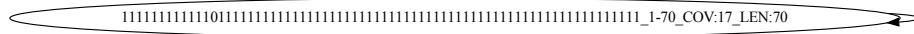
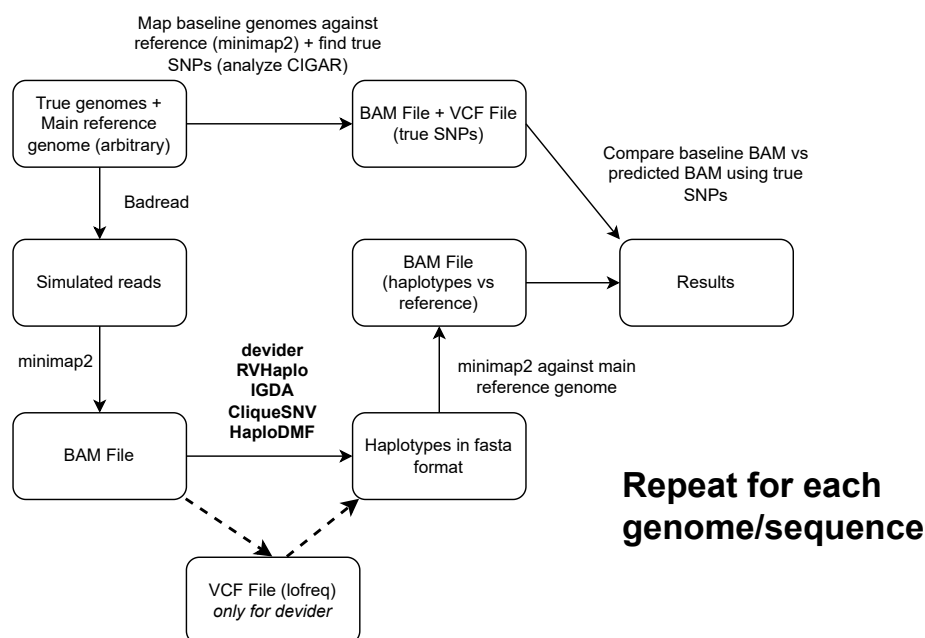


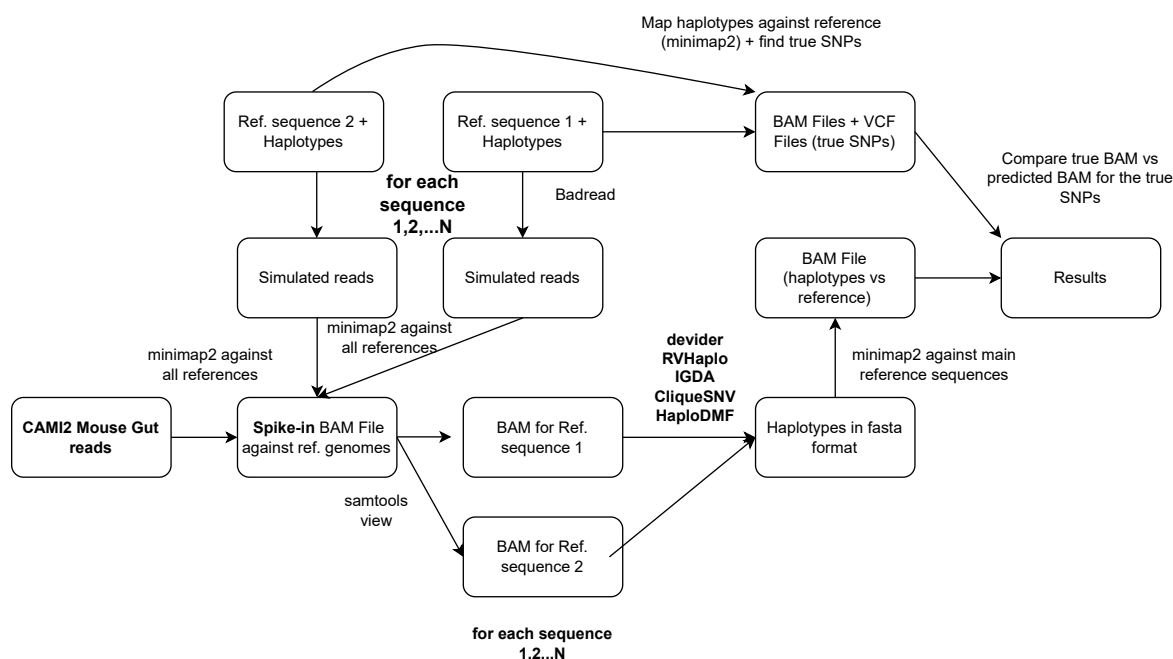
**Supplementary Figure 1.** Unitig graph for long reads (95% accuracy) for a single HIV-1 genome. The k-mer coverage cutoff was 2. The format is (alleles)\_(start-end)\_COV:(coverage)\_LEN:(number of alleles spanned). Notice that low coverage nodes and nodes with deletions (i.e.,  $\text{end} - \text{start} + 1 \neq \text{length}$ ) are more likely to be erroneous.



**Supplementary Figure 2.** Cleaned unitig graph for long reads (95% accuracy) for a single HIV-1 genome. The graph contains one unitig, as expected. The format is (alleles)\_(start-end)\_COV:(coverage)\_LEN:(number of alleles spanned).



**Supplementary Figure 3.** Benchmarking flowchart for the HIV-1 and antimicrobial resistance gene synthetic benchmarks.



**Supplementary Figure 4.** Benchmarking flowchart for the antimicrobial resistance gene **spike-in** synthetic benchmarks. The AMR gene reads were spiked into a mouse gut metagenome, and BAM files were generated for each AMR gene of interest with **samtools view** prior to haplotyping.

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Genome accessions for HIV communities

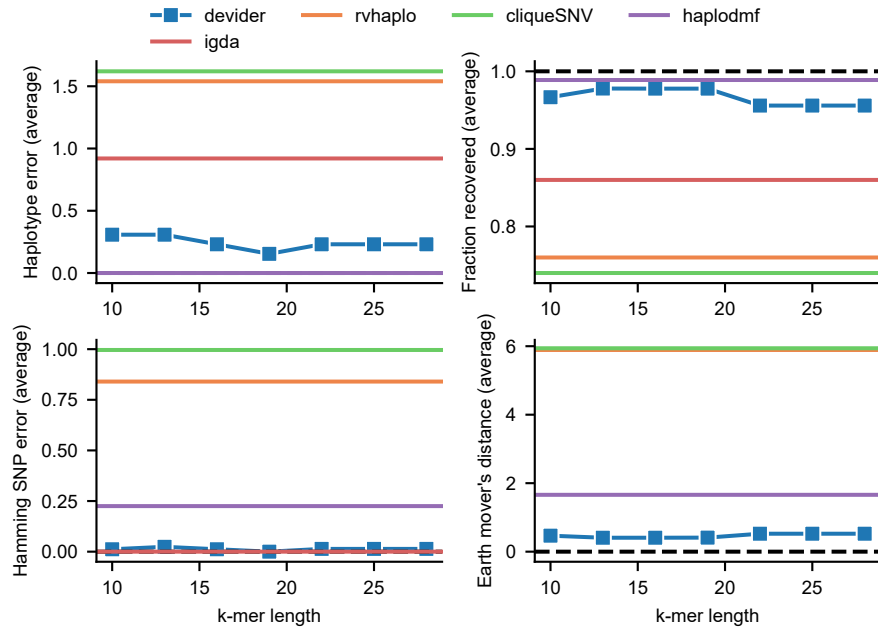
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- OR483999  
 - OR483988  
 - OR483990  
 - OR483987  
 - OR484006  
 - OR484007  
 - OR484012  

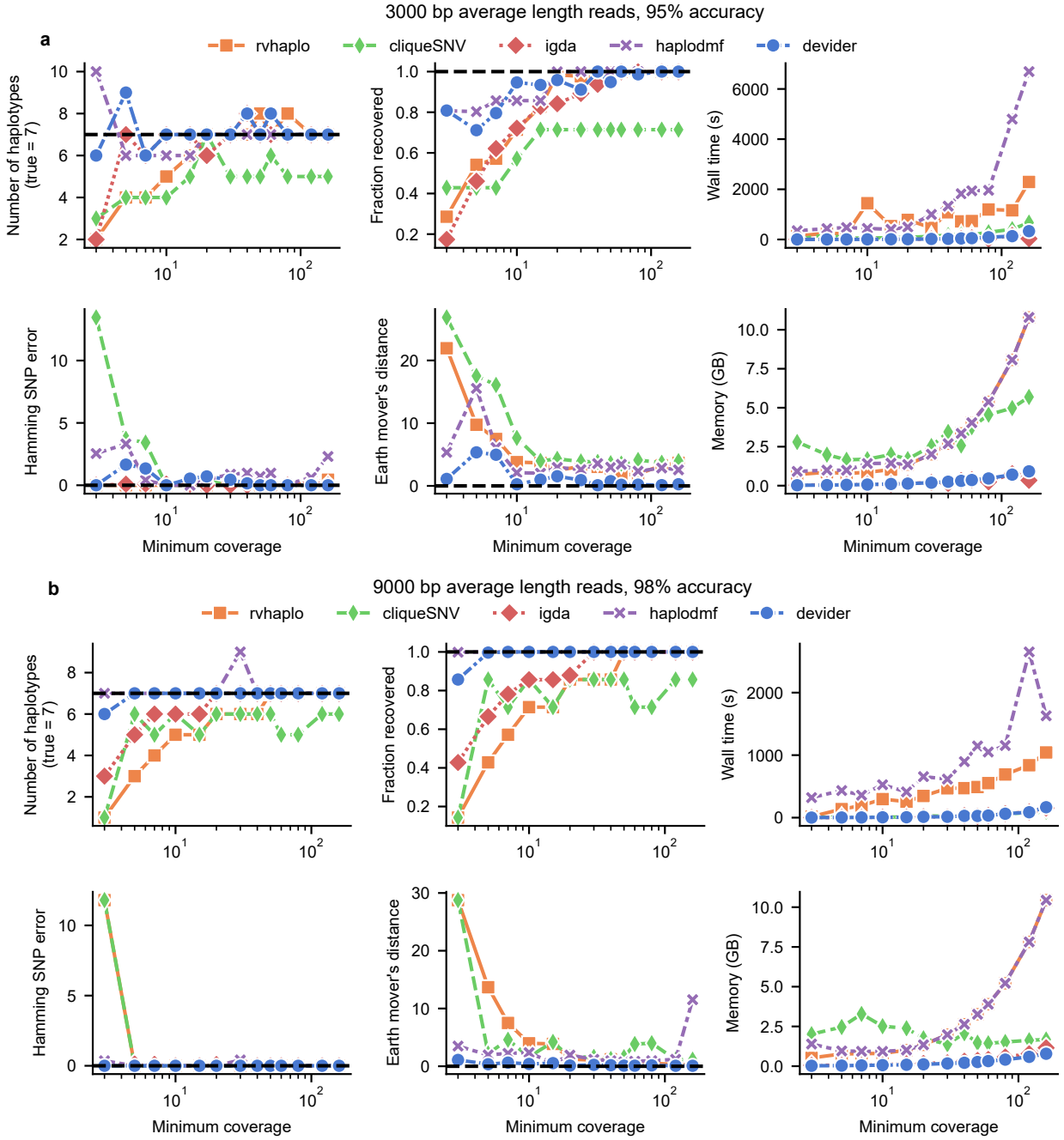

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 - OR484013  
 - OR484014  
 - OR484015  
 - OR484016  
 - OR483993  
 - OR483994  
 - OR483989  
 - OR483992  
 - OR483997  
 - OR484000  
 - OR484002  
 - OR483986  
 - OR484017  
 - OR484018  
 - OR484019  
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 - OR484022  
 - OR484001  
 - OR483985  
 - OR484004  
 - OR484005  
 - OR483996

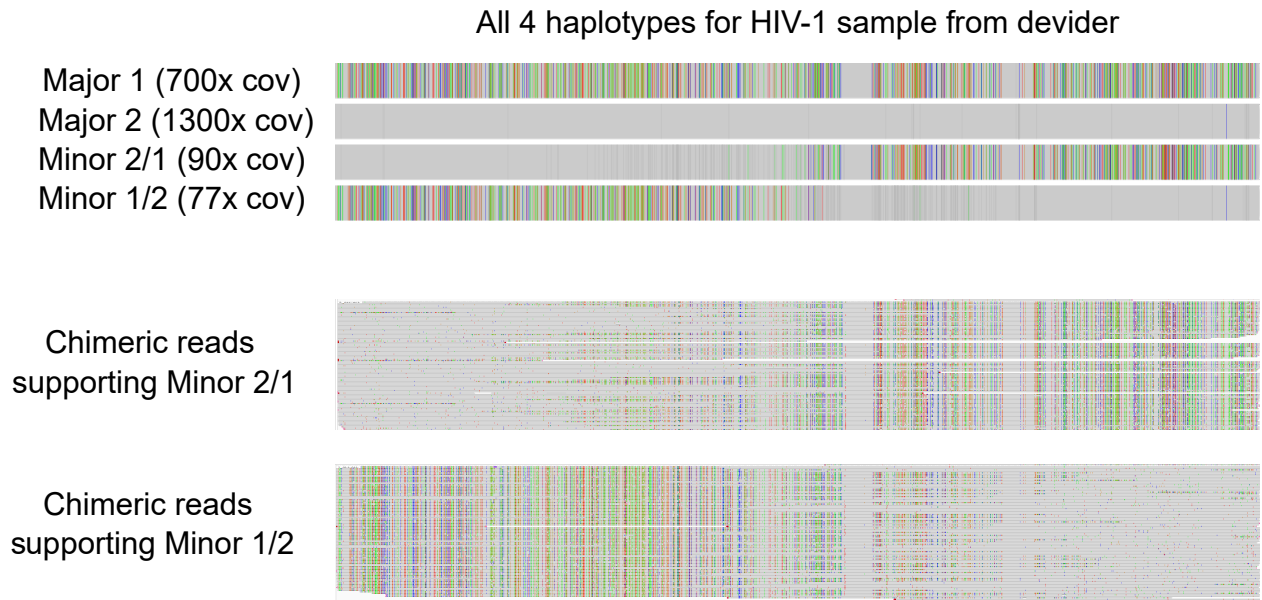
**Supplementary Table 1.** Genomes used for the HIV community. The first 7 (delimited by an horizontal line) are used for the 7 strain community and all 30 are used when benchmarking communities with 2 - 30 strains.



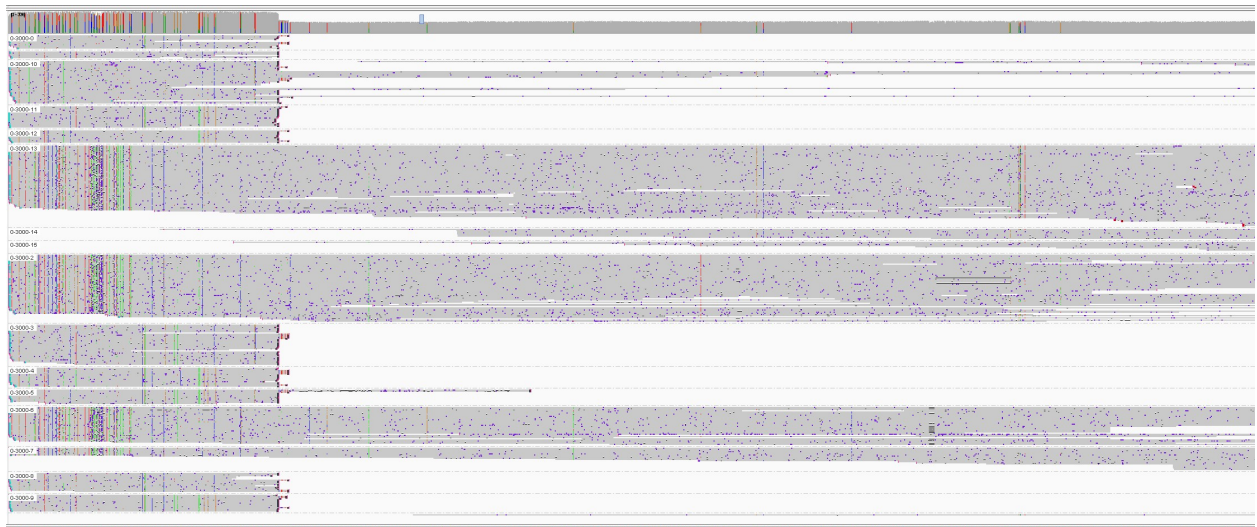
**Supplementary Figure 5.** devider's results over varying values of  $k$ -mer length  $k$  on the 7-strain HIV community with 9000 bp length and 95% accuracy simulated reads. devider dynamically selects  $k$  based on the data by default; instead, we fixed  $k$  from 10 to 30 and show averaged results over all data points. The other three methods are displayed as constant horizontal lines. The dashed black line shows the optimal values.



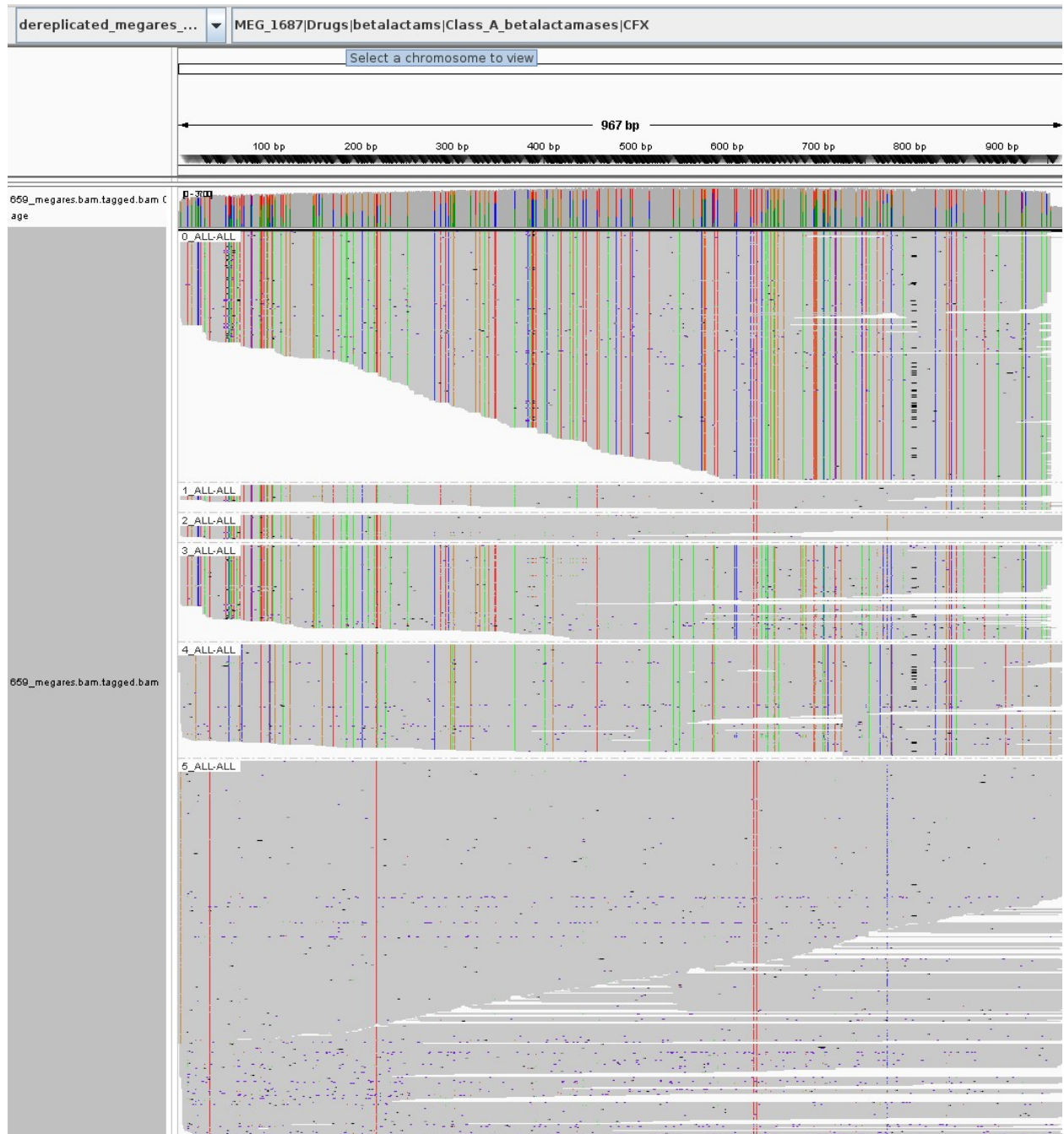
**Supplementary Figure 6.** The same experiment as Fig. 2a in the main paper on the 7-strain HIV synthetic community with staggered abundances. **a.** simulated reads have 3000bp average length and 95% accuracy. **b.** simulated reads have 9000 bp average length and 98% accuracy.



**Supplementary Figure 7.** All four haplotypes found by devider on the HIV-1 co-infection dataset. Two major haplotypes were found and two minor haplotypes were also found. Gray bars indicate alleles with < 66% of the reads supporting that allele. For the “chimeric” minor haplotypes, supporting reads assigned by devider are shown. The reference sequence for all alignments is a consensus generated by abPOA [1] from Major Haplotype 2’s reads.

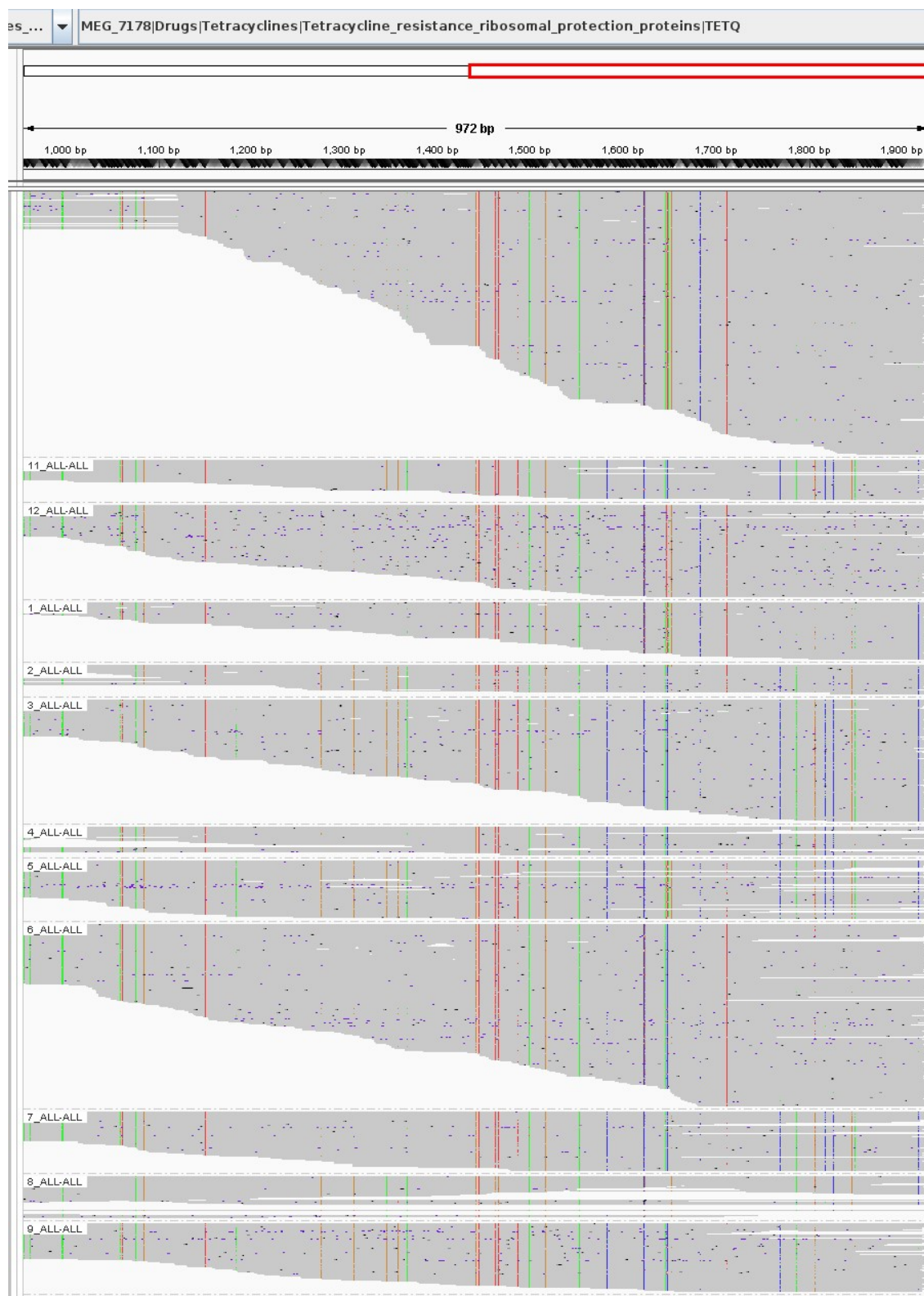


**Supplementary Figure 8.** A region with 16 estimated haplotypes by devider for the mussel-bacterial symbiont community. An elevated region of high coverage at the start of the contig (mean > 300x coverage vs estimated 220x single-copy coverage) and supplementary mapping reads suggest a duplicated region, which makes the number of haplotypes estimated higher.



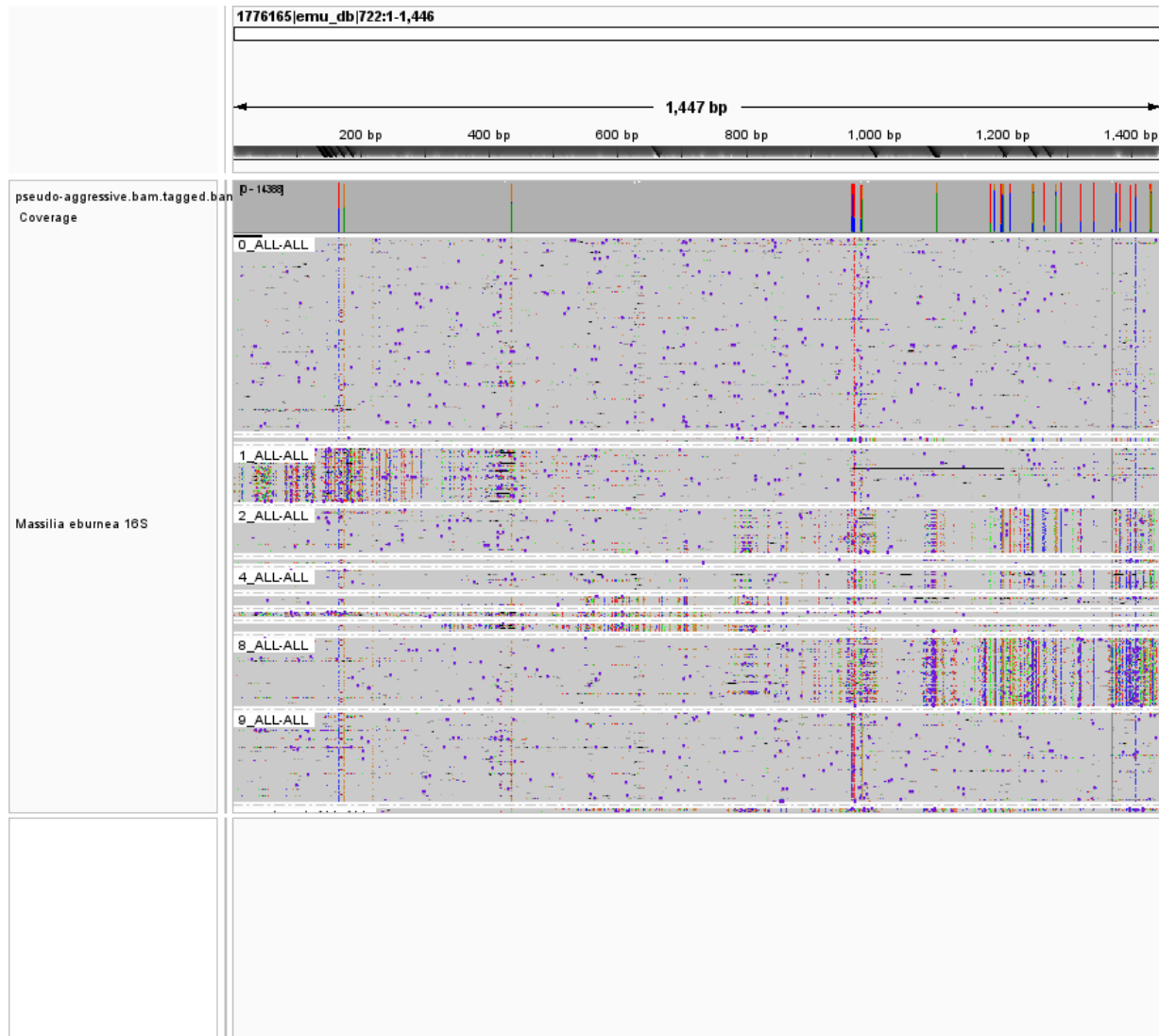
**Supplementary Figure 9.** Reads supporting the CfxA2 haplotypes shown in Fig. 4 c. Reads were downsampled to 1000x coverage per 1000 bp window for visualization purposes.



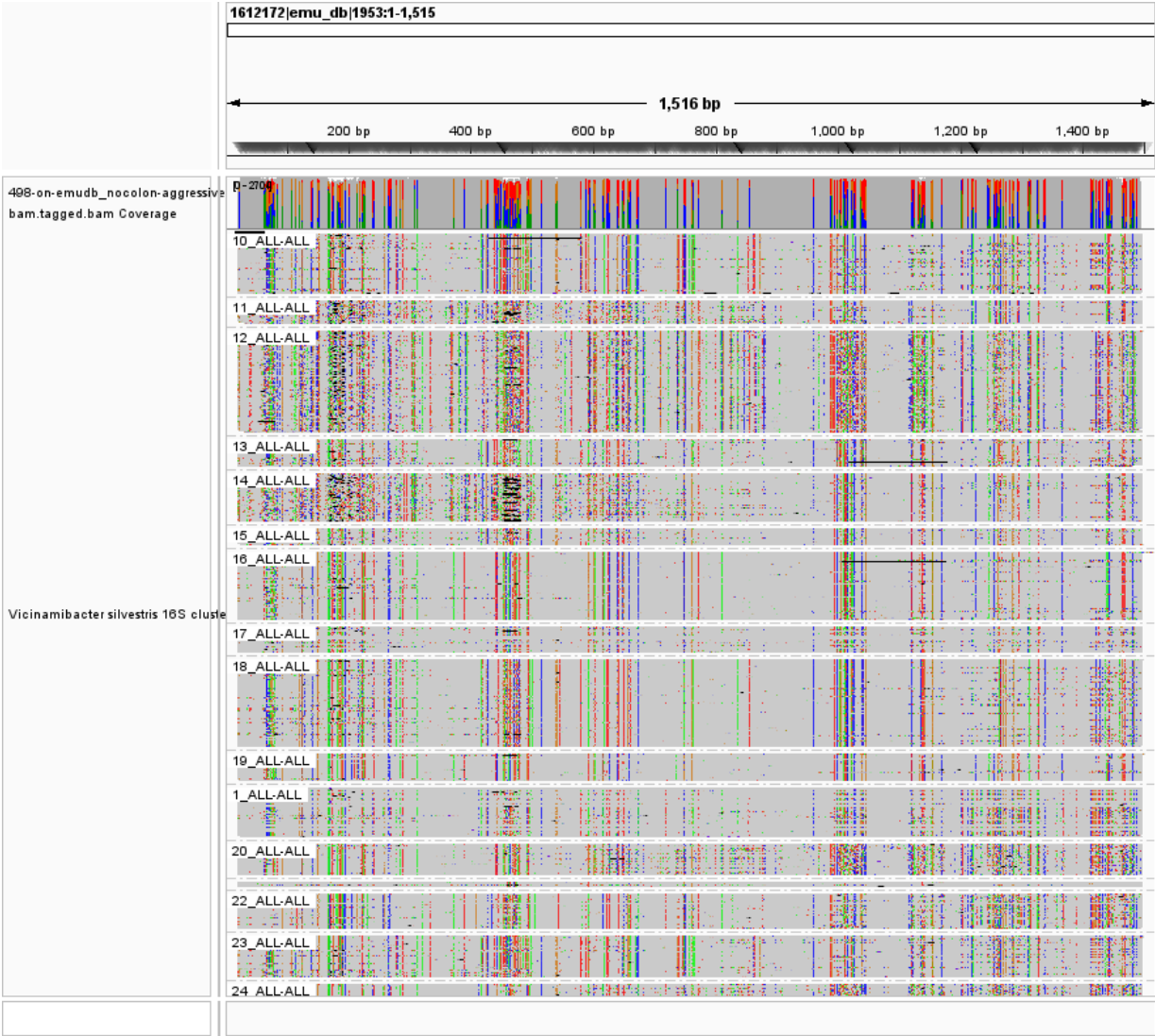


**Supplementary Figure 10.** Reads supporting a subset of the tet(Q) haplotypes shown in Fig. 4 c. Reads were downsampled to 1000x coverage per 1000 bp window for visualization purposes. Only the last 972 bp of the alignments are shown; Fig. 4 c also shows only the last 972 bp.





**Supplementary Figure 11.** Reads supporting the 16S consensus sequences mapping to *M. eburnea* for the 16S R10.4 ONT sequencing dataset. Reads were downsampled to 500x coverage for visualization purposes.



**Supplementary Figure 12.** Reads supporting a subset of 16S consensus sequences mapping to *V. silvestris* for the 16S R10.4 ONT sequencing dataset. Reads were downsampled to 500x coverage for visualization purposes.

## References

1. Gao, Y. *et al.* abPOA: An SIMD-based C library for fast partial order alignment using adaptive band. *Bioinformatics* **37**, 2209–2211 (2021).