

SUPPLEMENTAL FIGURES

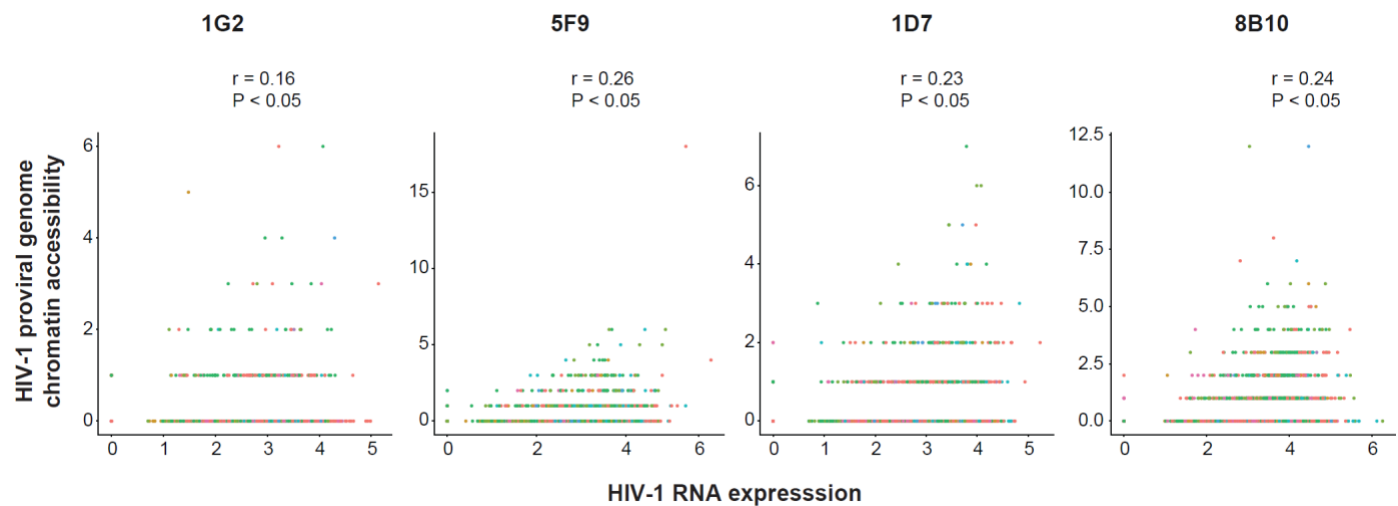


Figure S1. Correlations between HIV-1 RNA expression levels and HIV-1 genome chromatin

accessibility. Correlation coefficient (r) and P values were determined by Pearson correlation in each cell line across all cells in that line.

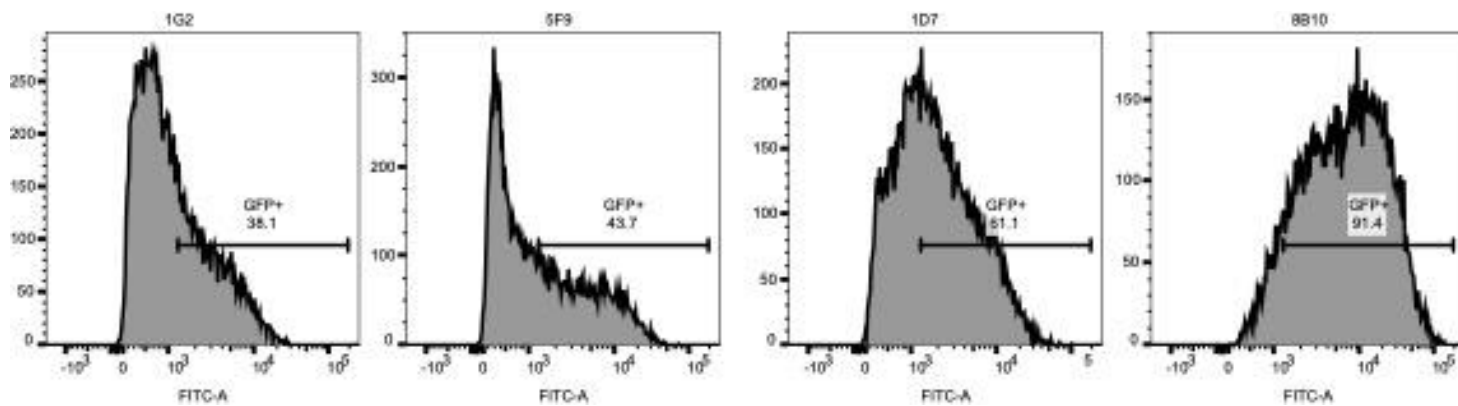


Figure S2. HIV-1 GFP expression is heterogenous across cell line clones. FACS plots showing the GFP+ of cell line clones, ordered from least to most chromatin accessibility.

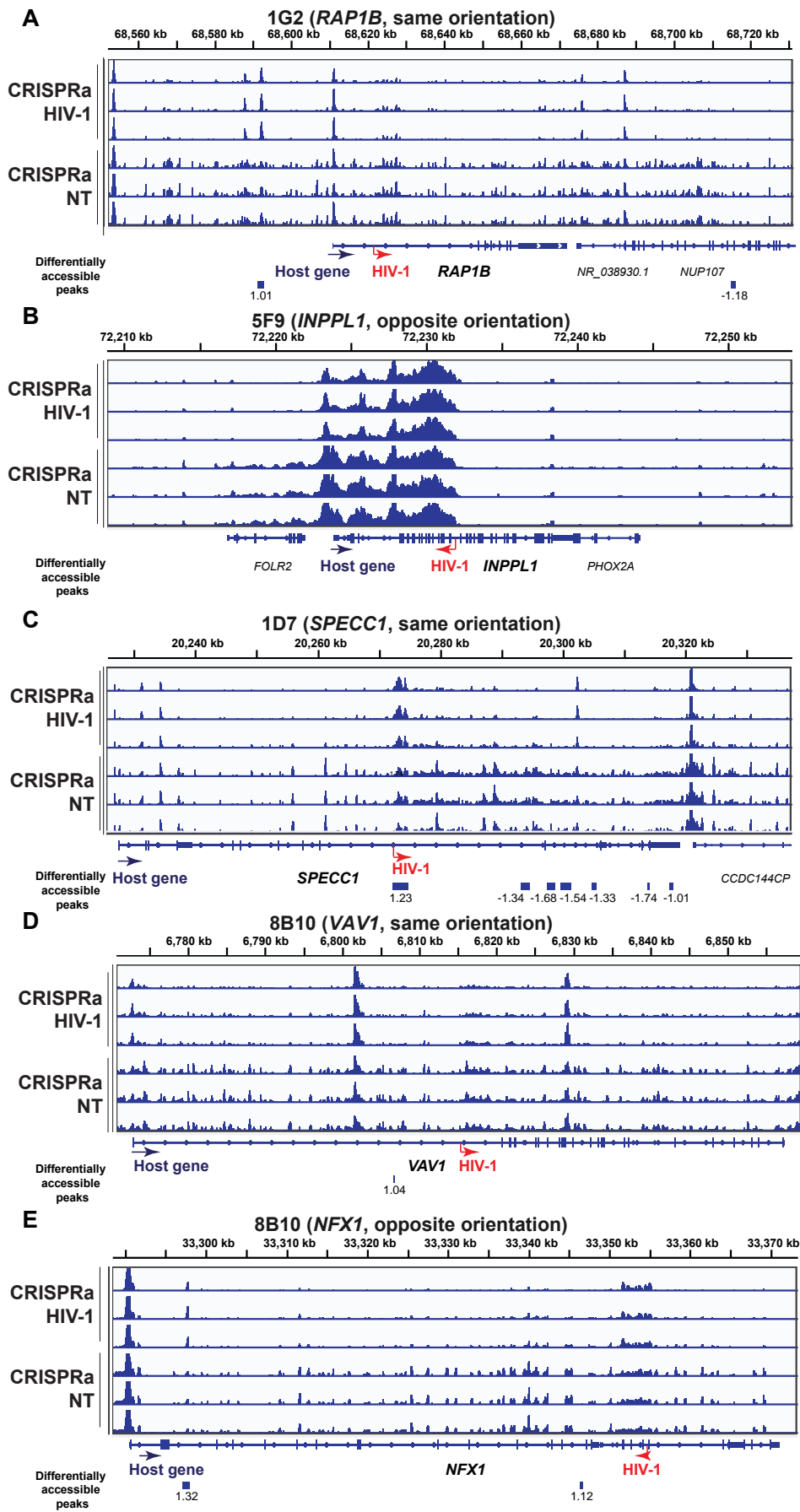


Figure S3. Impact of HIV-1-targeted CRISPR activation across cell lines. Genomic tracks and differentially accessible regions in CRISPRa expressing cell lines with an HIV-1-targeting guide RNA or a nontargeting guide RNA in (A) 1G2, (B) 5F9, (C) 1D7, or (D-E) 8B10. Significance determined by DESeq2 Tracks are CPM normalized and scaled to the fraction of reads in peaks, a range of 0-3 is shown on each track. Each peak is annotated with its Log2 fold change relative to the nontargeting control.

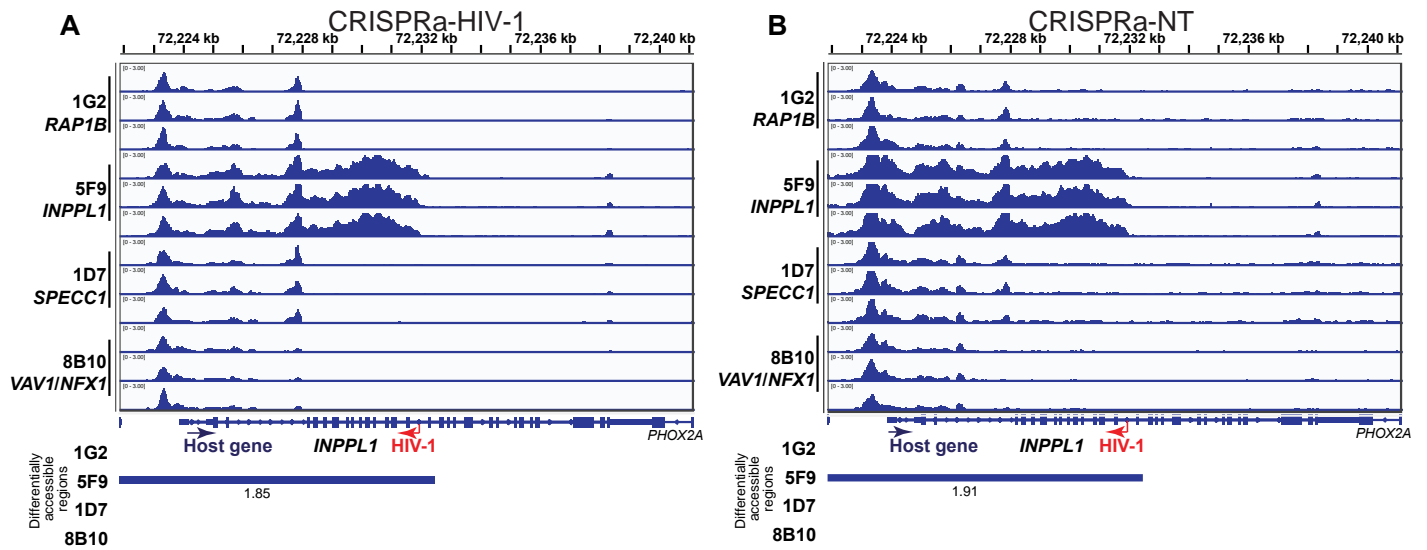


Figure S4. HIV-1-targetting CRISPRa fails to increase HIV-1-associated host chromatin accessibility.

The chromatin accessibility and significantly differentially accessible regions across cell lines at the *INPPL1* gene in CRISPRa-transduced cell lines with an (A) HIV-1-targetting guide RNA or (B) a nontargeting guide RNA.

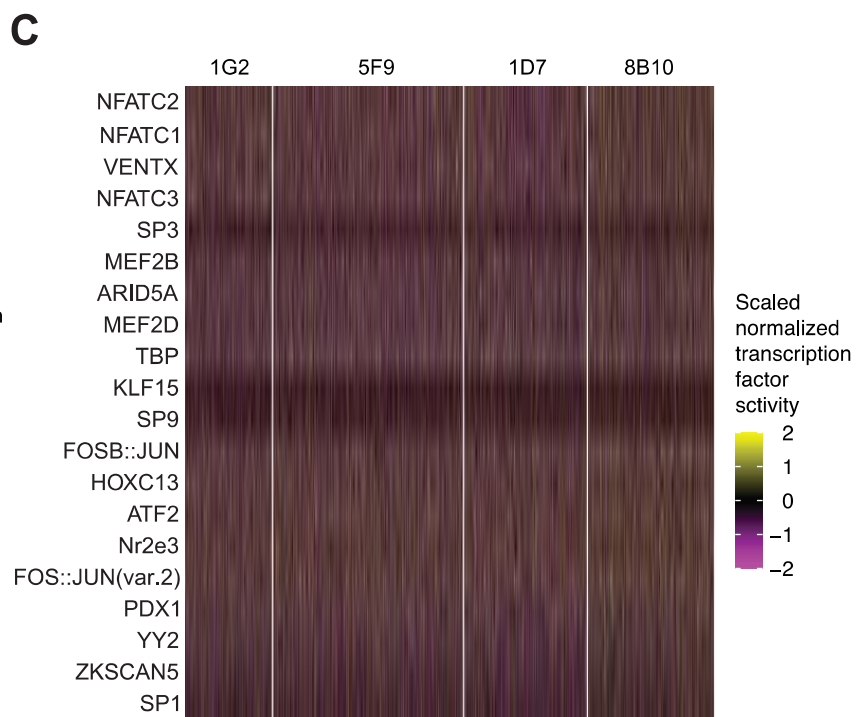
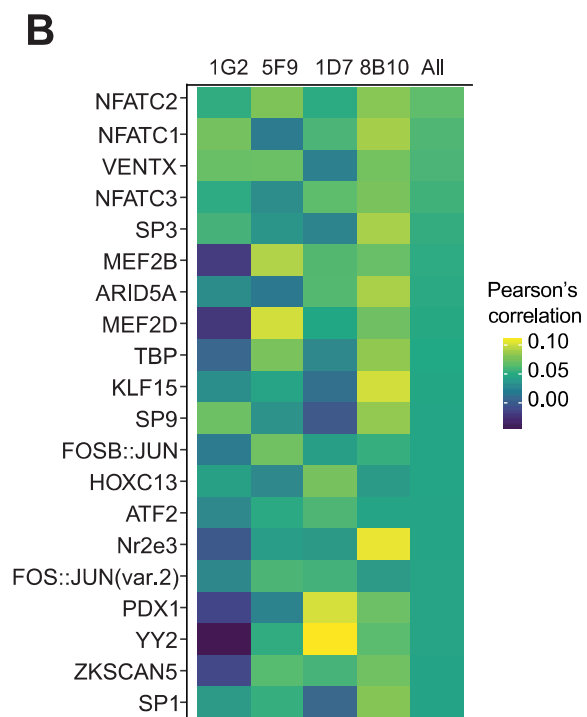
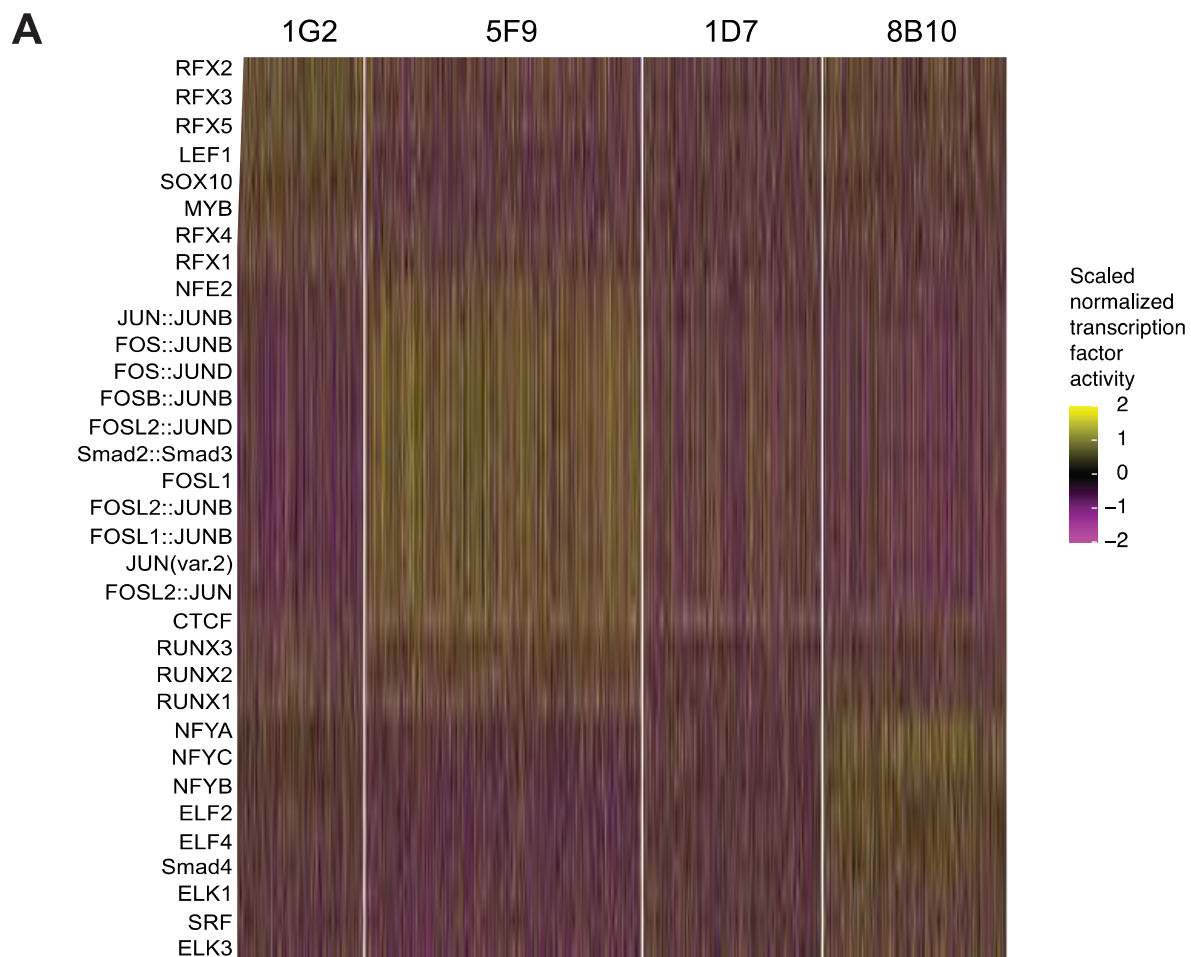
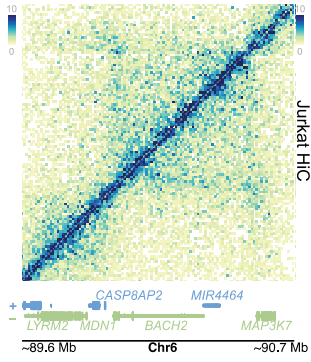


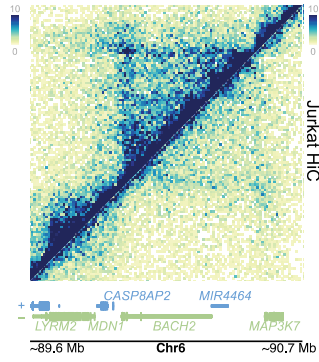
Figure S5. Correlations between transcription factor activity and HIV-1 chromatin accessibility differ in cell line clones having different HIV-1 integration sites independent of clone-clone differences. HIV-1 chromatin accessibility was measured by DOGMA-Seq. Transcription factor activity was measured by ChromVar. (A) Transcription factor activity of the top 10 positively enriched transcription factors in each cell line. (B) Heatmap of the Pearson correlation between transcription factor activities and HIV-1 chromatin accessibility. Pearson correlation calculated using R. Top 20 motifs by average correlation across cell lines shown, ordered by decreasing aggregate average correlation. (C) Transcription activity of the same 20 high correlation motifs across cell lines.

Hi-C (*BACH2* Locus)

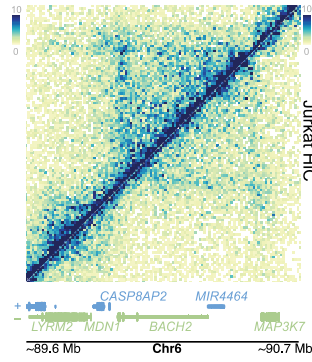
A 1G2 (*RAP1B*, same orientation)



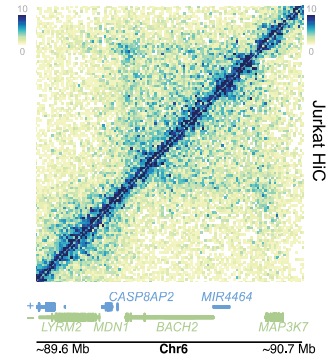
B 5F9 (*INPPL1*, opposite orientation)



C 1D7 (*SPECC1*, same orientation)
Replicate 1

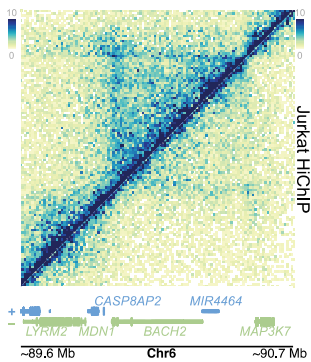


Replicate 2

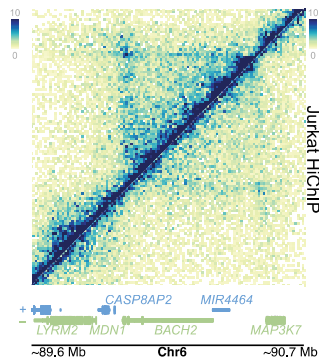


H3K27ac HiChIP (*BACH2* Locus)

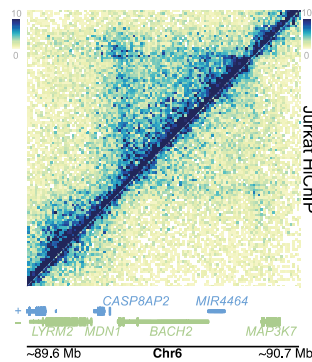
D 1G2 (*RAP1B*, same orientation)



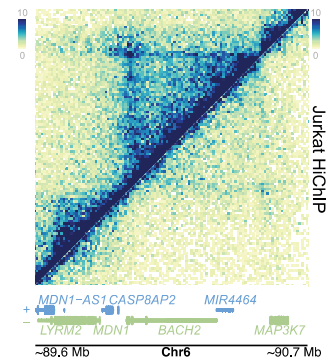
E 5F9 (*INPPL1*, opposite orientation)



F 1D7 (*SPECC1*, same orientation)
Replicate 1

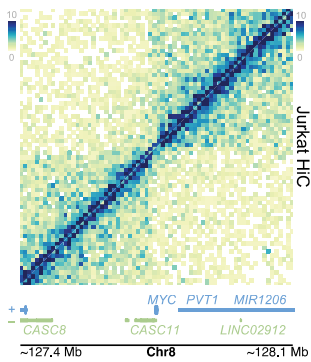


Replicate 2

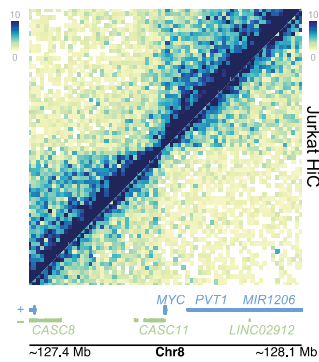


Hi-C (*MYC* Locus)

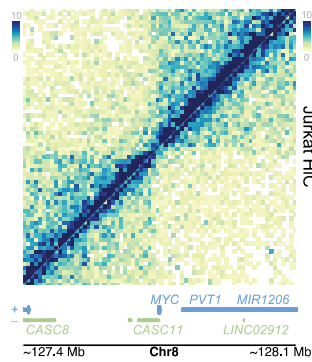
G 1G2 (*RAP1B*, same orientation)



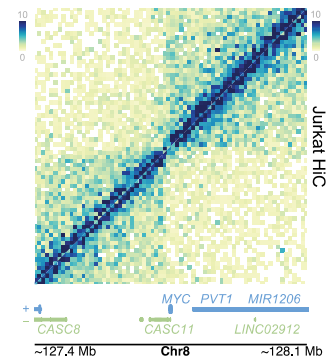
H 5F9 (*INPPL1*, opposite orientation)



I 1D7 (*SPECC1*, same orientation)
Replicate 1

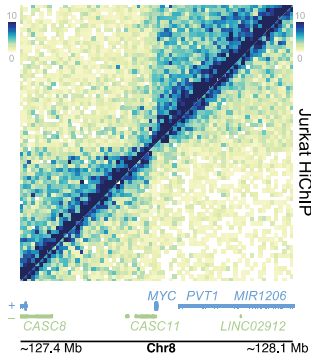


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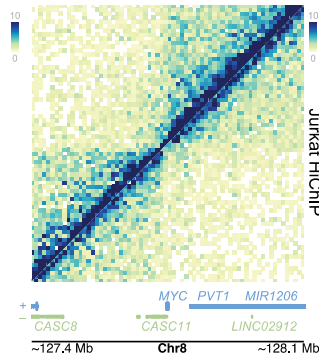


H3K27ac HiChIP (*MYC* Locus)

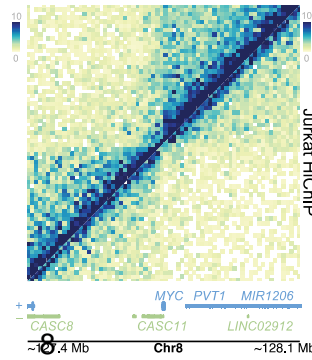
J 1G2 (*RAP1B*, same orientation)



K 5F9 (*INPPL1*, opposite orientation)



L 1D7 (*SPECC1*, same orientation)
Replicate 1



Replicate 2

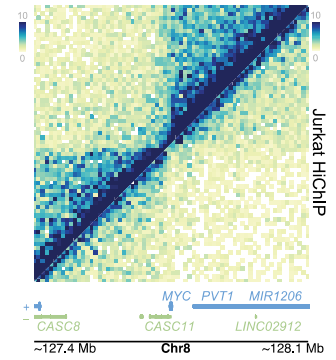


Figure S6. Impact of HIV-1 on genome wide 3D chromatin conformation and H3K27ac enhancer loops at the heterologous sites. Heatmap shows Kight-Ruiz (KR) normalized signal at a 10 kb resolution surrounding BACH2 (A-F) and MYC (G-L) by Hi-C (A–C, G-I) and H3K27ac HiChIP (D–F, J-L).

