

***Candida albicans* isolates contain frequent heterozygous structural variants and transposable elements within genes and centromeres**

Authors

Ursula Oggenfuss¹, Robert T. Todd^{1,2}, Natthapon Soisangwan¹, Bailey Kemp¹, Alison Guyer¹, Annette Beach¹, Anna Selmecki^{1, #}

1 Department of Microbiology and Immunology, University of Minnesota, Minneapolis, MN, USA

2 Department of Biology, Bard College, Annandale-on-Hudson, New York, NY, USA

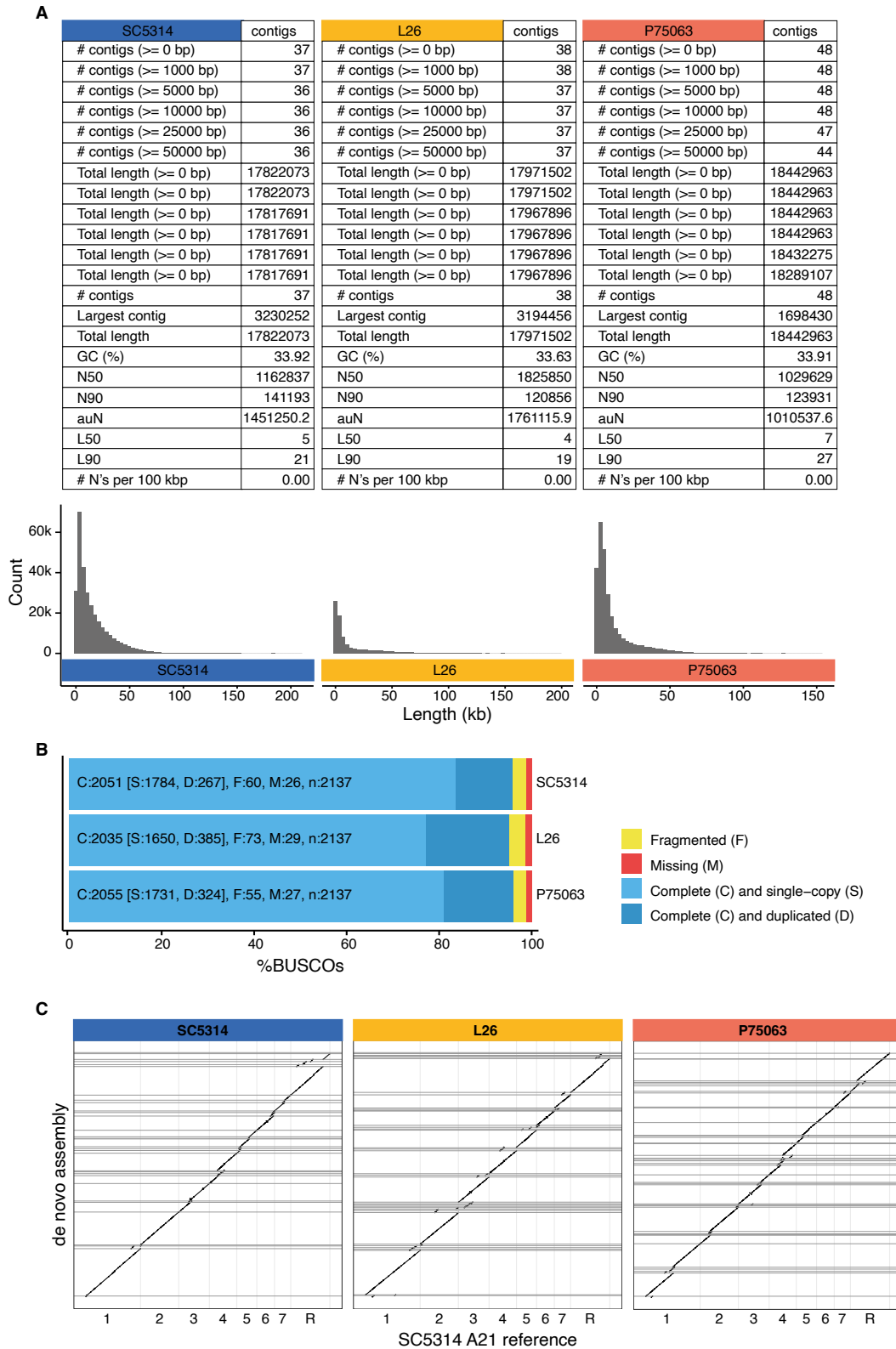
Supplementary Figures and Tables

Supplementary Table S1: TE fasta consensus sequences from the literature

Supplementary Table S2. Strain Table clinical isolates (including SRA for the long reads and PCR results)

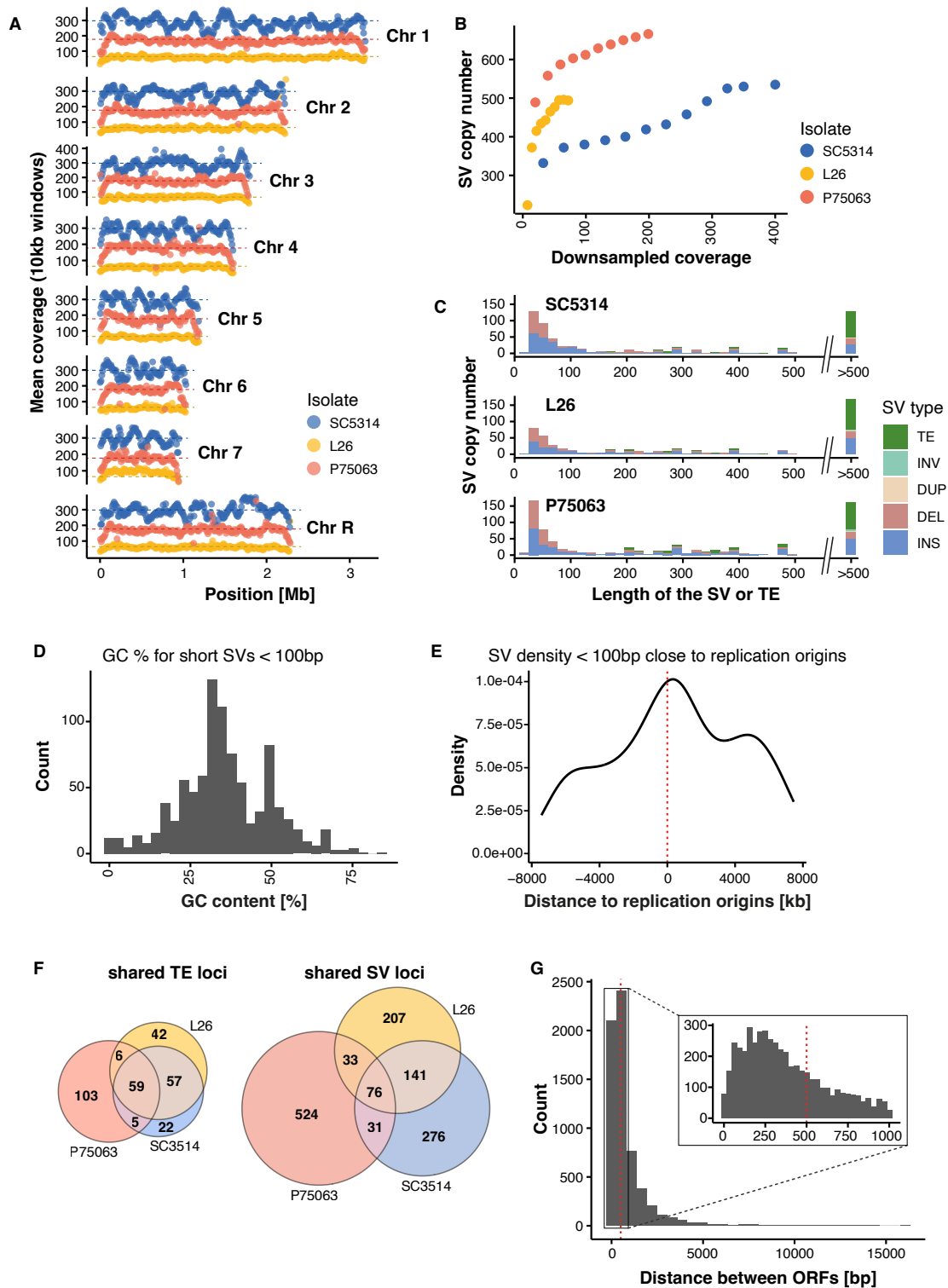
Supplementary Table S3. Primer table and primer conditions

Supplementary Table S4: SVs

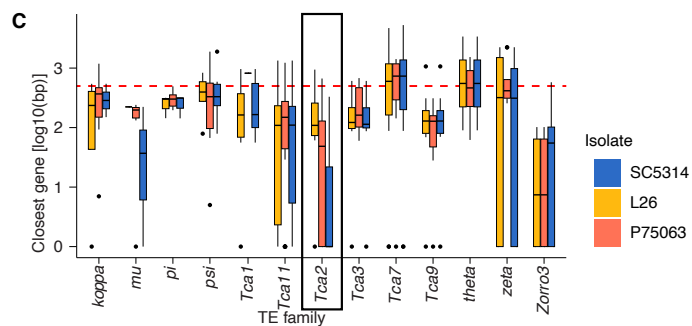
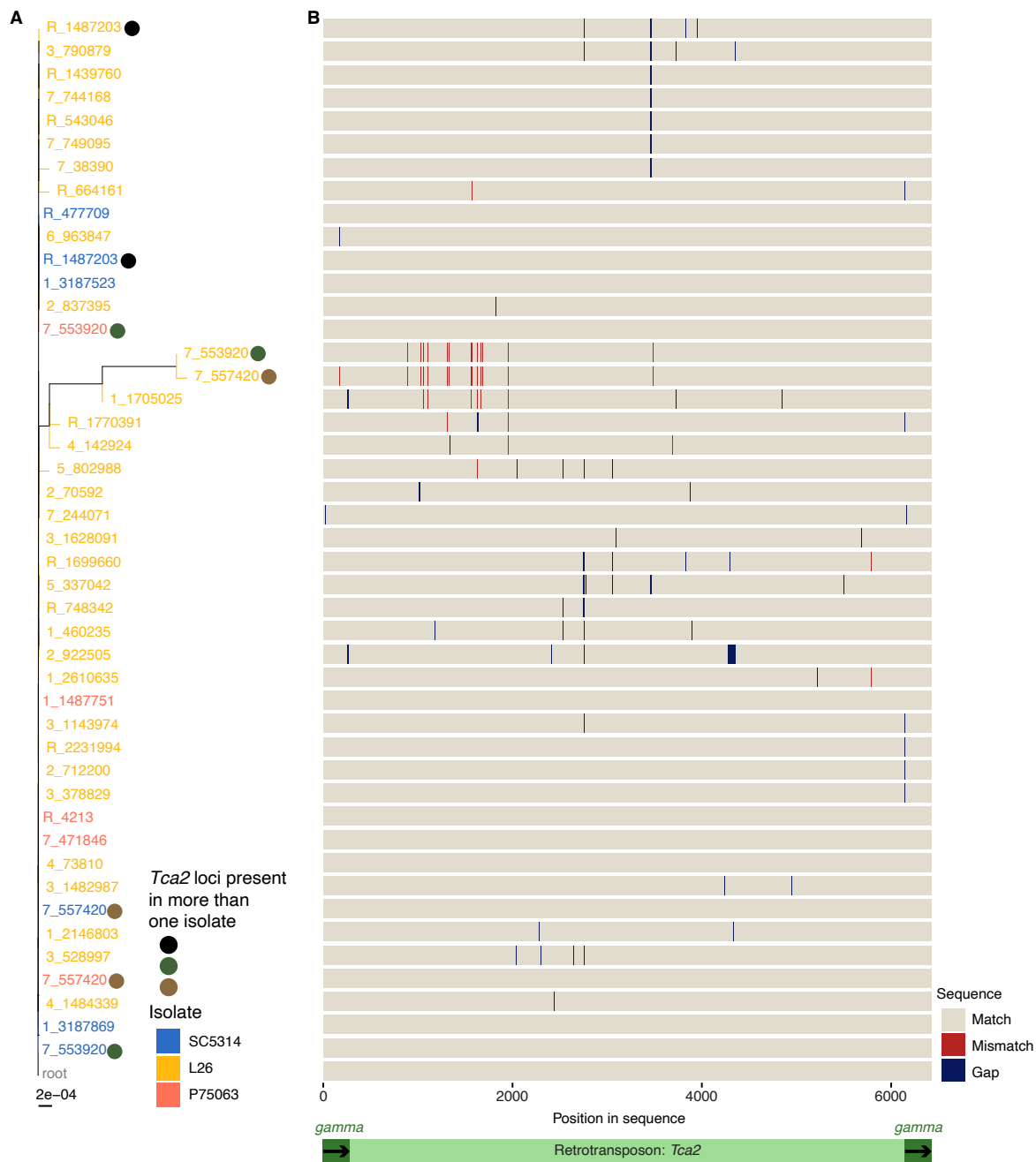


Supplementary Figure S1. *De novo* assembly. A) Assembly quality assessment with QUAST. The Histogram shows read length distribution per isolate. The longest reads have a length of 208.83 kb

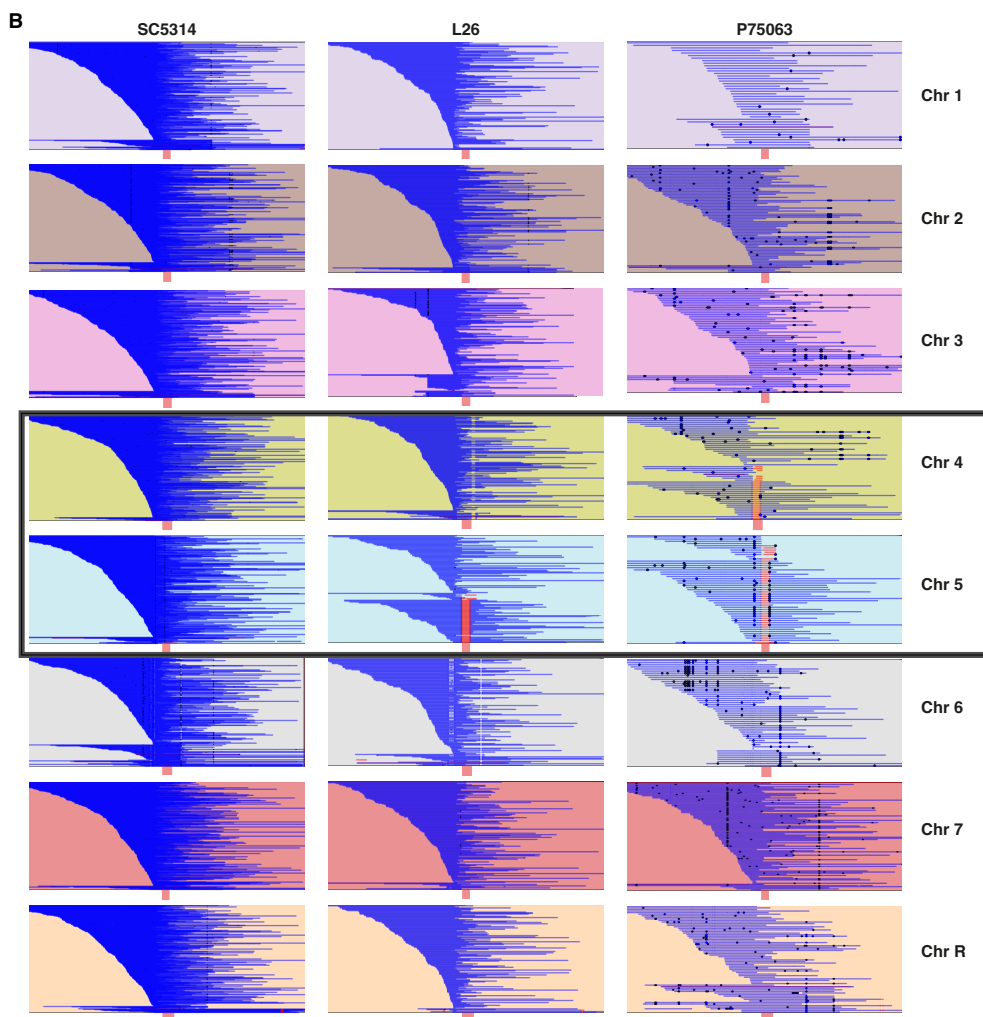
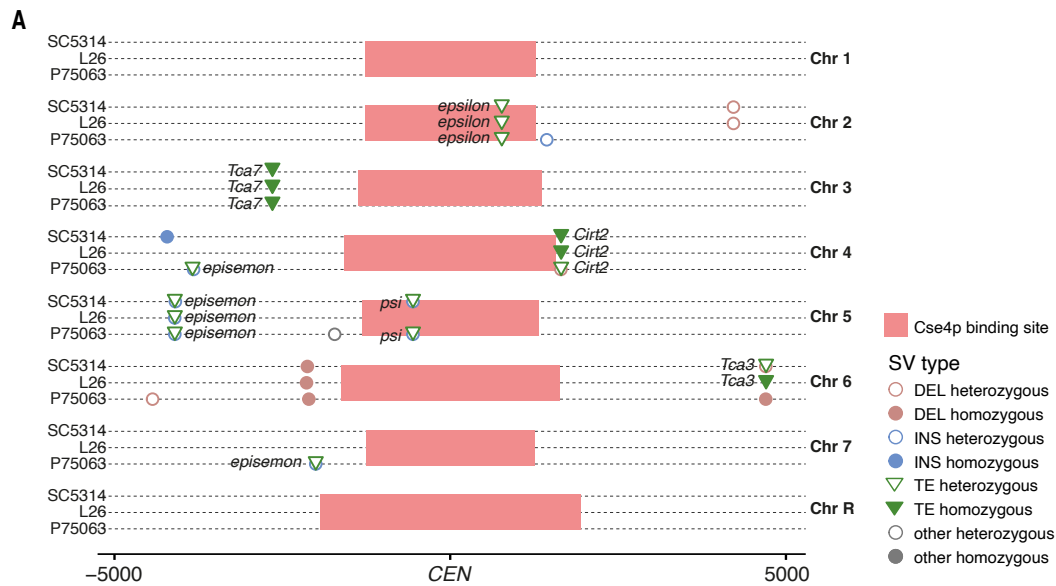
(SC5314), 198.49 kb (L26) and 153.01 kb (P75063). **B**) Genome completeness with BUSCO. BUSCO was tested on 2137 genes c conserved in Saccharomycotina. **C**) Contigs of the de novo assembly aligned to the reference genome SC5314 A21 (x-axis).



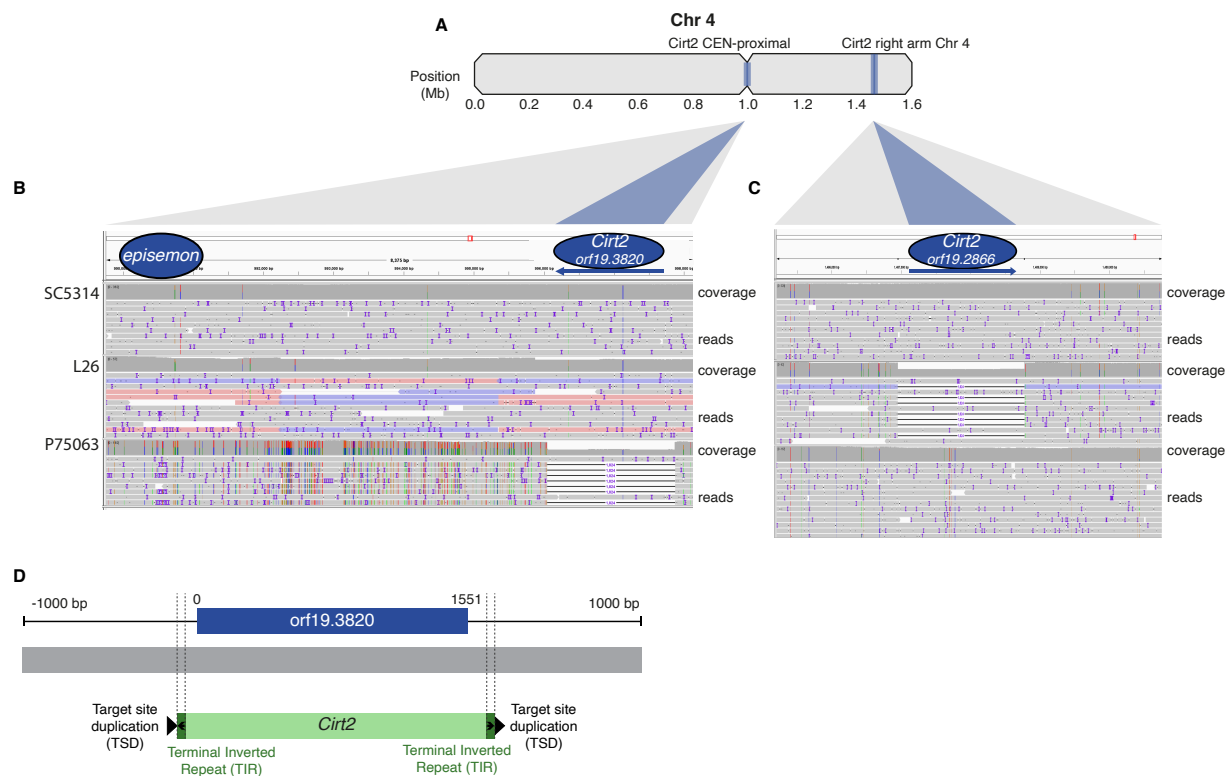
Supplementary Figure S2. Genomic landscape, TEs and SVs in three clinical isolates. **A)** Mean read depth coverage for 10,000 bp windows for SC5314, L26 and P75063. Dotted lines indicate the genome-wide mean coverage for each genome (73.7x in L26, 331.5x in SC5314 and 203.9x in P75063). **B)** Read depth downsampling analysis. Each dot represents the number of detected SVs with DELLY on a reduced set of reads. **C)** Length distribution of SVs. For SVs, the length is taken from the output of DELLY, for TEs the length of the consensus sequence. Colors by SV type (green: TE = transposable element; turquoise: INV = inversion; apricot: DUP = duplication; red: DEL = deletion; blue: INS = insertion). **D)** GC content distribution for short SVs (< 100 bp) **E)** Density of SVs < 100 bp detected around replication origins in a window of -7.5kb to 7.5kb. Replication origin positions were extracted from CHIPseq data (Tsai et al (2014)). **F)** TEs and SV identical in location between the isolates SC5315, L26 and P75063. **G)** Distribution of distance between ORFs. The red line indicates a distance of 500bp. The box shows a more fine-scale zoom in to a distance of 100 bp.



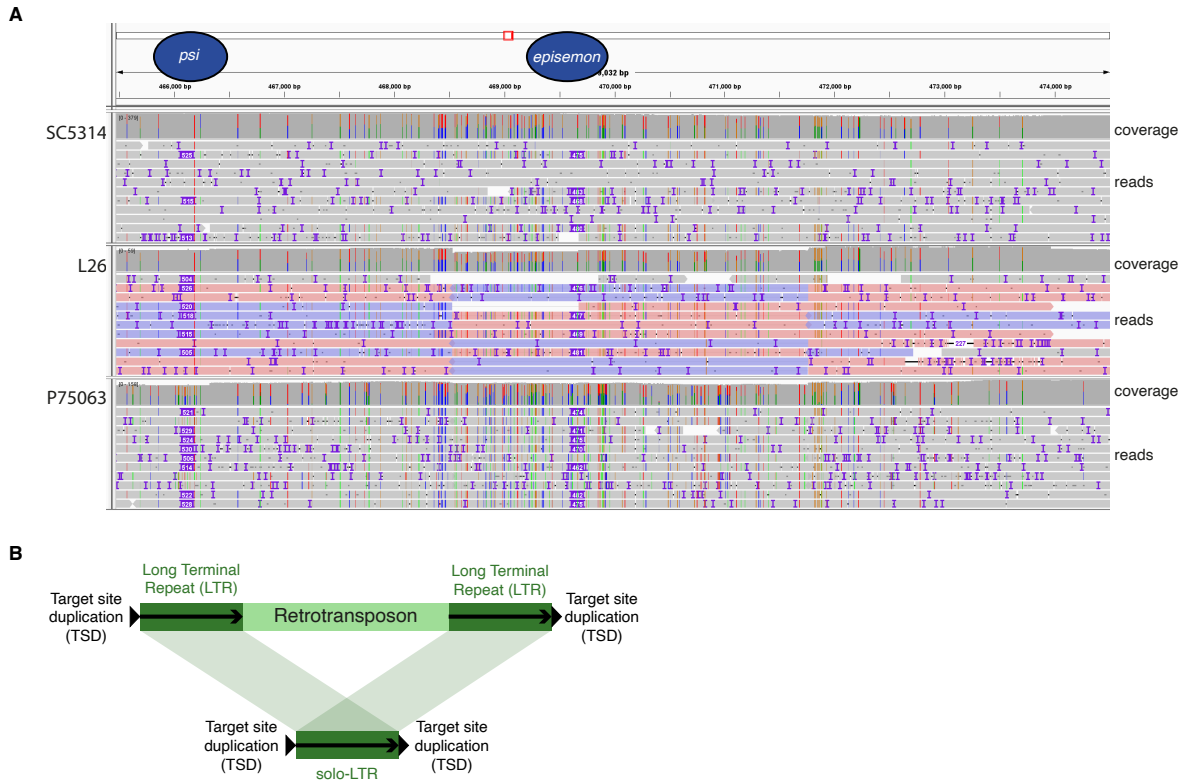
Supplementary Figure S3. Positions and schematic of the *Tca2* TE. **A)** Phylogenetic tree based on the multiple sequence alignment. Circles indicate copies that are present in more than one isolate (Fixed = present in all isolates; 2 copies = present in two isolates). Colors indicate the isolate. The tree is rooted with the consensus sequence. **B)** Multiple sequence alignment for the whole sequence of *Tca2*. Colors indicate sequence similarity (grey = match; red = mismatch; blue = gap). The schematic at the bottom represents *Tca2* and its *gamma* LTRs **C)** Distance to the closest annotated ORF for each TE copy. Only TE families with ten or more copies were used. Colors indicate the isolate.



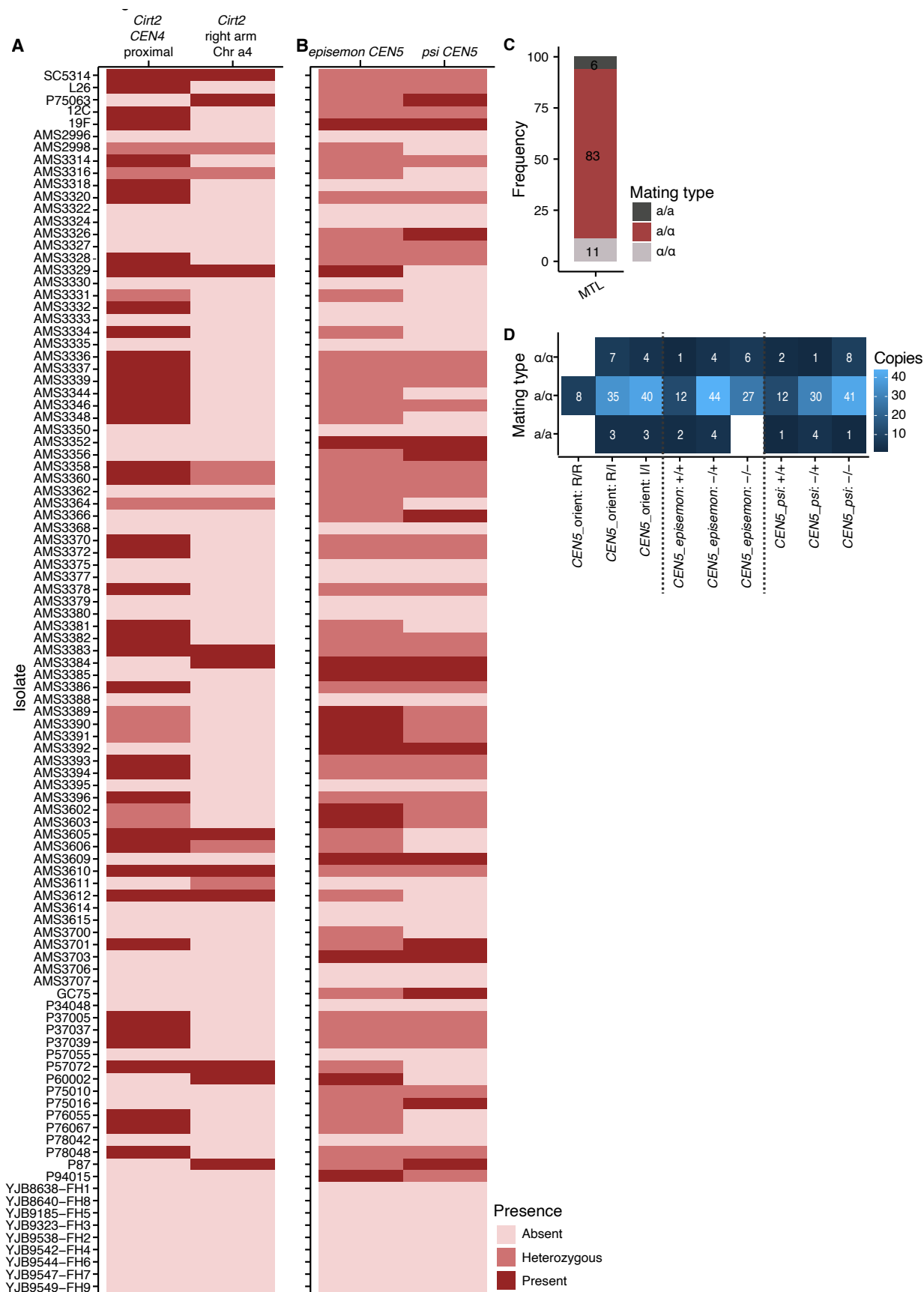
Supplementary Figure S4. SVs at *CEN* positions. **A)** Schematic of SVs at each Cse4p binding site (pink region) and each 5000 bp upstream and downstream. Figure adapted from main Figure 1. Colors indicate the SV type in comparison with the reference genome SC5314 A21 (green: TE = transposable element; red: DEL = deletion; blue: INS = insertion, grey = other types of SV (inversions, duplications, translocations)). The fill represents zygosity (filled symbol = homozygosity, unfilled symbol = heterozygosity). The names of TE families are added. **B)** Ribbon plots for each *CEN* position for each isolate. The pink box represents the Cse4p binding site for each chromosome. Cse4p/CENP-A binding delineates the central core sequence of the centromere (Sanyal et al. 2004; Ketel et al. 2009).



Supplementary Figure S5. Positions and schematic of the *Cirt2* TE. **A)** Schematic of chr4 and *Cirt2* positions (blue lines). **B)** Mapped reads (IGV screenshot) of *CEN4* and nearby features for a subset of SC5314, L26, and P75063 reads. The *CEN4* solo-LTR *episemon* and *Cirt2* are indicated above the reads. **C)** Mapped reads in the region around *orf19.2866*, right arm copy of *Cirt2* in SC5314, L26, and P75063. The chr4:1,486,953-1,488,776 deletion (“*orf19.2866* deletion”) is indicated above the reads, which is 1,824 bp and spans the annotated *orf19.2866* plus some bases upstream and downstream. **D)** Schematic of the *Cirt2* transposon, that contains a transposase as a coding region (blue), terminal inverted repeats at both sides of the element (green), and is flanked by a short target site duplication (typically TA; black).



Supplementary Figure S6. *CEN5* structural variants depicted by whole genome sequencing (WGS) visualization. A) Mapped reads (IGV screenshot) of *CEN5* and nearby features for SC5314, L26, and P75063. The *CEN5* solo-LTRs are indicated in the track above the reads. The locations of the ~470 bp *psi* and ~523 bp *episemon* solo-LTRs are indicated above the reads with a triangle and in the reads with a purple bar. **B)** Schematic of retrotransposon non-homologous recombination that leads to the presence of solo-LTRs. Full length LTR-retrotransposons contain two long terminal repeats (LTR; dark green), and superfamily specific coding regions (bright green). Upon insertion, a short target site duplication (TSD) is created. After the non-homologous recombination, only the TSD and a single copy of the LTR remains. Non-homologous recombination can also happen between two full length copies or between two LTRs of independent copies, leading to a deletion beyond the TE.



Supplementary Figure S7. Clinical isolates. **A)** Presence/absence and zygoty of *Cirt2* in the pericentromeric region and on the right arm of chr4. **B)** Mating type distribution of the 100 clinical isolates. **C)** Presence/absence and zygoty of the solo-LTRs *episemon* and *psi* in the pericentromeric region of chr5. **D)** Distribution of mating types over the different pericentromeric TEs.