

SUPPLEMENTAL FIGURES

Rapid and accurate demultiplexing of direct RNA nanopore sequencing datasets with SeqTagger

Leszek P Pryszzcz^{1,2,3,*}, Gregor Diensthuber^{1,2,3,4}, Laia Llovera¹, Rebeca Medina¹, Anna Delgado-Tejedor^{1,2}, Luca Cozzuto¹, Julia Ponomarenko¹ and Eva Maria Novoa^{1,2,3,*}

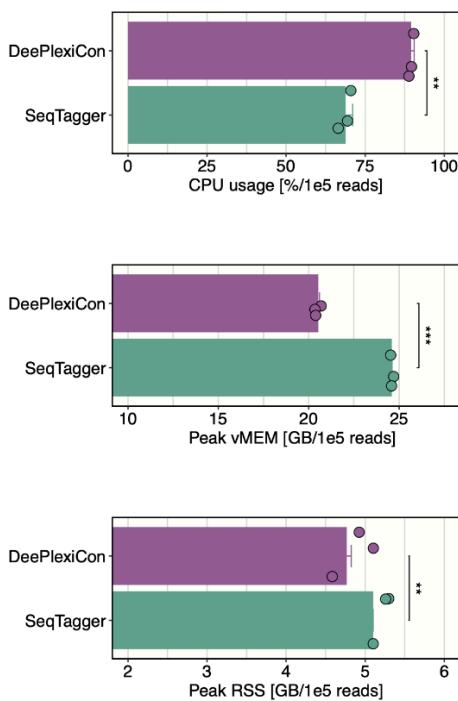
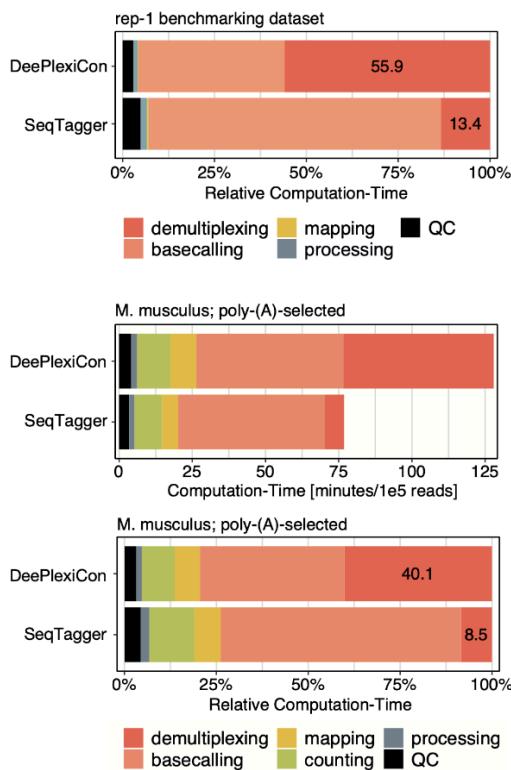
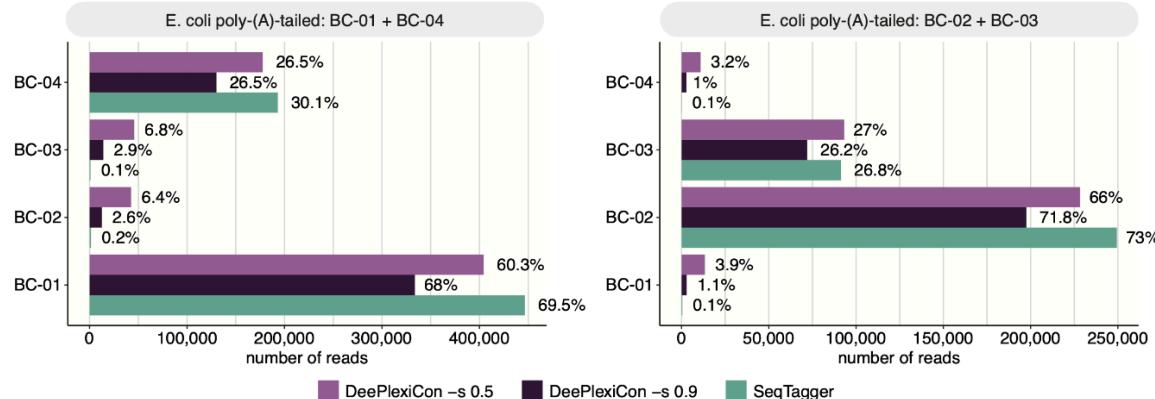
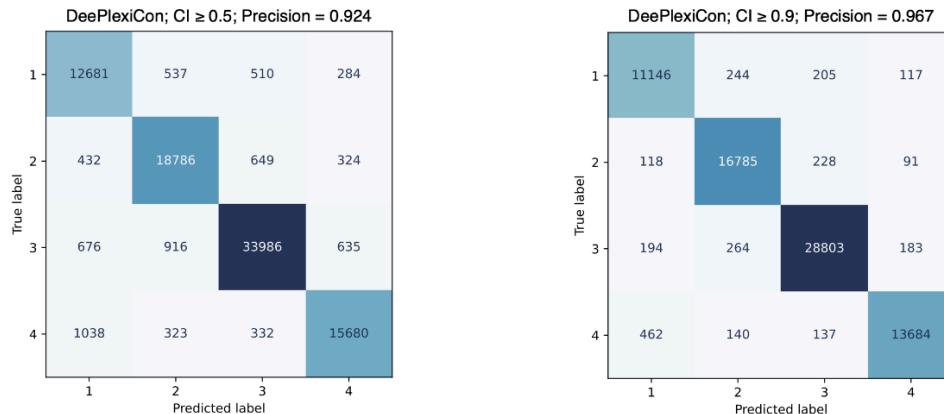
¹*Centre for Genomic Regulation (CRG), The Barcelona Institute of Science and Technology, Dr. Aiguader 88, Barcelona 08003, Spain*

²*Universitat Pompeu Fabra (UPF), Barcelona, Spain*

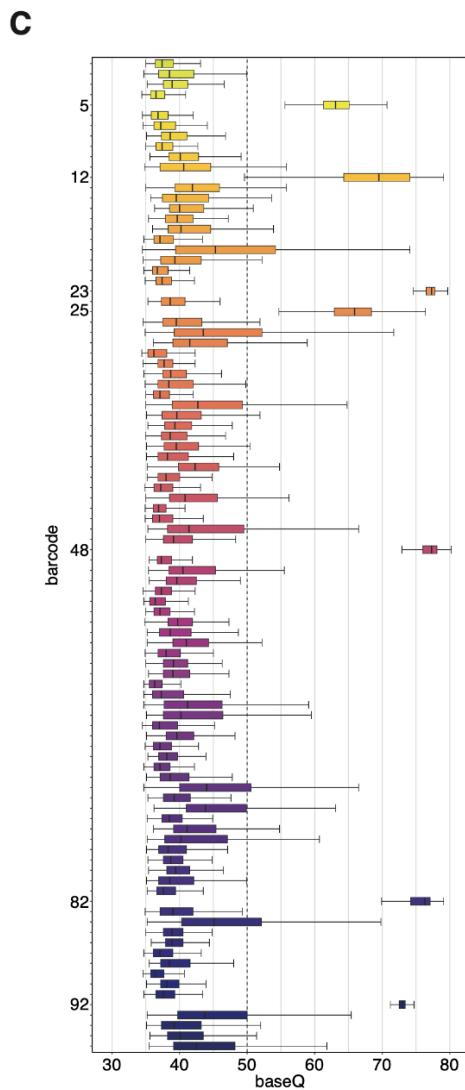
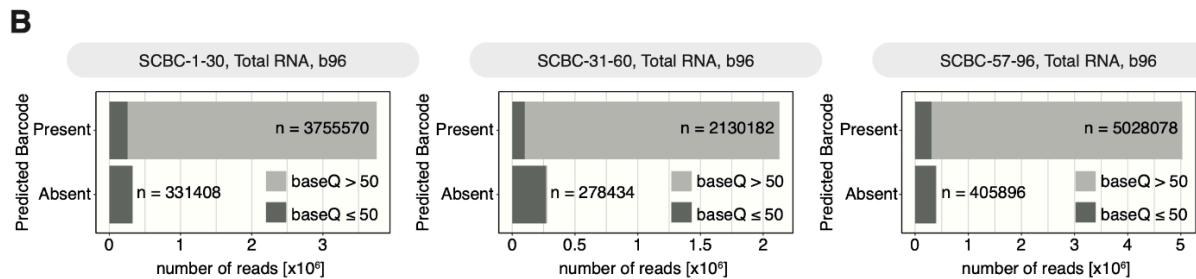
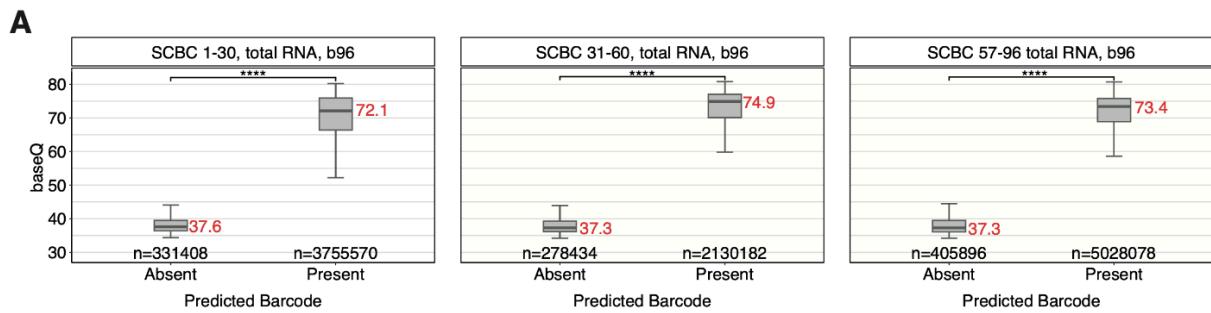
³*ICREA, Pg. Lluís Companys 23, Barcelona, España*

[#] *These authors contributed equally*

* Correspondence to: Leszek P Pryszzcz (leszek.pryszzcz@crg.eu) and Eva Maria Novoa (eva.novoa@crg.eu)

A**B****D****E**

Supplemental Figure S1. Benchmarking results for computational requirements and model performance **(A)** Barplots depicting system requirements (CPU usage; peak vMEM; peak RSS) of DeePlexiCon and SeqTagger on the benchmarking dataset. Dots represent individual replicates with bars representing the mean value and error bars depicting +/-1 standard deviation. To determine statistical significance, a two-sided t-test was performed and results were corrected for multiple hypothesis testing using the Benjamini-Hochberg procedure. **(B)** Barplot depicting the relative contributions of each preprocessing workflow to the overall computation time on rep-1 of the benchmarking dataset. **(C)** Barplots depicting the absolute and relative contributions to the overall computation time of 100,000 reads sampled from a mouse poly(A)-selected sample aligned to the mm39 genome. **(D)** Barplots representing the percentage of reads assigned to each barcode for two runs of total RNA from *E.coli* (poly(A)-tailed). The first run (left) contained BC-01 and BC-04 while the second run (right) contained barcodes BC-02 and BC-03. Runs were demultiplexed with either SeqTagger (b04_RNA002) or DeePlexiCon (resnet20-final.h5) with high recovery (-s 0.5) or high accuracy (-s 0.9) settings (see *Methods*). **(E)** Confusion matrices corresponding to DeePlexiCon results for high recall (-s 0.5) and high precision (-s 0.9) on rep-1 of the benchmarking dataset.



Supplemental Figure S2. Performance of SeqTagger's 96 barcode model on independent test data. **(A)** Boxplots showing the baseQ distribution for barcodes present and absent in the three independent test runs. The Number of reads is indicated by n with the median value shown in red. Statistical analysis was performed using a two-sided non-parametric Wilcoxon test. Results were corrected for multiple-hypothesis testing using the Bonferroni procedure to obtain adjusted p-values (ns: $p > 0.05$, *: $p \leq 0.05$, **: $p \leq 0.01$, ***: $p \leq 0.001$, ****: $p \leq 0.0001$). **(B)** Barplots representing the total number of reads (n) for three independent test runs demultiplexed with SeqTagger's 96 barcode model (b96_RNA002). Colors indicate different baseQ thresholds. **(C)** Boxplots of base quality (baseQ) per barcode for an additional independent test run containing SCBC-05, SCBC-12, SCBC-23, SCBC-25, SCBC-48, SCBC-82 and SCBC-96. For Figures S2A and S2C the box is limited by the lower quartile Q1 (bottom) and upper quartile Q3 (top). Whiskers are defined as $1.5 * \text{IQR}$ with outliers not shown for visualization purposes.
