

Supplementary Table 1

species	tissue	Sequencing depth	Sequencing depth (>25nt)	R²	Complexity
human	brain	8303	3238	0.76	0.638
human	cerebellum	8098	1605	0.62	0.635
human	heart	8123	514	0.33	0.498
human	kidney	8644	1926	0.4	0.712
mouse	brain	11154	1196	0.52	0.712
mouse	cerebellum	11664	904	0.53	0.660
mouse	heart	14818	1130	0.69	0.707
mouse	kidney	9788	706	0.48	0.668
mouse	testis	11572	8518	0.41	0.780
chicken	brain	20838	448	0.56	0.747
chicken	cerebellum	19313	388	0.45	0.773
chicken	heart	28251	2696	0.71	0.756
chicken	testis	27806	15281	0.29	0.846

Sequencing depth is measured as the number of sequenced reads per megabase, considering the size of the reference genome used. R² is the coefficient of determination from the regression analysis comparing the expression levels of coding genes retrieved from small and total RNA sequencing libraries. Complexity refers to

the Normalised Shannon entropy, defined as $\frac{\sum_i p_i \log_2 p_i}{\log_2(N)}$ where p_i is the proportion of reads of type i and N

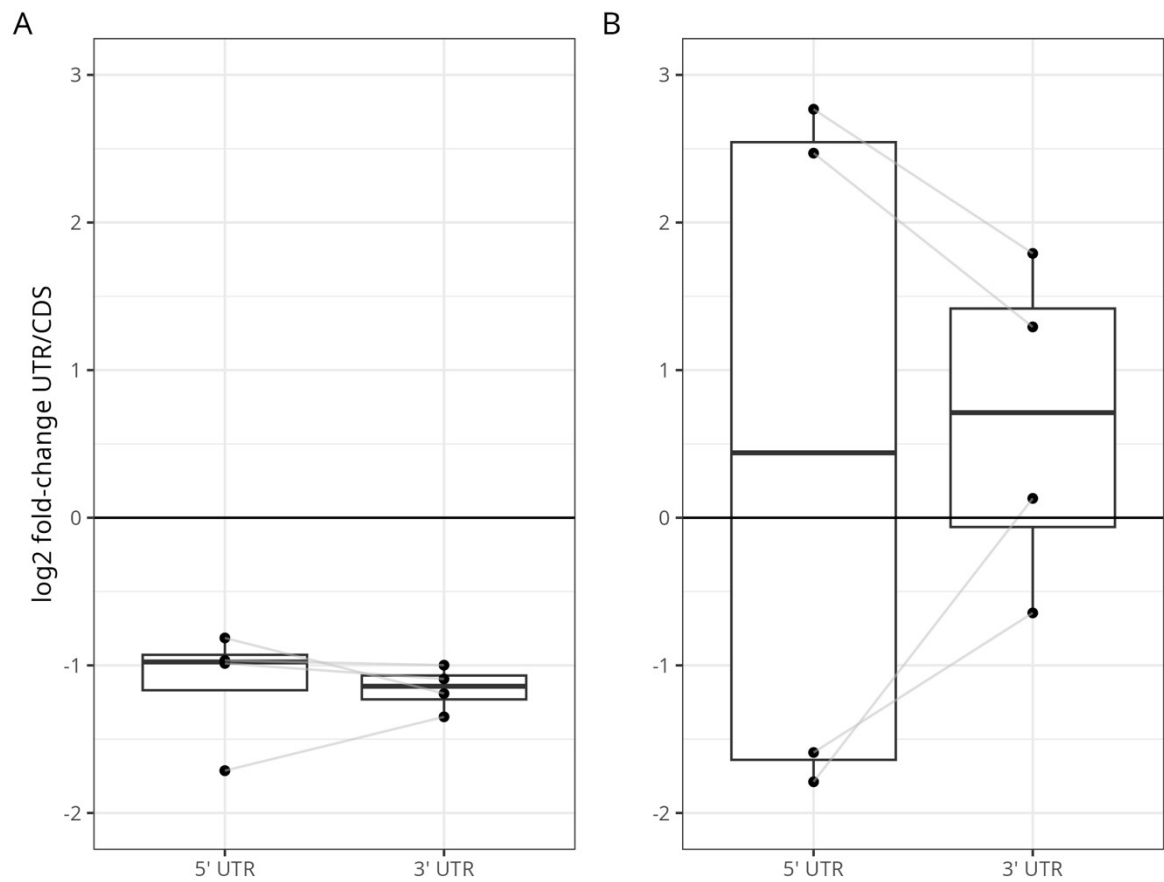
is the total number of unique reads.

Supplementary Table 2

Term (linear model)				
Tissue	Statistics	Total RNA	Half-life	Interaction
Brain	Slope	0.659	-0.017	-0.001
	SE	0.007	0.004	0.001
	p	0.000	0.000	0.452
Cerebellum	Slope	0.446	-0.011	0.001
	SE	0.006	0.003	0.001
	p	0.000	0.001	0.098
Heart	Slope	0.246	-0.018	-0.001
	SE	0.006	0.003	0.001
	p	0.000	0.000	0.375
Kidney	Slope	0.273	-0.019	0.002
	SE	0.007	0.004	0.001
	p	0.000	0.000	0.010

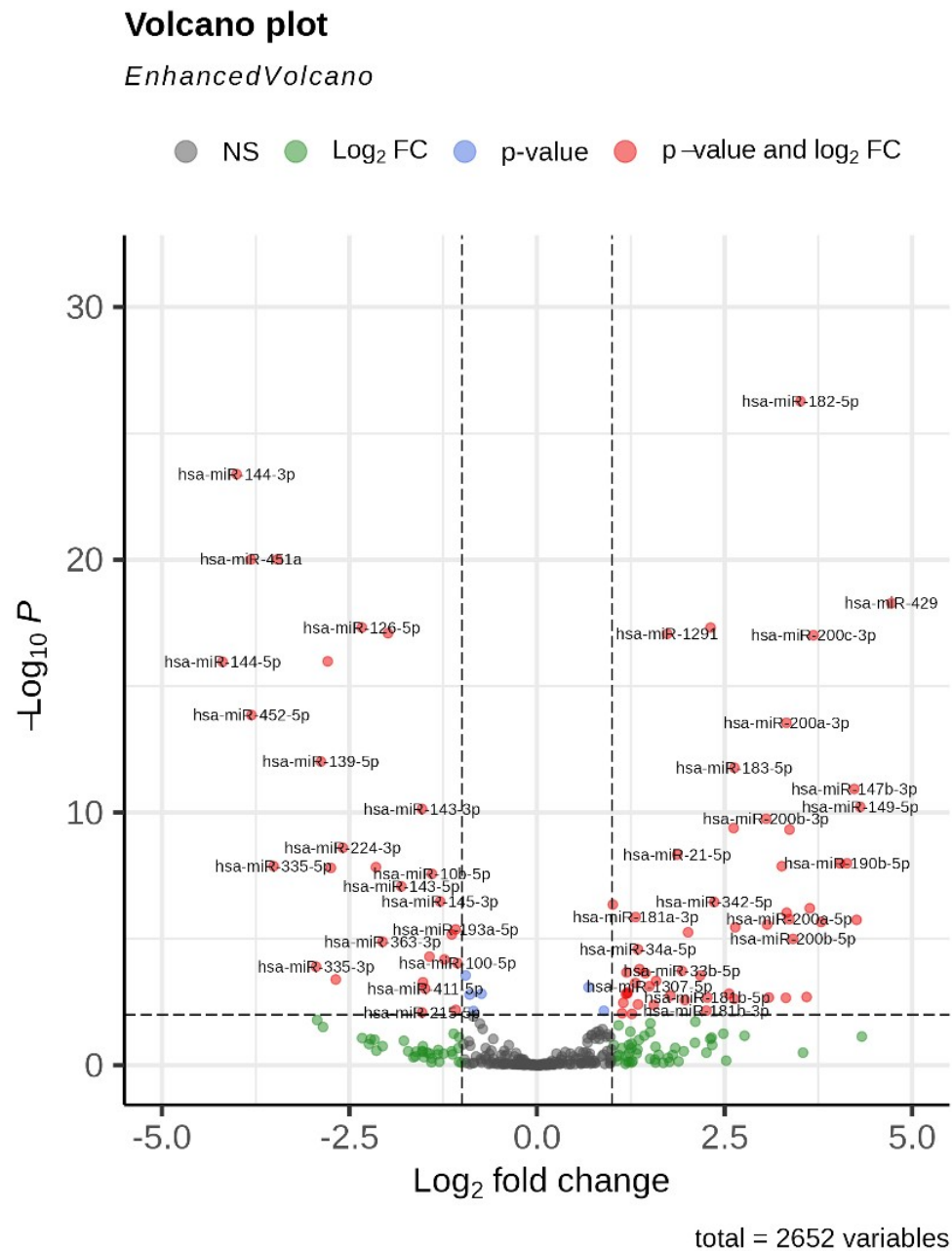
Statistical analysis of the linear model fitting the inferred gene expression levels from small RNA libraries against the levels from total RNA libraries and the half-life of the analysed transcripts.

Supplementary Figure 1



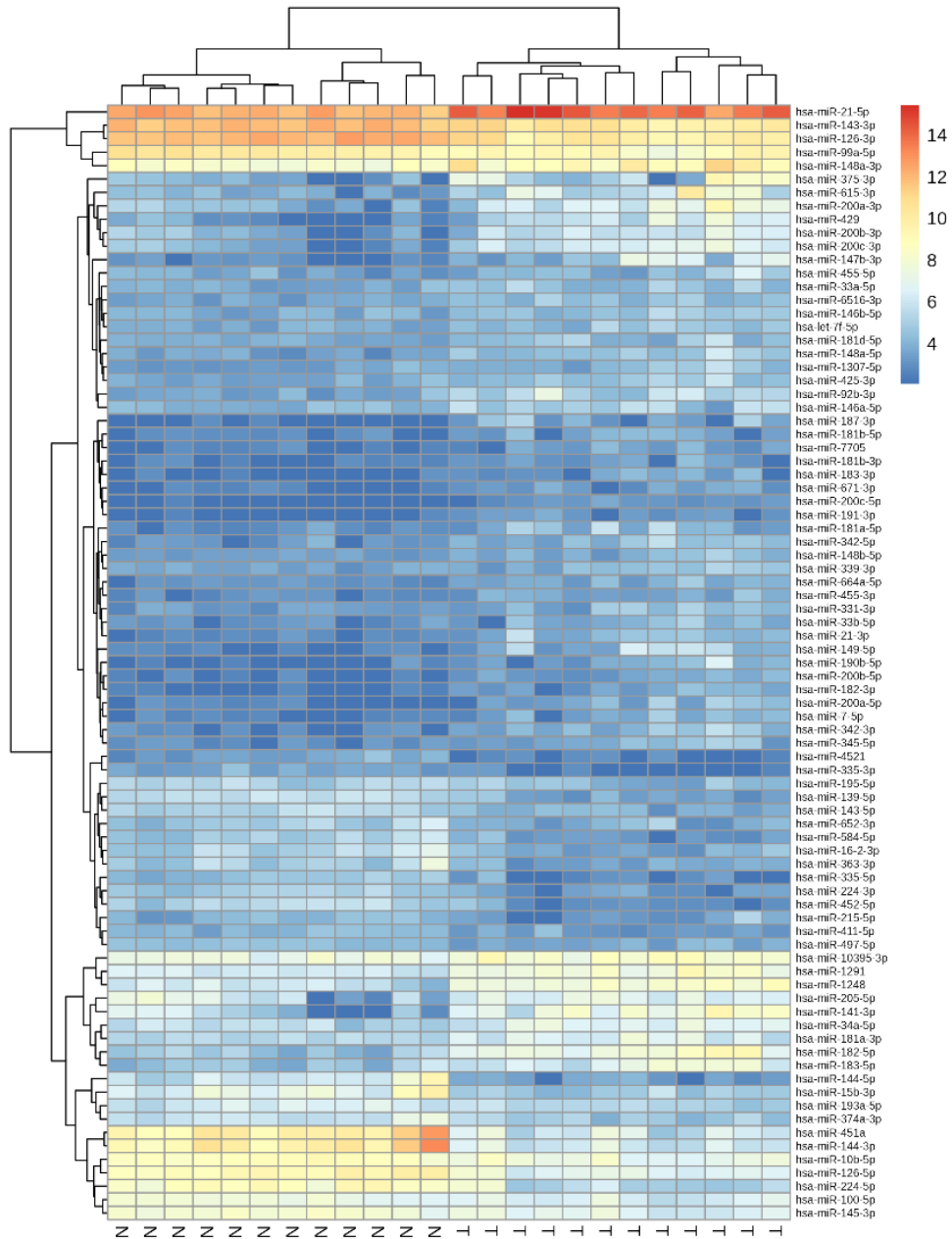
Supplementary Figure 1. Relative mapping of sequencing reads to untranslated regions. Log2 fold-change of the number of reads mapped to either the 5' untranslated region (5' UTR) or the 3' UTR with respect to reads mapped to the coding sequence. Values over 1 show an enrichment whilst value under 0 indicate a paucity of reads mapped to the UTR. Grey lines connect identical datasets (A) UTR fold-enrichment for reads from total RNA libraries. (B) UTR fold-enrichment for reads from small RNA libraries.

Supplementary Figure 2



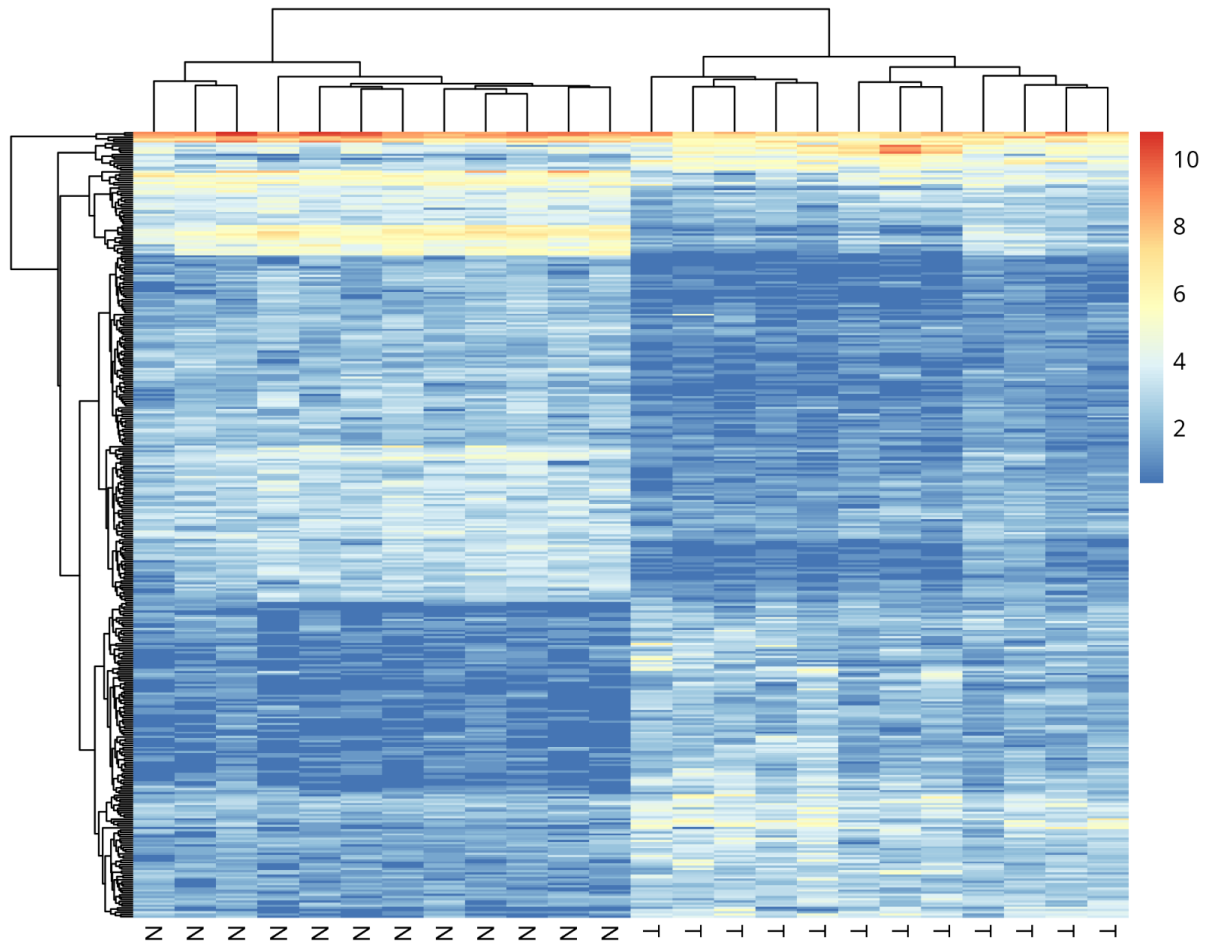
Supplementary Figure 2. Differential gene expression of microRNAs from breast cancer paired sRNAseq experiments. Volcano plot representing the expression fold change (DESeq2) in the x-axis of paired breast cancer samples (see main text) against the $-\log_{10}$ of the q value (FDR corrected p value) generated during the differential gene expression analysis. Identified differentially expressed genes are shown in red.

Supplementary Figure 3



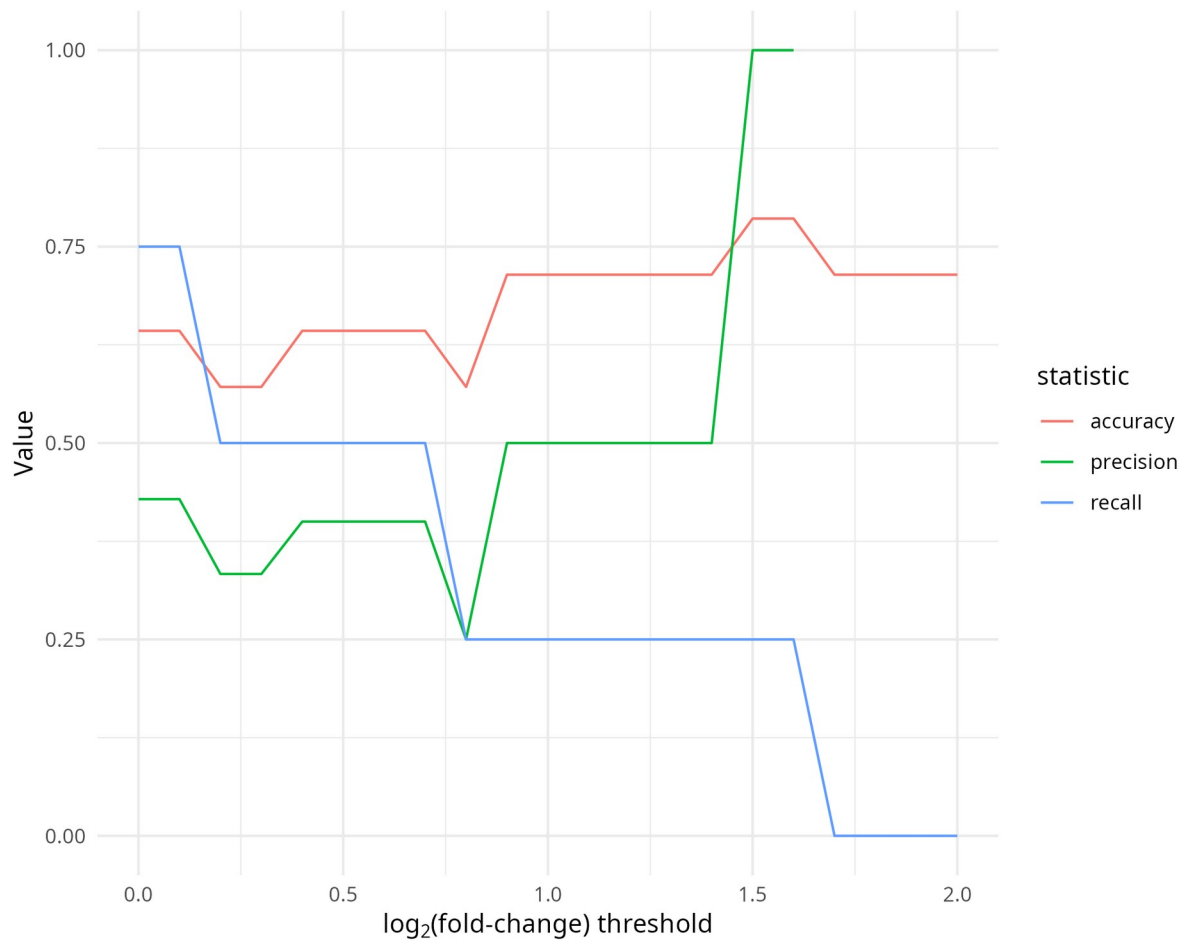
Supplementary Figure 3. Gene expression profile of microRNAs from breast cancer. Heatmap showing the expression levels of microRNAs from small RNA sequencing experiments in paired breast cancer samples.

Supplementary Figure 4



Supplementary Figure 4. Gene expression profile of protein-coding transcripts from breast cancer. Heatmap showing the expression levels of protein-coding genes from small RNA sequencing experiments in paired breast cancer samples.

Supplementary Figure 5



Supplementary Figure 5. Accuracy, precision and recall when comparing the DGE analysis with the gold standard. For thresholds of $\log_2(\text{fold-changes})$ in gene expression of the Differential Gene Expression analysis of Breast Cancer samples, the resulting upregulated genes were compared with the gold standard generated by Meerson et al. (2019). Values shown for accuracy, precision and recall.