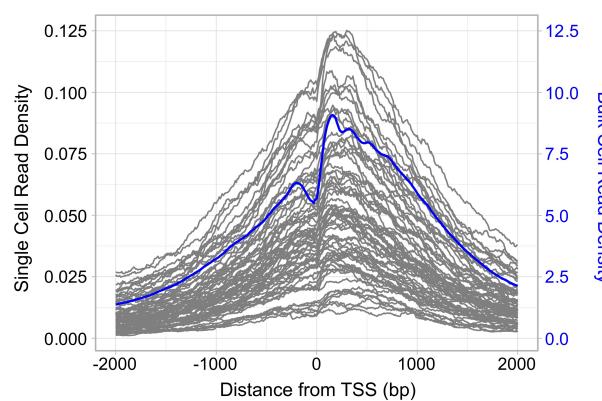
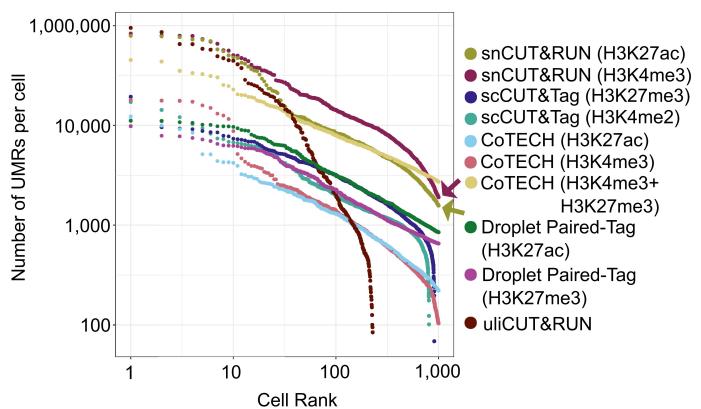
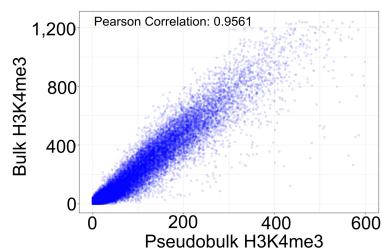
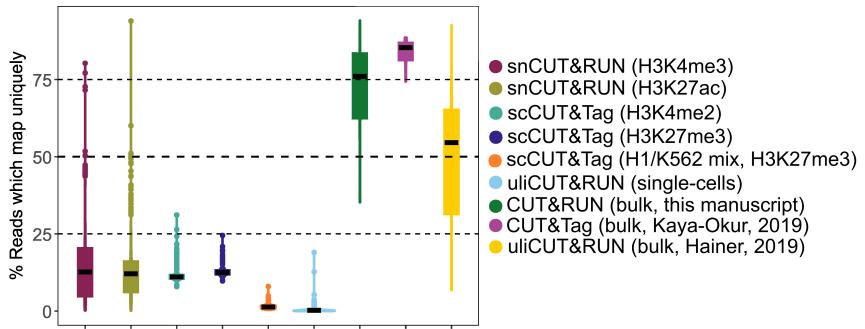
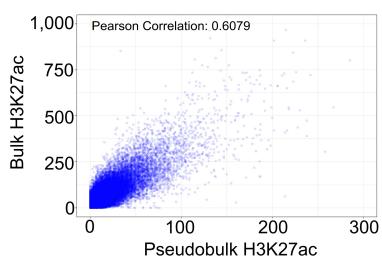
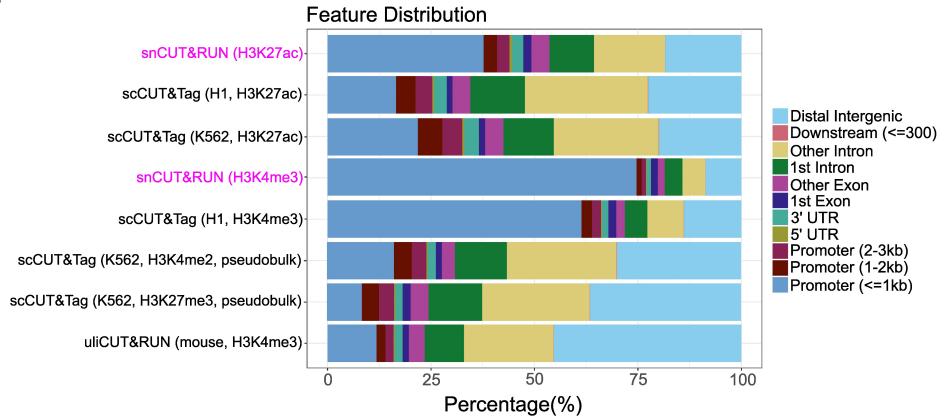


Supplementary Figures

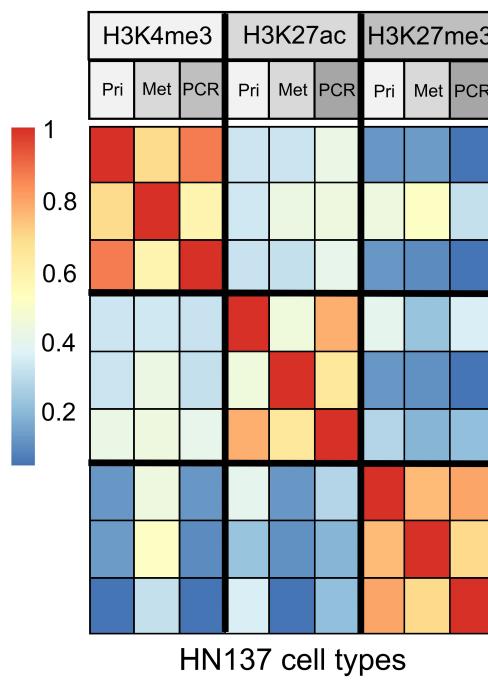
Single-nucleus CUT&RUN deciphers the function of intrinsic and genomics-driven epigenetic heterogeneity in head and neck cancer progression

Howard J. Womersley, Daniel Muliaditan, Ramanuj DasGupta*, Lih Feng Cheow*

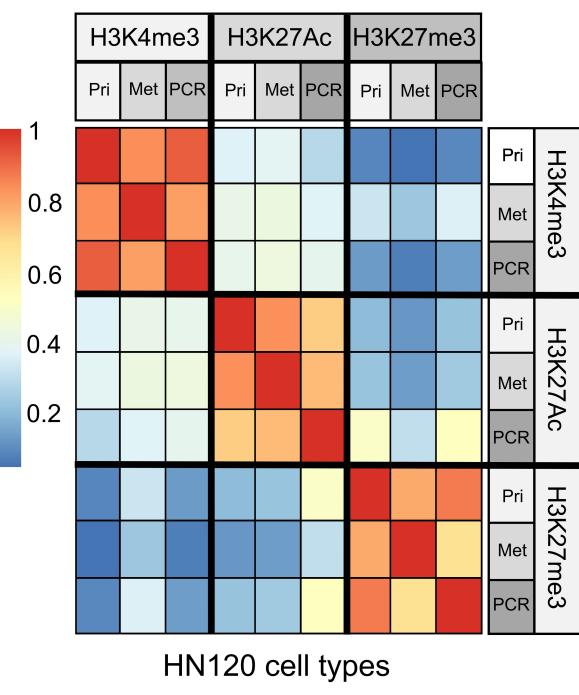
*Corresponding authors. Email: dasguptar@gis.a-star.edu.sg (RD), bieclf@nus.edu.sg (LFC)

A**D****B****E****C****F**

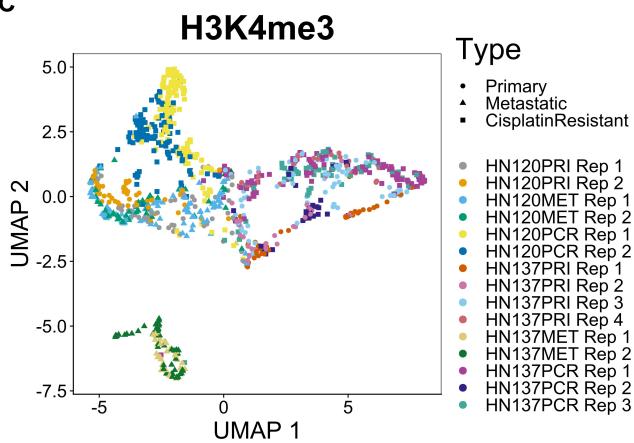
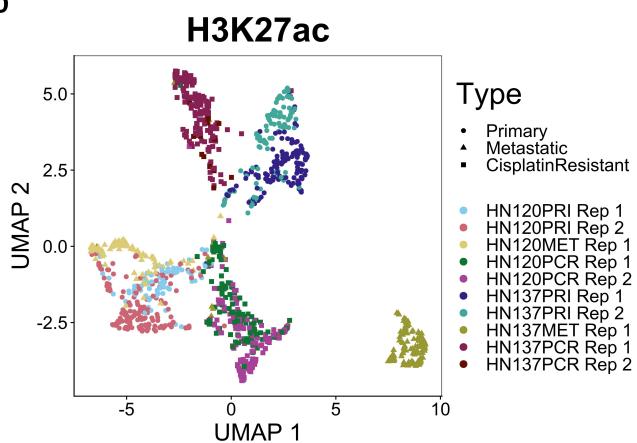
Supplemental Fig. S1. Quality metrics for training and validation data set. (A) Comparison of single cell and bulk cell read density for H3K4me3 histone mark relative to global transcriptional start sites. Read density profiles for single cells closely match bulk cell data, with clear nucleosome depletion regions. (B) Pearson's correlation between training set single cells and bulk data for H3K4me3 histone mark. (C) Pearson's correlation between training set single cells and bulk cell data for H3K27ac histone mark. (D) Scatter plot ranking the top 1,000 libraries with the most to the least number of unique mapped reads. snCUT&RUN data is compared with scCUT&Tag (Kaya-Okur et al. 2019), CoTECH (Xiong et al. 2021) and Droplet Paired-Tag (Xie et al. 2023). (E) Violin- and boxplots indicating the percentage of reads which map to the genome uniquely after PCR duplicates have been removed. (F) ChIPseeker (Yu et al. 2015) annotation of the genomic localization of H3K27ac and H3K4me3 peaks in the snCUT&RUN dataset, compared to the genomic localization of probed histone marks in the scCUT&Tag and uliCUT&RUN bulk/pseudobulk datasets.

A

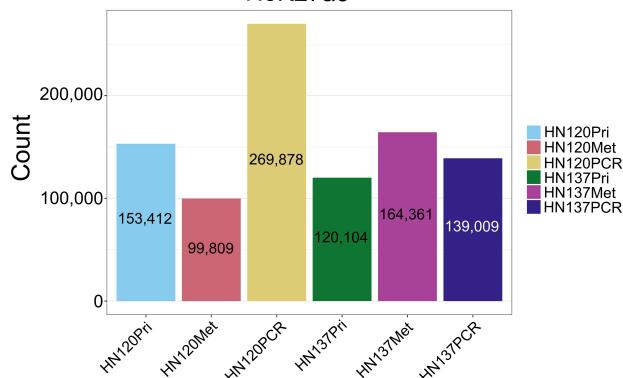
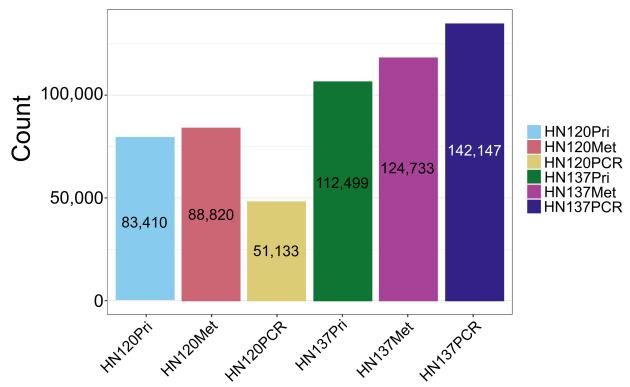
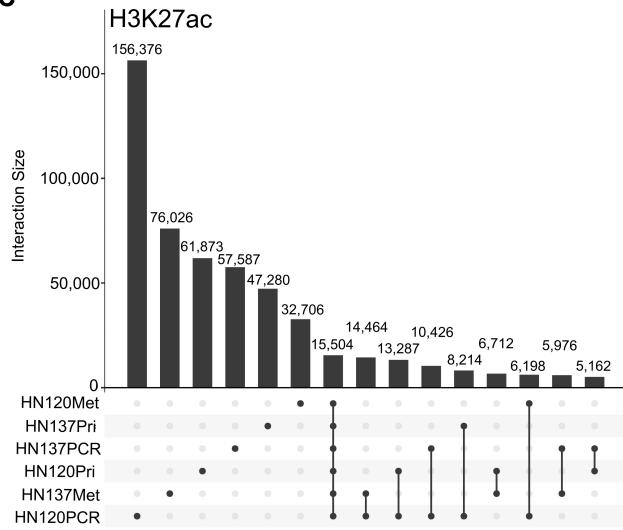
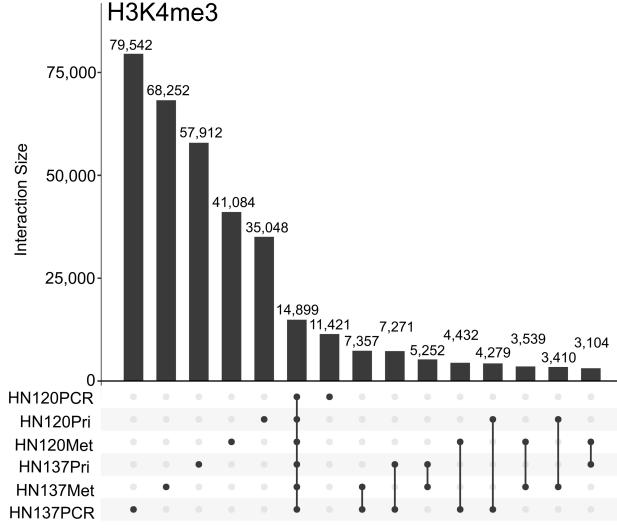
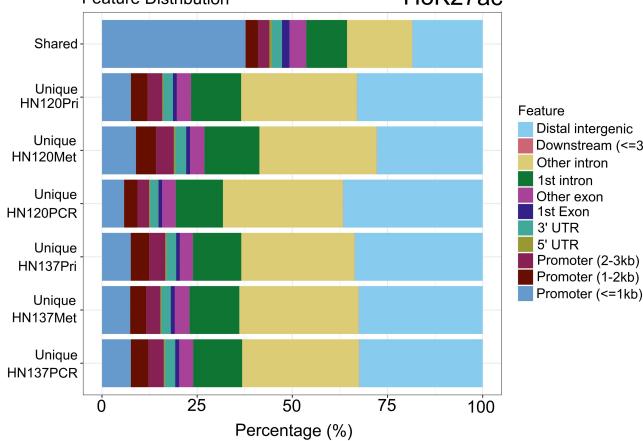
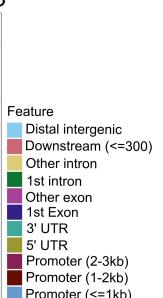
HN137 cell types

B

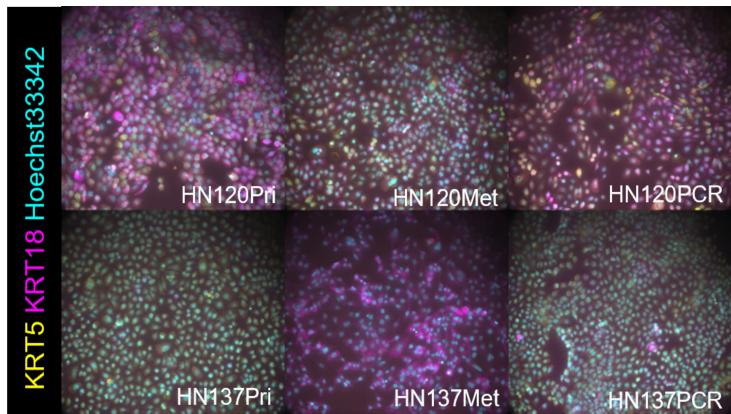
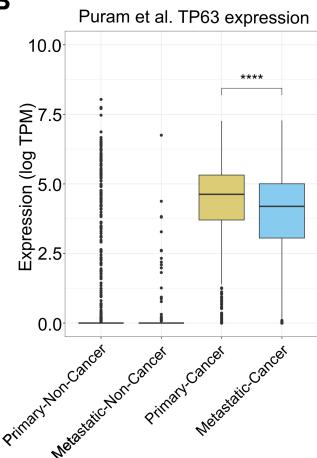
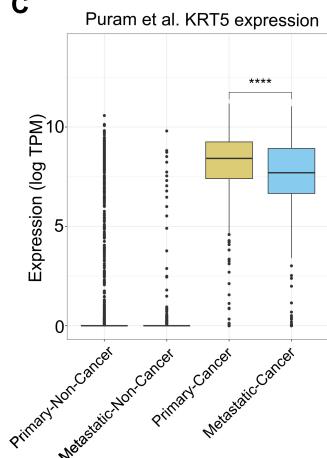
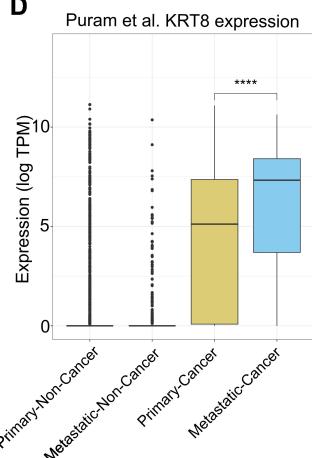
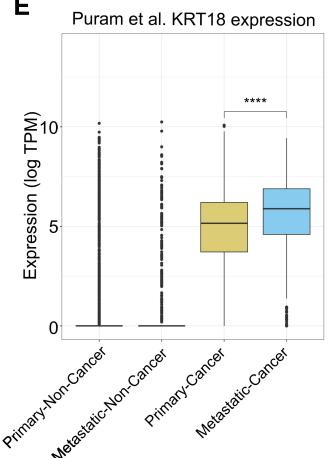
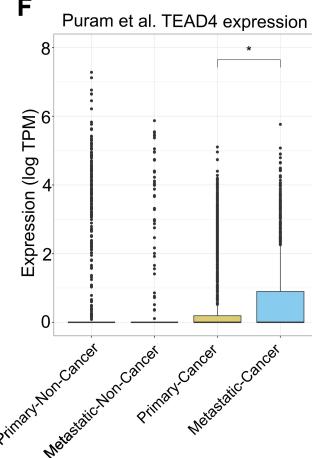
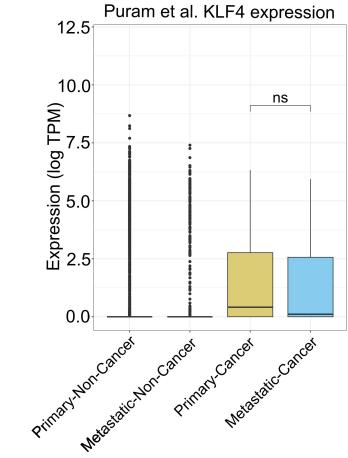
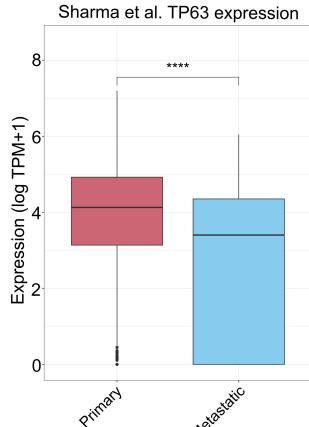
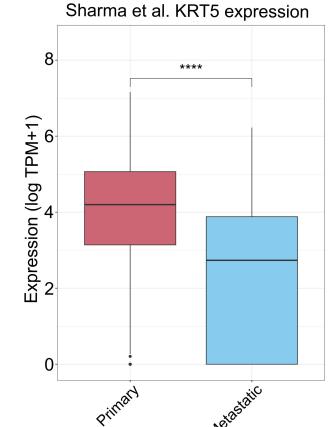
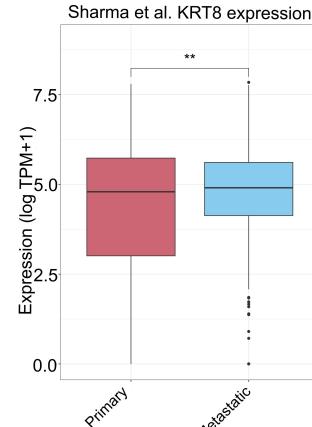
HN120 cell types

C**D**

Supplemental Fig. S2. Pearson correlation matrices of PRI, MET and PCR cell types for H3K4me3, H3K27ac and H3K27me3 histone marks. (A) Similarity matrix for cells from patient HN137. For all histone marks, HN137MET cells have diverged more from HN137PRI than HN137PCR cells. The largest distinction between cell types is between HN137PRI and HN137MET for H3K27ac. (B) Similarity matrix for cells from patient HN120. HN120PRI and HN120PCR are more closely associated than HN120MET for both H3K4me3 and H3K27me3 histone marks. For H3K27ac, HN120PCR cells are more distinct from HN120PRI and HN120MET cells. (C) and (D) UMAPs of H3K4me3 (C) and H3K27ac (D) profiles of HN120 and HN137 cells annotated by replicates.

A**H3K27ac****B****H3K4me3****C****D****E****Feature Distribution****H3K27ac****Feature Distribution****H3K4me3**

Supplemental Fig. S3. Global H3K4me3 and H3K27ac profiles of primary and progressed PDCs. (A) and (B) Number of H3K27ac (A) and H3K4me3 (B) peaks achieved per cell line profiled with snCUT&RUN. (C) and (D) snCUT&RUN number of H3K27ac (C) and H3K4me3 (D) peaks that are either cell line specific or shared by two or more cell lines. (E) and (F) ChIPseeker (Yu et al. 2015) annotation of the genomic localization of shared and sample unique H3K27ac (E) and H3K4me3 (F) peaks in the snCUT&RUN dataset.

A**B****C****D****E****F****G****H****I****J**

Supplemental Fig. S4. Differential keratin expression between primary and progressed HNSCC

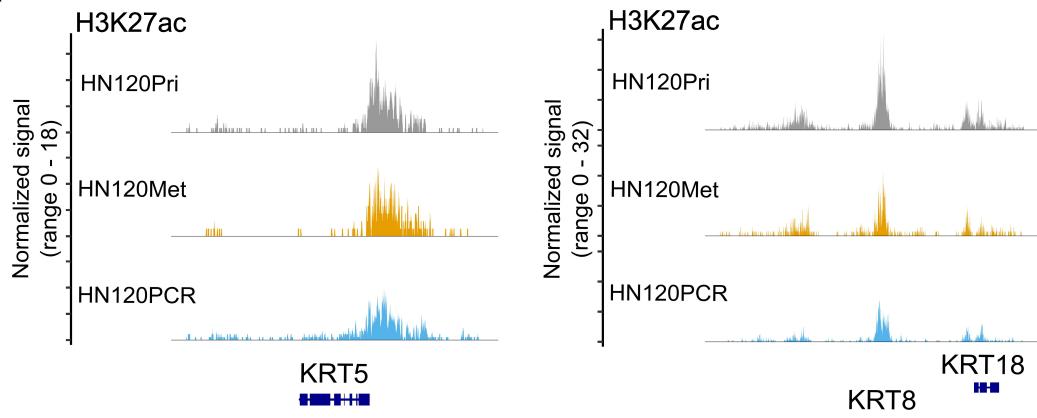
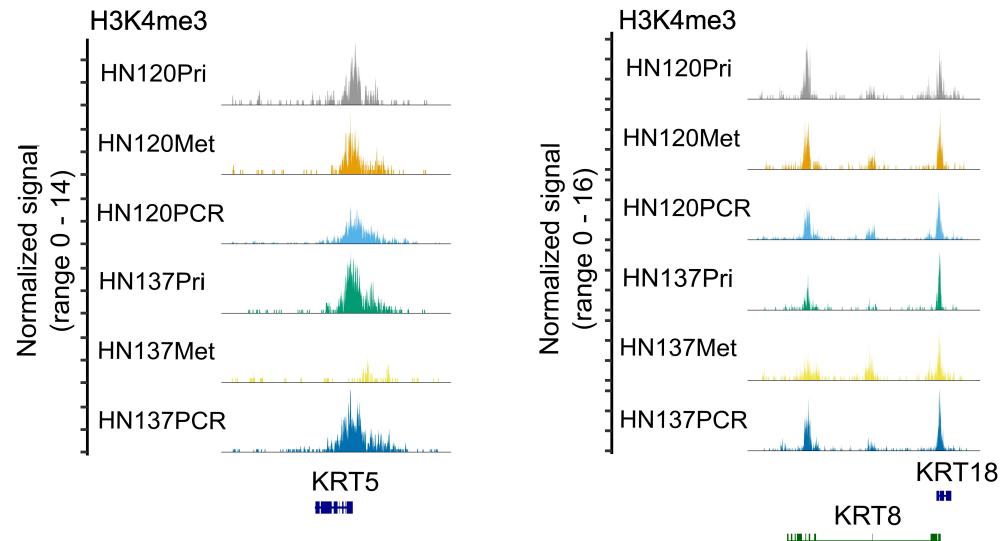
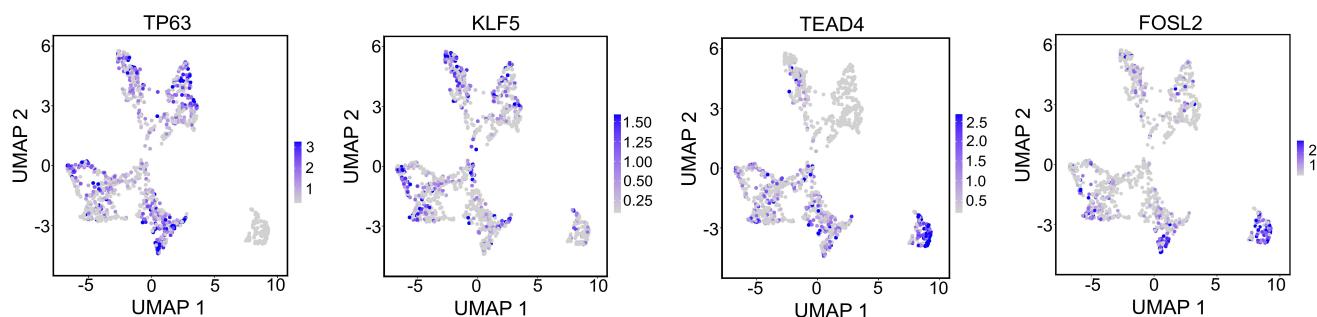
(A) Immunofluorescence image of KRT5 (yellow) and KRT18 (magenta) in HN120 and HN137

PDCs. **(B-G)** Expression data from Puram et al. (Puram et al. Cell, 2017) of various keratins and key

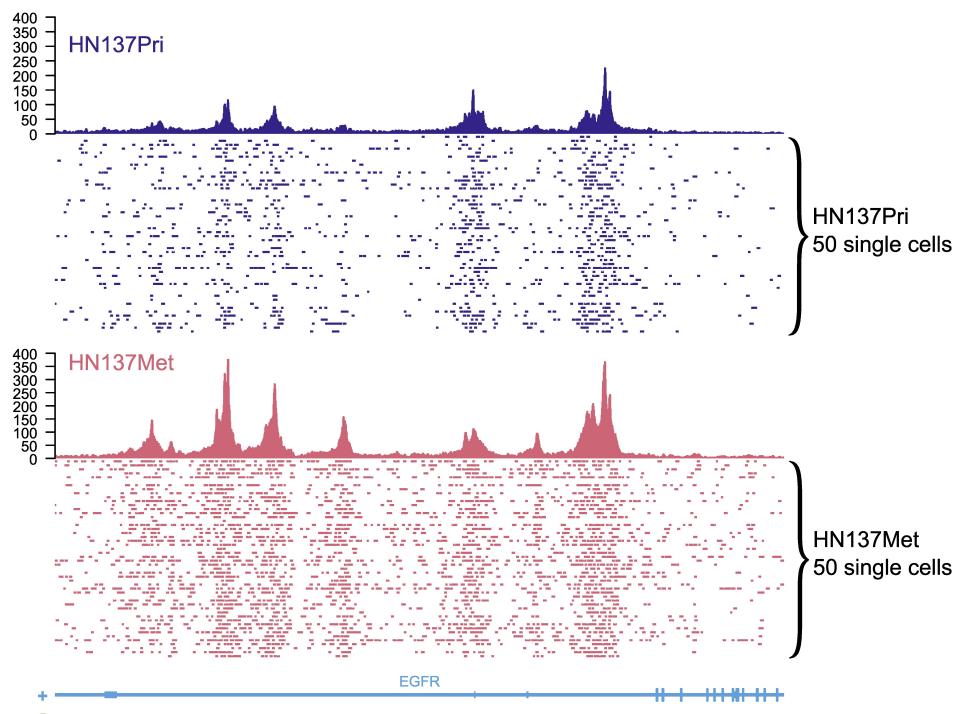
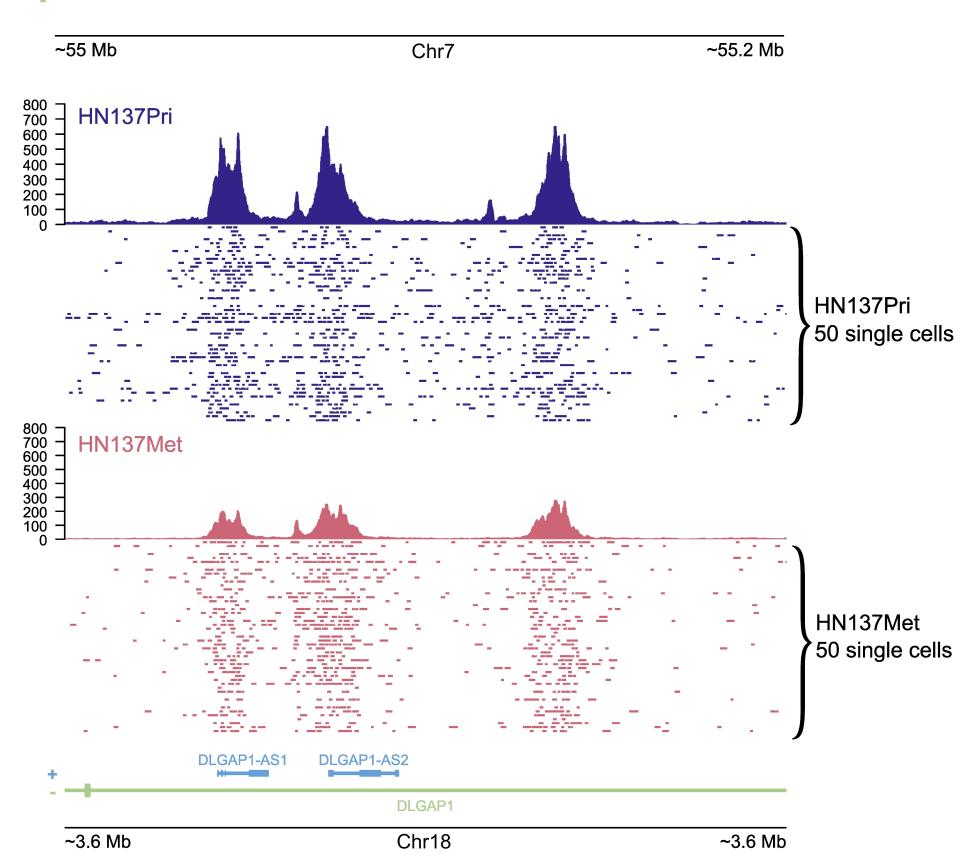
transcription factors in keratinocyte differentiation, comparing primary tumours and metastatic

tumours. **(H-J)** Gene expression of *TP63*, *KRT5* and *KRT8*, comparing primary tumour and metastatic

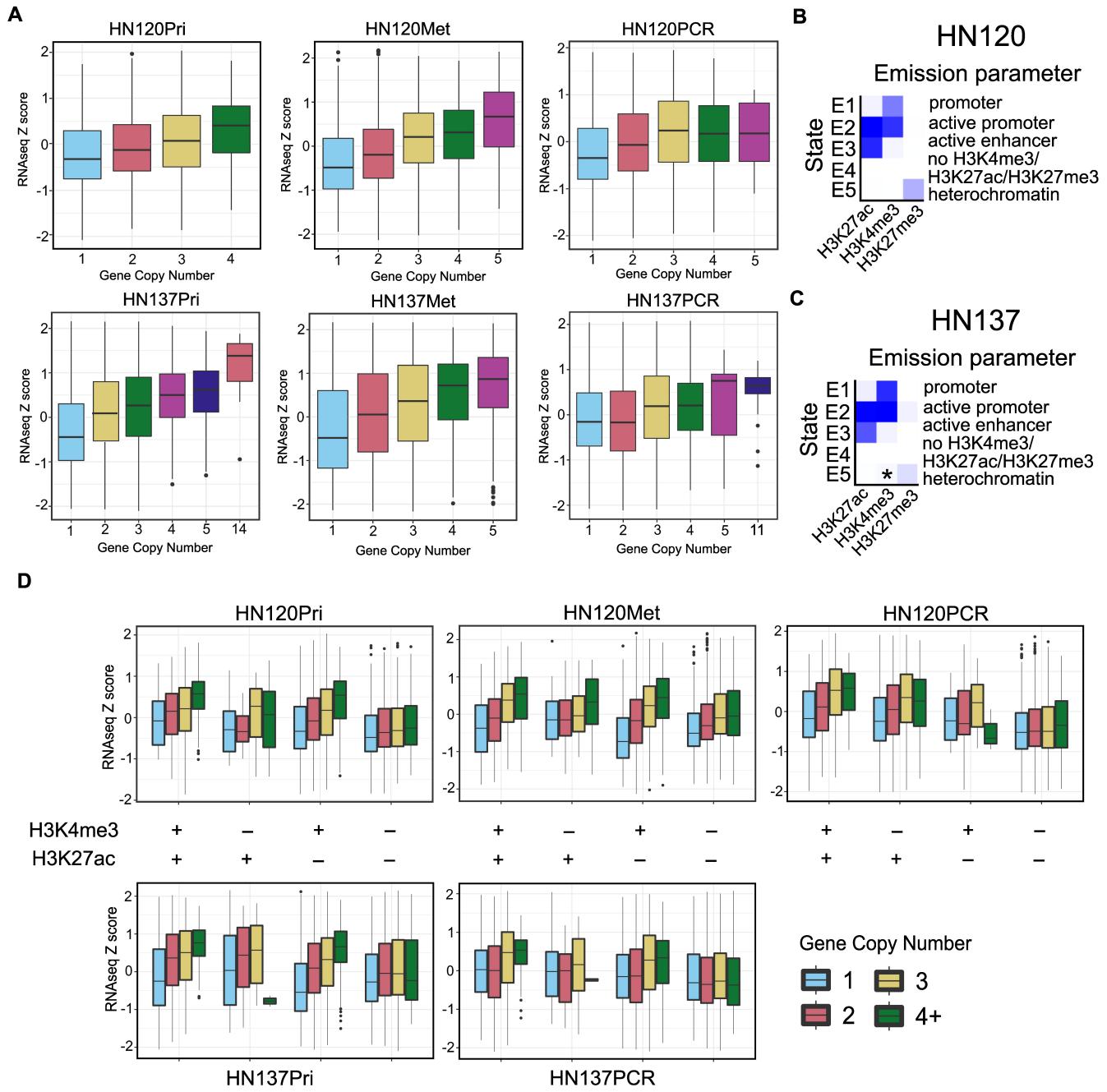
tumours. Data from Sharma et al. Nature Communications, 2018.

A**B****C**

Supplemental Fig. S5. Inference of transcription factor activity determining epithelial cell identity with H3K27ac and H3K4me3 signal. (A) H3K27ac CoveragePlots at the *KRT5* (left) and *KRT8/18* (right) loci for HN120 PDCs. **(B)** H3K4me3 CoveragePlots for HN120 and HN137 PDCs at the *KRT5* (left) and the *KRT8/18* (right) loci. **(C)** Motif activity UMAPs depicting the TF activities of TP63, KLF5, TEAD4 and FOSL2 at single cell level.

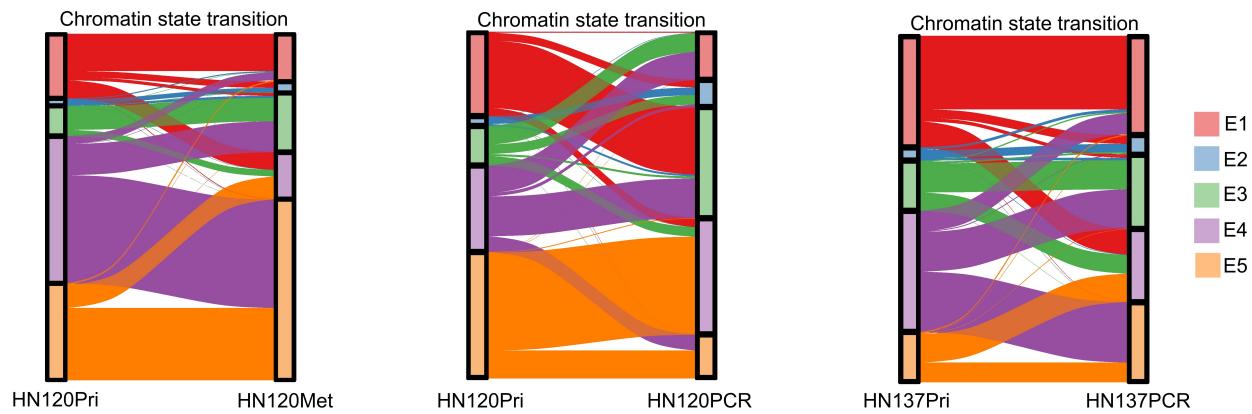
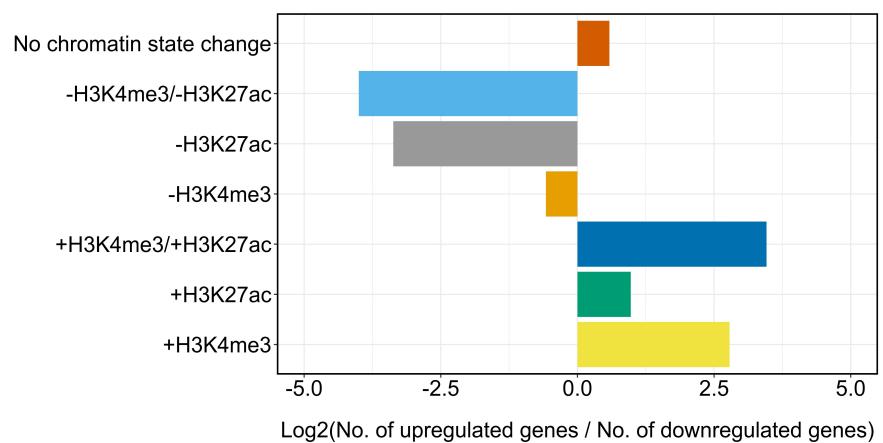
A**B**

Supplemental Fig. S6. Genomic regions with copy number amplifications exhibit intense read density with snCUT&RUN. (A) *EGFR* locus has a higher read density in HN137Met cells than HN137Pri cells, suggesting further amplification of this region might drive primary cancer cells towards metastasis. **(B)** Read density at Chr18p11.31 is significantly higher in both HN137Pri and HN137Met cells compared to the rest of the genome.

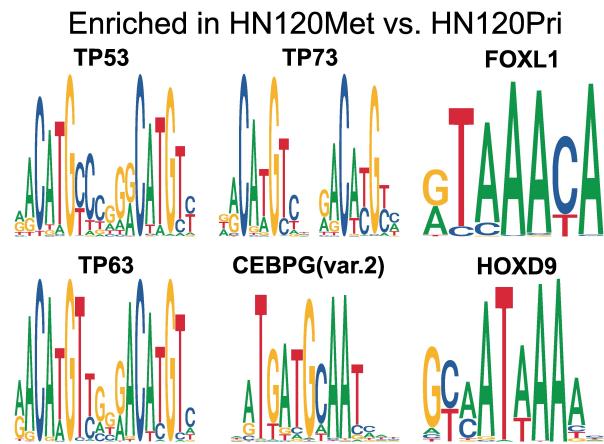
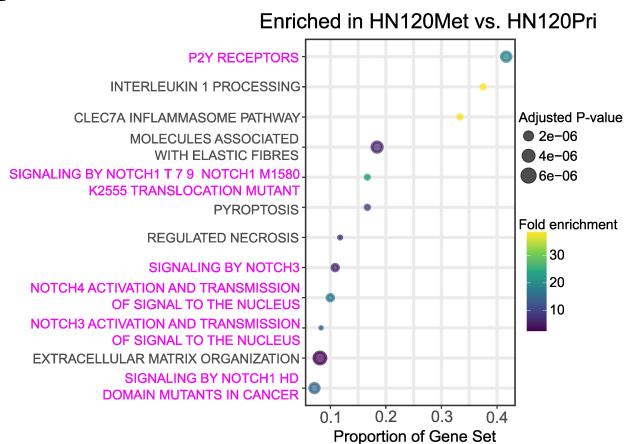
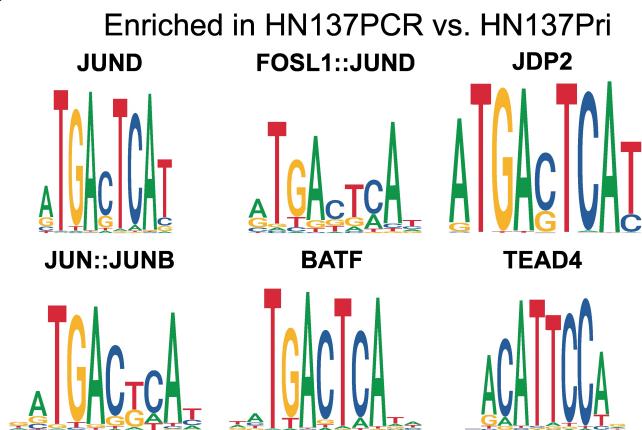
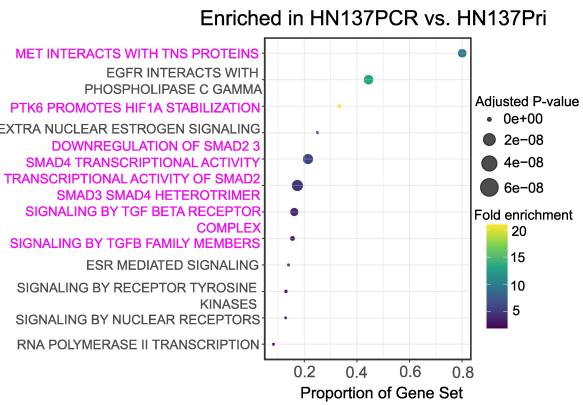
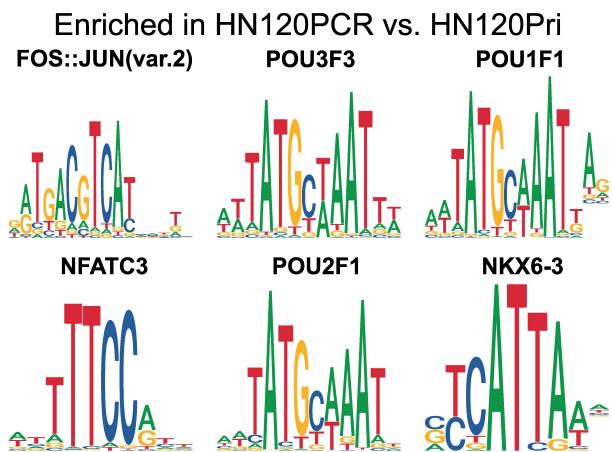
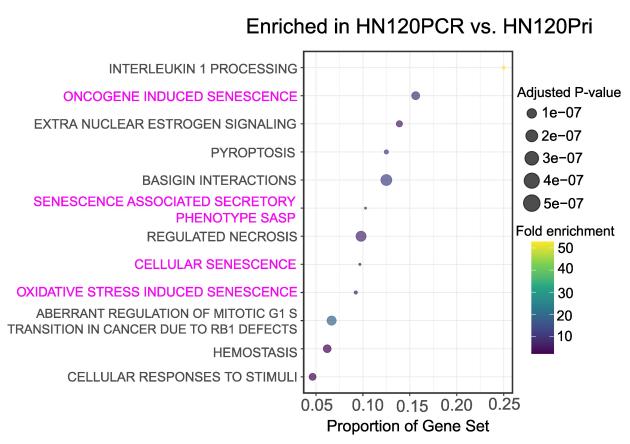


Supplemental Fig. S7. Correlation between gene CN, chromatin state, and gene expression (A)

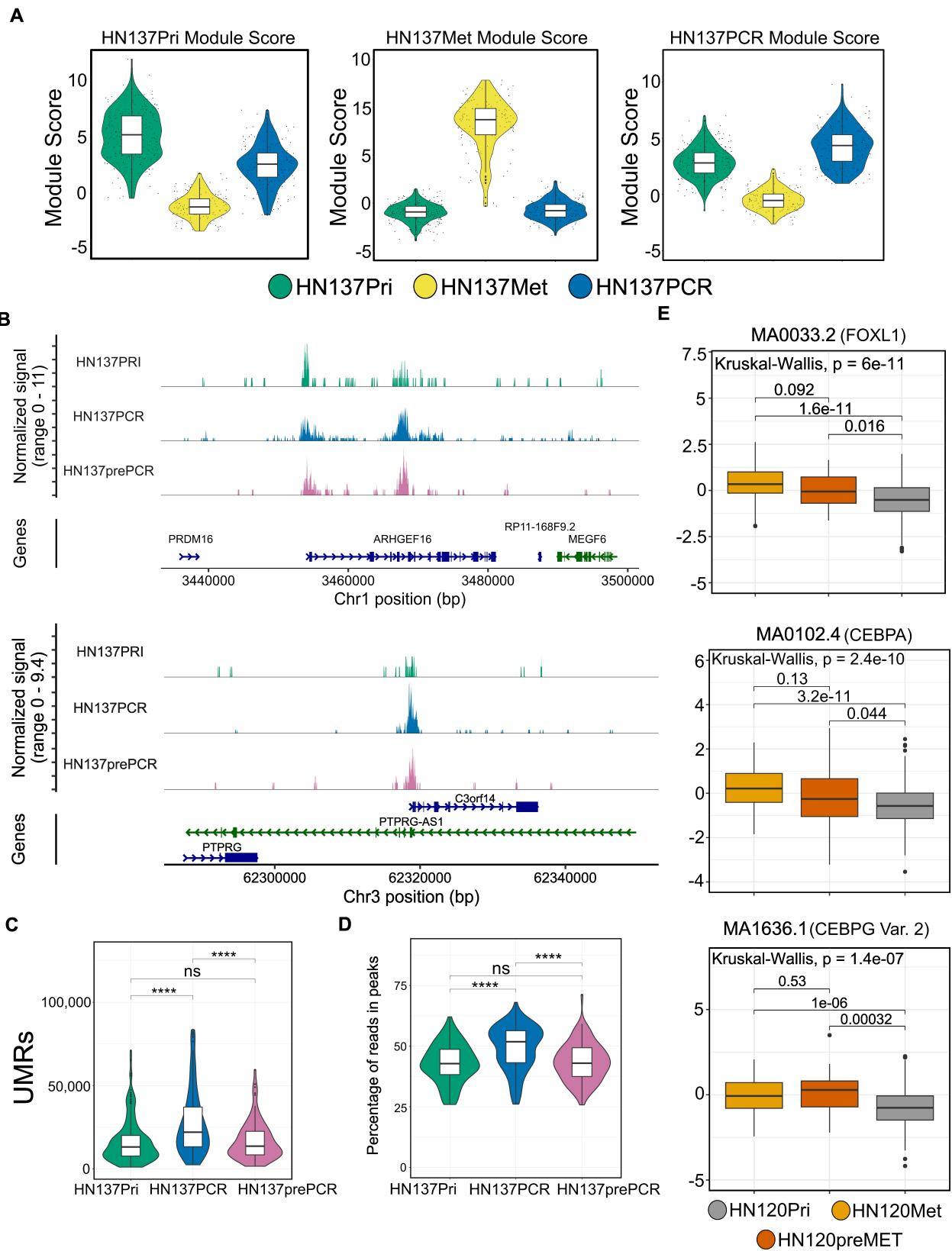
Boxplots of RNAseq Z-scores against gene CN of the six PDCs. **(B)** ChromHMM results showing chromatin state combinations based on HN120 samples alone. **(C)** ChromHMM results showing chromatin state combinations based on HN137 samples alone. The weak bivalent H3K4me3/H3K27me3 signal is annotated with an asterisk. **(D)** Boxplots correlating RNAseq Z-scores against H3K27ac/H3K4me3 activity in HN120Pri, HN120Met, HN120PCR, HN137Pri and HN137PCR.

A**B**

Supplemental Fig. S8. Chromatin state transitions between primary tumour and progressed HNSCC. **(A)** Alluvial plot depicting the chromatin state changes between HN120Pri and HN120Met PDC (left), HN120Pri > HN120PCR transition (middle) and HN137Pri > HN137PCR transition (right). E1 (red): regions with H3K4me3 only. E2 (blue): regions with both H3K4me3 and H3K27ac. E3 (green): regions with H3K27ac only. E4 (purple): unmodified regions. E5 (orange): regions with H3K27me3 only. **(B)** Log2 ratio of the number of upregulated ($\text{Log2 FC} > 1, p < 0.001$) genes to the number of downregulated ($\text{Log2 FC} < -0.5, p < 0.001$) genes per chromatin state change during metastatic progression of patient HN137 (HN137Pri > HN137Met).

A**B****C****D****E****F**

Supplemental Fig. S9. Transcription factor (TF) activity related to stemness is correlated with HNSCC progression. **(A)** Top six enriched TF motifs in HN120Met vs. HN120Pri. **(B)** Top 12 enriched REACTOME pathways in HN120Met vs. HN120Pri. **(C)** Top six enriched TF motifs in HN137PCR vs. HN137Pri. **(D)** Top 12 enriched REACTOME pathways in HN137PCR vs. HN137Pri. **(E)** Top six enriched TF motifs in HN120PCR vs. HN120Pri. **(F)** Top 12 enriched REACTOME pathways in HN120PCR vs. HN120Pri.



Supplemental Fig. S10. HN137prePCR and HN120preMET analysis. (A) PDC-specific HN137 H3K27ac module scores. Each module consists of the top 50 peaks for a particular PDC, and each module score was back calculated for each single-cell using Signac's AddChromatinModule function. (B) CoveragePlots of the *ARHGEF16* and *PTPRG-AS1* loci showing higher H3K27ac localization in HN137prePCR compared to remaining HN137Pri cells. (C) UMR count for HN137Pri, HN137PCR and HN137prePCR cells. (D) FRiP count for HN137Pri, HN137PCR and HN137prePCR cells. (E) ChromVAR scores of the activity of several TFs enriched in HN120Met vs. HN120Pri, visualized after the subset of HN120Pri to HN120preMET and remaining HN120Pri cells.