

Supplemental material to:

Optical genome mapping enables accurate testing of large repeat expansions

Bart van der Sanden¹, Kornelia Neveling¹, Syukri Shukor², Michael D. Gallagher², Joyce Lee², Stephanie L. Burke², Maartje Pennings¹, Ronald van Beek¹, Michiel Oorsprong¹, Ellen Kater-Baats¹, Eveline Kamping¹, Alide A. Tieleman³, Nicol C. Voermans³, Ingrid E. Scheffer⁴, Jozef Gecz^{5, 6, 7}, Mark A. Corbett⁷, Lisenka E.L.M. Vissers¹, Andy Wing Chun Pang², Alex Hastie², Erik-Jan Kamsteeg^{1,*}, Alexander Hoischen^{1,8,*}

1 Department of Human Genetics, Research Institute for Medical Innovation, Radboud University Medical Center, Nijmegen, the Netherlands

2 Bionano Genomics Clinical and Scientific Affairs, San Diego, CA, USA

3 Department of Neurology, Radboud university medical center, Nijmegen, the Netherlands

4 Epilepsy Research Centre, Department of Medicine, University of Melbourne, Austin Health, VIC, Australia; Department of Pediatrics, University of Melbourne, Royal Children's Hospital, Florey and Murdoch Children's Research Institutes, VIC, Melbourne, Australia.

5 South Australian Health and Medical Research Institute, Adelaide, SA, Australia

6 Genetics and Molecular Pathology, SA Pathology, Adelaide, SA, Australia

7 Robinson Research Institute and Adelaide Medical School, University of Adelaide, Adelaide South Australia, Australia

8 Department of Internal Medicine, Radboud Expertise Center for Immunodeficiency and Autoinflammation and Radboud Center for Infectious Disease (RCI), Radboud university medical center, Nijmegen, the Netherlands

* These authors jointly supervised the work

Corresponding Authors

Erik-Jan Kamsteeg, PhD

Department of Human Genetics

Radboud university medical center

6500HB Nijmegen, The Netherlands

Email: erik-jan.kamsteeg@radboudumc.nl

AND

Alexander Hoischen, PhD

Department of Human Genetics & Department of Internal Medicine

Radboud university medical center

6500HB Nijmegen, The Netherlands

Email: alexander.hoischen@radboudumc.nl

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Supplemental Table S1: Deviation between the manual *de novo* assembly workflow and the local guided assembly workflow. For each data point in Fig 2, this table presents the estimated size of the largest allele resulting from both workflows, as well as the deviation between the two sizing workflows. Deviation is calculated as: $((\text{local}-\text{manual})/\text{local}) \times 100$. For this analysis the deviation was only calculated one way.

Sample	Manual de novo assembly	Local guided assembly	Deviation (%) [#]
CNBP_02	6331	6544	3.4
CNBP_03	6517	5374	-17.5
CNBP_04	7155	5652	-21.0
CNBP_05	8042	6322	-21.4
CNBP_07	5212	5173	-0.7
CNBP_08	2502	2661	6.3
CNBP_10	2874	3980	38.5
CNBP_11	375	254	-32.2
CNBP_12	3471	3346	-3.6
CNBP_13	4634	4330	-6.6
CNBP_14	5244	4331	-17.4
CNBP_15	2183	2092	-4.2
CNBP_16	3221	4747	47.4
CNBP_17	6275	5759	-8.2
CNBP_18	1915	2976	55.4
CNBP_19	1460	1574	7.8
CNBP_20	3977	4852	22.0
CNBP_21	288	244	-15.4
CNBP_22	1683	1725	2.5
CNBP_23	2131	2515	18.0
CNBP_25	1476	2626	77.9
CNBP_26	3737	3241	-13.3

Sample	Manual de novo assembly	Local guided assembly	Deviation (%) [#]
DMPK_01	269	247	-8.1
DMPK_02	456	473	3.8
DMPK_03	252	116	-54.0
DMPK_04	66	60	-9.1
DMPK_05	457	485	6.1
DMPK_06	64	82	28.1
DMPK_07	378	358	-5.2
DMPK_09	71	68	-4.2
DMPK_10	2829	2825	-0.1
DMPK_11	233	231	-1.0
DMPK_12	213	219	3.0
DMPK_13	163	167	2.7
DMPK_14	202	188	-6.9
DMPK_15	1839	1768	-3.9
DMPK_16	85	52	-38.6

DMPK_17	491	510	3.8
DMPK_18	71	69	-2.4
DMPK_19	54	61	13.7
DMPK_21	1366	1347	-1.4
DMPK_23	372	369	-0.9
DMPK_24	82	93	13.4
DMPK_25	79	109	38.0
DMPK_28	440	393	-10.6
DMPK_29	320	310	-3.1
DMPK_30	290	131	-54.8

Sample	Manual de novo assembly	Local guided assembly	Deviation (%) [#]
RFC1_01	1487	1497	0.6
RFC1_02	738	751	1.8
RFC1_03	883	897	1.6
RFC1_04	750	760	1.3
RFC1_05	1167	1175	0.7
RFC1_06	1565	1579	0.9
RFC1_07	625	643	2.9
RFC1_08	840	856	1.9
RFC1_09	1506	1499	-0.5
RFC1_10	1289	1307	1.4
RFC1_11	812	818	0.7
RFC1_12	1106	1121	1.4
RFC1_13	927	933	0.6
RFC1_14	725	740	2.0
RFC1_15	873	887	1.6
RFC1_16	1104	1097	-0.6
RFC1_17	711	723	1.6
RFC1_18	1444	1474	2.1
RFC1_19	223	235	5.6
RFC1_20	494	502	1.6
RFC1_21	703	715	1.6
RFC1_22	1161	1180	1.6
RFC1_23	1028	1054	2.5
RFC1_24	602	615	2.1
RFC1_25	855	868	1.6
RFC1_26	714	734	2.8
RFC1_27	973	987	1.5
RFC1_28	875	893	2.0
RFC1_29	1071	1073	0.2
RFC1_30	743	754	1.5

[#] for calculating the average deviations in the manuscript, all deviations presented in this table were taken as positive value.

Supplemental Table S2: Repeat expansion sizes in repeat units at the three different repeat loci in ten control samples based on the manual *de novo* workflow. OGM only identified repeat sizes below the pathogenic repeat sizing threshold of *CNBP* and *DMPK*. In the case of *RFC1*, OGM identified three heterozygous and one homozygous expansion beyond the pathogenic repeat size threshold of 20 units that was used in this manuscript. However, the 20 units we used is different from the formal pathogenic repeat size threshold of *RFC1* at >400 units (**Methods** and **Table 3**). None of the calls exceeded this formal threshold and therefore the four samples are considered carriers. This data suggests that OGM has a very low false positive rate for the detection of repeat expansions.

	<i>CNBP</i>		<i>DMPK</i>		<i>RFC1</i>	
Control #	Allele 1	Allele 2	Allele 1	Allele 2	Allele 1	Allele 2
1	3	3	23	23	-3	-3
2	6	-39	30	30	59	13
3	17	17	14	14	-3	-4
4	25	-53	15	15	117	117
5	-37	-37	-17	-17	-1	-1
6	-26	-26	35	35	71	24
7	55	55	-11	-11	0	0
8	-5	-5	13	13	-4	-4
9	-42	-42	21	21	106	-1
10	-17	-17	-31	-46	14	14

Supplemental Table S3: The OGM results of the 12 additional samples with a known repeat expansion in *ATXN10*, *C9orf72*, *FXN*, *NOP56* or *STARD7*. For all samples except *ATXN10_03*, OGM detected a repeat expansion. In addition, OGM allowed to distinguish between the two repeat alleles for *FXN_02*. *ATXN10* (Morato Torres et al. 2022) and *C9orf72* (Barseghyan et al. 2022) repeat cases were also published before. For the molecule distance script “A” denotes multiple consensus maps and “B” denotes a gradient in the molecule distances. We now considered somatic instability in cases where both “A + B” provided suggestive evidence.

Sample	Repeat motif	Pathogenic repeat size threshold	SOC	Manual <i>de novo</i> assembly	Local guided assembly	Suggestive evidence of somatic instability from molecule distance script
ATXN10_01	ATTCT	>850	>850	-10 / 1,249	0 / 1,206	A
ATXN10_02	ATTCT	>850	>850	-11 / 1,076	0 / 1,056	A
ATXN10_03	ATTCT	>850	>850	-4 / 1,257	0 / 1,263	-
ATXN10_04	ATTCT	>850	>850	-10 / 1,249	2 / 2,711	A + B
C9orf72_01	GGCCCC	>250	>250	3 / 1,579	15 / 1,615	B
C9orf72_02	GGCCCC	>250	>250	7 / 2,314	16 / 16	A + B
FXN_01	GAA	>65	8 / >65	22 / 483	33 / 501	A
FXN_02	GAA	>65	>65 / >65	221 / 886	172 / 912	A
NOP56_01	GGCCTG	>670	5 / >40	16 / 1,081	19 / 1,077	A
NOP56_02	GGCCTG	>670	5 / >40	19 / 1,189	0 / 9	A + B
STARD7_01	ATTTC	>660	1,135*	-1 / 1,257	0 / 1,258	-

* SOC for *STARD7* was performed using long-range PCR followed by Oxford Nanopore sequencing.

Supplemental Table S4: Overview of which repeat expansions can be detected using our OGM analysis approach. The table contains all known repeat expansion loci published in (Depienne and Mandel 2021). “Yes” means the pathogenic repeat size threshold is >500 bp and repeats at this locus can be detected using our approach. “Possible” means the pathogenic repeat size threshold starts <300 bp, but extends beyond the 300 bp or even 500 bp mark. For these loci it depends on the actual size of the repeat expansion whether our approach works for these genes.

“No” means the pathogenic repeat size threshold remains <300 bp and repeat expansions at this locus cannot be detected using the OGM approach. For each gene the respective disorder, repeat unit, pathologic repeat size threshold is given. Table adjusted from (Depienne and Mandel 2021).

Gene	Disorder	Repeat motif	Pathologic repeat size threshold (units)	Pathologic repeat size threshold (base pairs)	OGM capable to detect pathogenic repeat expansion
<i>AFF2</i>	Fragile XE syndrome	CCG	≥200-900	≥600-2700	Yes
<i>AR</i>	Spinal and bulbar muscular atrophy	CAG	≥38-68	≥114-204	No
<i>ARX</i>	Early infantile epileptic encephalopathy type 1	GCN	23	69	No
<i>ATN1</i>	Dentatorubral-pallidoluysian atrophy	CAG	≥48-93	≥144-279	No
<i>ATXN1</i>	Spinocerebellar ataxia type 1	CAG	≥39-88	≥117-264	No
<i>ATXN10</i>	Spinocerebellar ataxia type 10	ATTCT/ATTGT	>280-4500	>1400-22500	Yes
<i>ATXN2</i>	Spinocerebellar ataxia type 2	CAG	≥32-500	≥96-1500	Possible
<i>ATXN3</i>	Spinocerebellar ataxia type 3	CAG	≥55-87	≥165-261	No
<i>ATXN7</i>	Spinocerebellar ataxia type 7	CAG	≥37-460	≥111-1380	Possible
<i>ATXN8/ATXN8OS</i>	Spinocerebellar ataxia type 8	CAG/CTG	>74-250	>222-750	Possible
<i>BEAN1/TK2</i>	Spinocerebellar ataxia type 31	TGGAA	≥110-760	≥550-3800	Yes
<i>C9ORF72</i>	Frontotemporal dementia / amyotrophic lateral sclerosis	GGGGCC	>30	>180	No
<i>CACNA1A</i>	Spinocerebellar ataxia type 6	CAG	≥20-33	≥60-99	No
<i>CNBP</i>	Myotonic dystrophy type 2	CCTG/CAGG	>50-11000	>200-44000	Possible
<i>CSTB</i>	Progressive myoclonus epilepsy type 1	CCCCGCCCGCG	30-75	360-900	Possible
<i>DAB1</i>	Spinocerebellar ataxia type 37	ATTTC	≥31-75	≥155-375	Possible
<i>DMPK</i>	Myotonic dystrophy type 1	CTG	>50-10000	>150-30000	Possible
<i>FMR1</i>	Fragile X syndrome	CGG	>200	>600	Yes
<i>FMR1</i>	Fragile X-associated premature ovarian infertility	CGG	55-200	165-600	Possible

<i>FMR1</i>	Fragile X-associated tremor ataxia syndrome	CGG	55-200	165-600	Possible
<i>FOXL2</i>	Blepharophimosis, ptosis and epicanthus inversus	GCN	19-24	57-72	No
<i>FXN</i>	Friedreich ataxia	GAA	≥66-1300	≥198-3900	Possible
<i>GIPC1</i>	Oculopharyngeal muscular dystrophy type 2	CGG	≥97-120	≥291-360	Possible
<i>GLS</i>	Global developmental delay, progressive ataxia, and elevated glutamine	GCA	≥680-1400	≥2040-4200	Yes
<i>HOXA13</i>	Hand-foot-genital syndrome	GCN	22	66	No
<i>HOXD13</i>	Synpolydactyly type 1	GCG	24	72	No
<i>HTT</i>	Huntington disease	CAG	≥36-250	≥108-750	Possible
<i>JPH3</i>	Huntington disease-like 2	CAG	≥41-58	≥123-174	No
<i>LRP12</i>	Oculopharyngodistal myopathy type 1	CGG	90-130	270-390	Possible
<i>MARCHF6</i>	Familial adult myoclonic epilepsy type 3	ATTTC	≥660-2800	≥3300-14000	Yes
<i>NOP56</i>	Spinocerebellar ataxia type 36	GGCCG	≥650-2500	≥3250-12500	Yes
<i>NOTCH2NLC</i>	Neuronal intranuclear inclusion disease	CGG	≥61-500	≥183-1500	Possible
<i>NUM2B-AS1</i>	Oculopharyngeal myopathy with leukoencephalopathy type 1	CGG/CCG	40-60	120-180	No
<i>PABPN1</i>	Oculopharyngeal muscular dystrophy	GCG	≥12-17	≥36-51	No
<i>PHOX2B</i>	Congenital central hypoventilation syndrome	GCN	25-29	75-87	No
<i>PPP2R2B</i>	Spinocerebellar ataxia type 12	CAG	43-78	129-234	No
<i>RAPGEF2</i>	Familial adult myoclonic epilepsy type 7	ATTTC	N/A	N/A	N/A
<i>RFC1</i>	Cerebellar ataxia, neuropathy and vestibular areflexia syndrome	AAGGG	≥400-2000	≥2000-10000	Yes
<i>RUNX2</i>	Brachydactyly and cleidocranial dysplasia	GCN	27	81	No
<i>SAMD12</i>	Familial adult myoclonic epilepsy type 1	ATTTC	≥440-3680	≥2200-18400	Yes
<i>SOX3</i>	Mental retardation with isolated growth hormone deficiency	GCN	26	78	No

<i>STARD7</i>	Familial adult myoclonic epilepsy type 2	ATTTC	≥661-735	≥3305-3675	Yes
<i>TAF1</i>	X-linked dystonia parkinsonism	CCCTCT	30-55	180-330	Possible
<i>TBP</i>	Spinocerebellar ataxia type 17	CAG or CAG/CAA	≥43-66	≥129-178	No
<i>TCF4</i>	Fuchs endothelial corneal dystrophy type 3	CTG	>50	>150	No
<i>TNRC6A</i>	Familial adult myoclonic epilepsy type 6	ATTTC	N/A	N/A	N/A
<i>XYLT1</i>	Baratela-Scott syndrome	CGG	120-800	360-2400	Yes
<i>YEATS2</i>	Familial adult myoclonic epilepsy type 4	ATTTC	N/A	N/A	N/A
<i>ZIC2</i>	Holoprosencephaly type 5	GCN	25	75	No

Supplemental Table S5: Genomic regions and label IDs used for the manual *de novo* assembly size estimate. Table adjusted from van der Sanden et al. (2024).

Gene	Genomic region of repeat expansion locus (GRCh38/hg38)	Start label of interest ID [#]	End label of interest ID [#]	Reference length between labels of interest (bp)
<i>CNBP</i>	chr3:129,169,450-129,181,839	26,243	26,246	12,389
<i>DMPK</i>	chr19:45,752,584-45,771,947	5,926	5,927	19,363
<i>RFC1</i>	chr4:39,343,732-39,350,590	7,723	7,724	6,858

The label IDs are chromosome specific.

For *CNBP* the locus entailed four labels to prevent resolution errors from labels in proximity that can be falsely read as one extra-bright signal.

Supplemental Table S6: Genomic regions and label IDs used for the Molecule Distance Script workflow. Table adjusted from van der Sanden et al. (2024).

Gene	Genomic region of repeat expansion locus (GRCh38/hg38)	Start label of interest ID#	End label of interest ID#	Reference length between labels of interest (bp)
<i>CNBP</i>	chr3:129,168,220-129,186,501	26,242	26,247	18,281
<i>DMPK</i>	chr19:45,740,976-45,829,452	5,925	5,930	88,476
<i>RFC1</i>	chr4:39,339,156-39,362,887	7,722	7,725	23,731

The label IDs are chromosome specific.

For all three genes the locus entailed more than two labels to make sure the labels of interest cover the entire gene region containing the repeat expansion since the molecule distance script workflow constitutes an automated workflow and does not allow for individual manual tweaking.

Sample	Manual de novo assembly	Local guided assembly
CNBP_01		
CNBP_02		
CNBP_03		
CNBP_04		
CNBP_05		
CNBP_06		
CNBP_07		
CNBP_08		
CNBP_10		
CNBP_11		
CNBP_12		
CNBP_13		
CNBP_14		
CNBP_15		
CNBP_16		
CNBP_17		
CNBP_18		
CNBP_19		
CNBP_20		
CNBP_21		
CNBP_22		
CNBP_23		
CNBP_24		
CNBP_25		
CNBP_26		
DMPK_01		
DMPK_02		
DMPK_03		
DMPK_04		
DMPK_05		
DMPK_06		
DMPK_07		
DMPK_08		
DMPK_09		
DMPK_10		
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DMPK_28		
DMPK_29		
DMPK_30		
RFC1_01		
RFC1_02		
RFC1_03		
RFC1_04		
RFC1_05		
RFC1_06		
RFC1_07		
RFC1_08		
RFC1_09		
RFC1_10		
RFC1_11		
RFC1_12		
RFC1_13		
RFC1_14		
RFC1_15		
RFC1_16		
RFC1_17		
RFC1_18		
RFC1_19		
RFC1_20		
RFC1_21		
RFC1_22		
RFC1_23		
RFC1_24		
RFC1_25		
RFC1_26		
RFC1_27		
RFC1_28		
RFC1_29		
RFC1_30		

Supplemental Fig S1: Heatmap presenting the individual results from each OGM workflow. Blue color means repeat expansion detected and orange color means no repeat expansion detected.

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