

SUPPLEMENTAL FIGURES

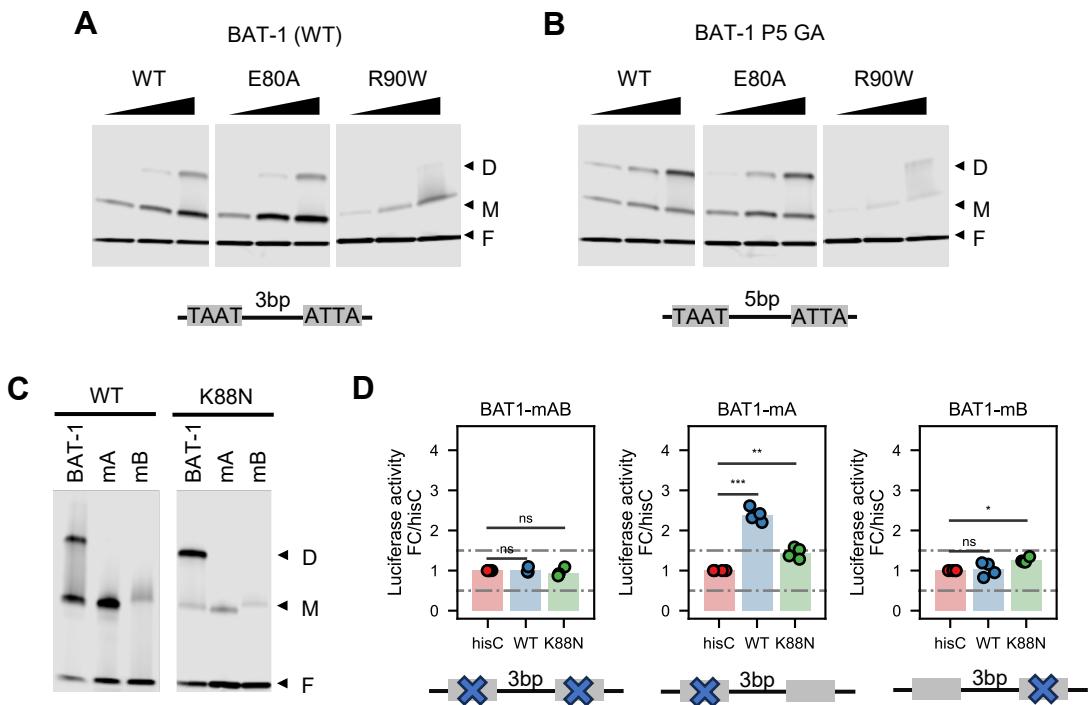
Aberrant homeodomain-DNA cooperative dimerization underlies distinct developmental defects in two dominant *CRX* retinopathy models

Authors

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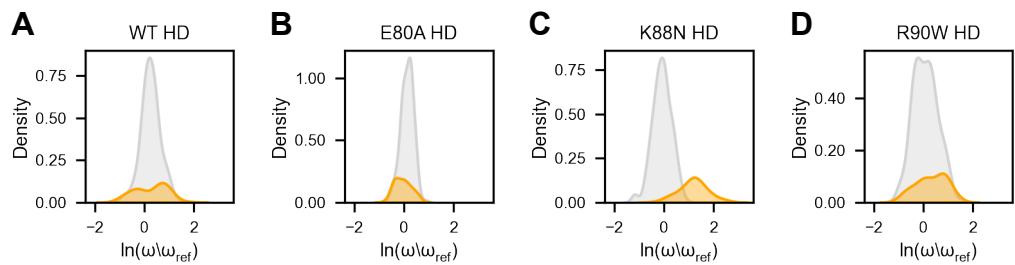
Affiliations

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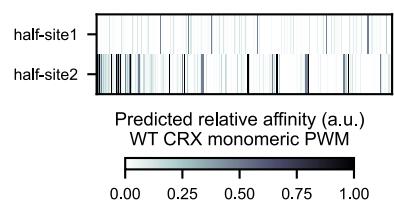
Supplemental Fig S1

Supplemental Fig S1 – P3 configuration and intact half-sites are required for K88N HD cooperative binding and transactivation activity. (A,B) EMSA gel images showing increasing amounts of WT, E80A, and R90W HD peptides bound to a fixed amount of BAT-1 (WT) or P5 probes. The cartoon underneath each gel image shows the dimeric HD motif configuration and is labeled with the spacer length. Note the WT HD EMSA gel images are the same as in Fig. 1C,D. (C) EMSA gel images showing WT and K88N HD peptides bound to BAT-1 (WT), mA and mB probes. (D) Barcharts of luciferase reporter assays comparing CRX WT and K88N transactivation activity at BAT-1 variant enhancer sequences. The cartoon underneath each barchart shows the HD core motif mutated. Bars represent the mean. *p*-values of one-way ANOVA are annotated: ns: >5e-2; *: <=5e-2; **: <=1e-2; ***: <=1e-3.

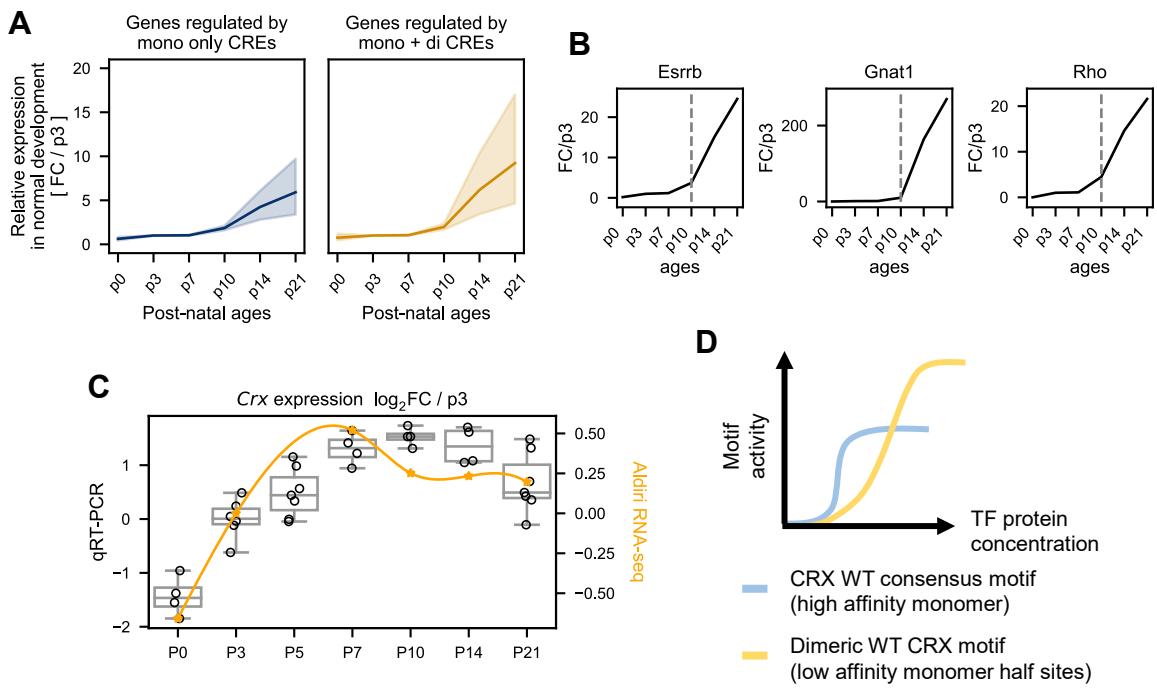


Supplemental Fig S2

Supplemental Fig S2 – WT and mutant CRX HDs demonstrate similar cooperativity profiles at P5 library but distinct cooperativity profiles at P3 library. (A-D) Histograms depicting the distribution of CRX HD relative cooperativity on members of the P5 library (grey) and the P3 library (orange). Relative cooperativity against the reference sequence TAATGCGCTATTA is plotted ($\omega/\omega_{\text{ref}}$). Note the relative cooperativity is presented in the Logarithmic scale. The full relative cooperativity matrix can be found in Supplemental Table S2.

A

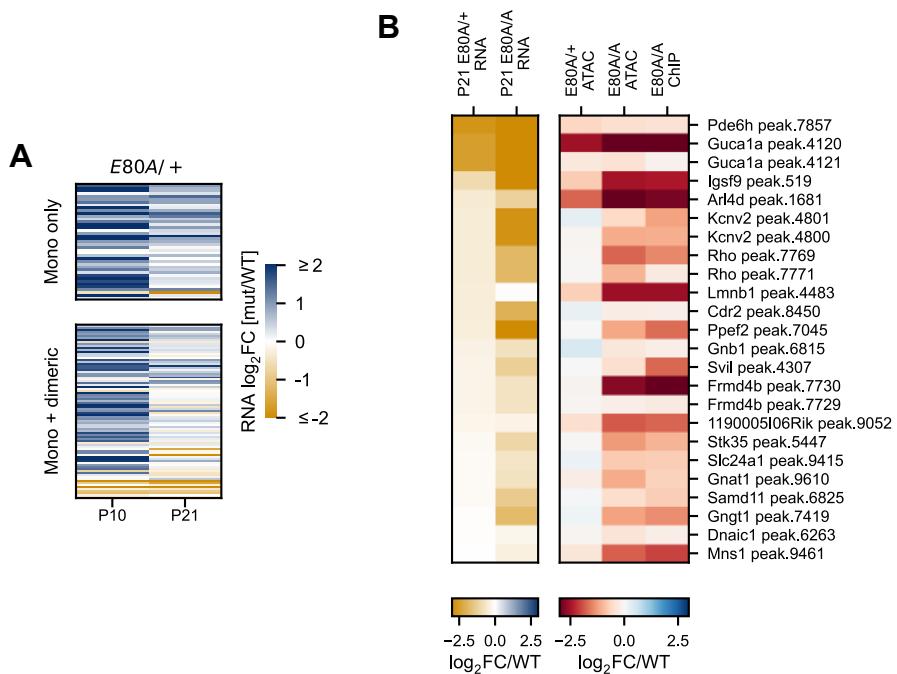
Supplemental Fig S3 – Dimeric K₅₀ HD motifs in Crx^{E80A/A}-reduced ATAC-seq peaks composed of low-affinity half-sites. (A) Heatmap depicting the predicted half-site affinity of FIMO-found dimeric K₅₀ HD motifs under Crx^{E80A/A}-reduced ATAC-seq peaks. The dimeric motifs are ordered by FIMO p-values. The half-site relative affinity, normalized to consensus TAATCC, is predicted using the published WT CRX PWM model (Supplemental Table S4; Methods).



Supplemental Fig S4

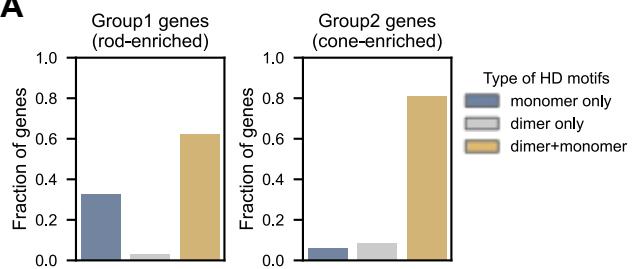
Supplemental Fig S4 – Dimeric K₅₀ HD motifs are associated with stage-specific gene regulation. (A)

Line plots showing the expression dynamics of genes associated with CREs with only monomeric or both monomeric and dimeric K₅₀ HD motifs. Expression levels are normalized to P3 and represented by fold-change. (B) Expression line plots of selected genes associated with monomeric and dimeric K₅₀ HD motifs. (C) Dual-axes box and line plot showing the Crx mRNA developmental expression patterns measured by qRT-PCR (left axis, box and strip plot) and by whole-retina RNA-seq (right axis, line and scatter plot). Expression levels are normalized to post-natal day 3 (P3) and presented by log-fold-change. The developmental RNA-seq data in Supplemental Fig. S4A-S4C is from Aldiri et al. 2017. (D) Diagram depicting the stage-specific activity model of CRX WT high-affinity monomeric motifs and dimeric motifs with individually low-affinity half-sites.



Supplemental Fig S5

Supplemental Fig S5 – CRX-DAGs regulated by dimeric K₅₀ HD motifs are selectively down-regulated in *CrxE80A* mutant retinas. (A) Heatmap comparing the CRX-DAG expression changes in the *CrxE80A/+* retinas at ages of post-natal day 10 (P10) and day 21 (P21). The gene sets in the heatmaps are as defined in Fig. 6A. The order of the genes is identical to that in Fig. 6B. (B) Heatmaps showing down-regulated genes with correspondingly decreased chromatin accessibility at associated CREs in the *CrxE80A* retinas. The genes and CREs are ordered first by RNA-seq logFC than by ATAC-seq logFC in the *CrxE80A/A* retinas. The peak.id denotes the unique CRX ChIP-seq peak associated with a gene. The full CRX bound CRE-gene association matrix can be found at Supplemental Table S6.

A

Supplemental Fig S6 – Dimeric K₅₀ HD motifs are associated with cell type-specific gene regulation.
(A) Barcharts showing the distribution of cone- vs rod-enriched CRX-DAGs associated with CREs containing monomeric, dimeric, or both types of K₅₀ HD motifs. The definition of Group1 and 2 genes is taken directly from our previous publication Zheng et al. 2023.

A

Primer F padding 10bp BC Primer R

GTAGCGTCTGCCGT **GAATTC** ----CRE Sequence---- **ACTAGT** **C** **GCATGC** NNNNNNNNNN **CGGCCG** CCAACTACTACTACAG

EcoRI Spel SphI EagI

B

| Category | | N | Include in MPRA lib | HD site mutants | # BC/CRE | Total testing CREs |
|----------|---|------|---------------------|-----------------|----------|--------------------|
| 1 | K88N/N gained | 466 | | | 4 | |
| 2 | E80A/A gained | 214 | | | 4 | |
| 3 | E80A/A lost | 309 | | | 4 | 9884 |
| 4 | CREs for selected genes | 55 | | | 4 | |
| 5 | Rest of ChIP peaks not in categories 1-4 (expected) | 8798 | 900 | 799 | 4 | 6796 |
| 6 | Basal | | 1 | | 20 | 20 |
| 7 | Scrambles | | 150 | | 4 | 600 |
| 8 | + Ctrl | | 20 | | 6 | 120 |
| | TOTAL | | 4345 | | | 17420 |

Supplemental Fig S7 – MPRA oligo design and library composition. (A) Diagram of a typical testing MPRA oligo. The restriction enzyme cut sites used for cloning, primer sites for amplification, and testing CRE and barcode locations are noted. (B) Table displaying the composition of MPRA library designed and ordered. The gained and lost categories were defined by comparing the ChIP-seq signal intensity (Methods; MPRA GitHub repository).