

Supplemental information

Targeted, programmable and precise tandem duplication in

mammalian genome

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Yao^{1*}

FANCF TD 50-bp

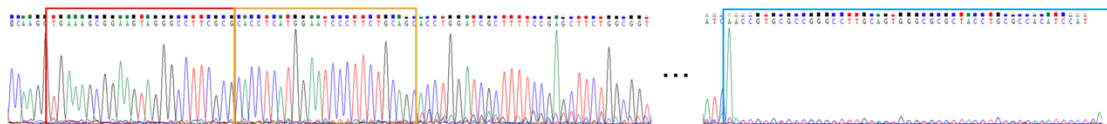
✓ Wild-type sequence:

PAM1 nick

CCAAGGTGAAAGCGGAAGTAGGGCCTTCGCGCACCTCATGGAATCCCTTCTGCAGCACCTGGATCGCTTTTC

Reference sequence:

CCAAGGTGAAAGCGGAAGTAGGGCCTTCGCGACCTCATGGAATCCCTTCTGCAGCTGAAAGCGGAAGTAGGGCCTTCGCGACCTCATGGAATCCCTTCTGCAGCACCTGGATCGCTTTTC



Supplemental Fig. S1 Representative Sanger sequencing results of TD-PE mediated 50-bp tandem duplication at *FANCF* site. PCR amplicons containing targeted tandem duplication was purified and directly subjected to Sanger sequencing. Wild-type sequence and TD reference sequence were shown on top of the sequencing chromatogram. PAM1 and PAM2 of the two pegRNAs were shown in red and yellow respectively. Red arrows indicated the positions of the nicks. The sequences in-between two nicks are the duplicated sequences, with the upstream end shown in red and the downstream end shown in yellow. The peaks in the sequencing chromatogram corresponding to each end were boxed in red and yellow respectively. The blue box marked the fragment that is located at the end of the double peaks. The length of the fragment is the same as that of the duplicated region, and the sequence corresponds to the 3' end of the amplicons that contain TDs.

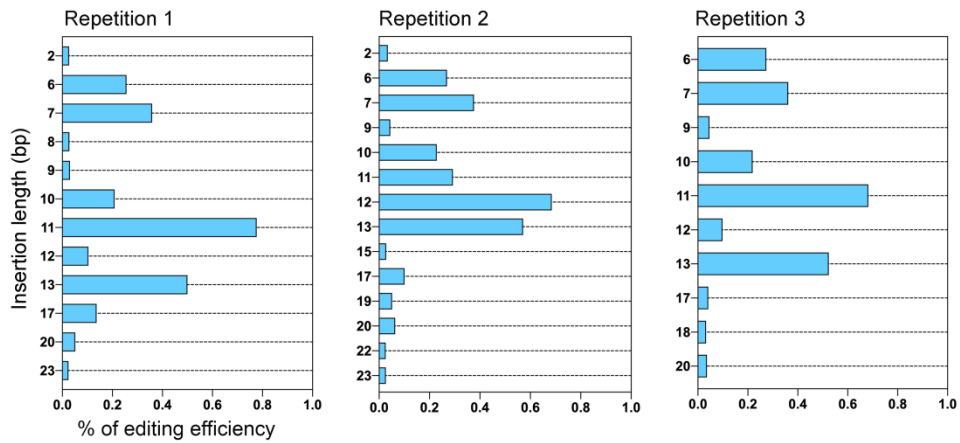
[illegible]

Supplemental Fig. S2 HTS sequences showing the editing result of TD-PE at the *FANCF* site. The fragments containing tandem duplications were amplified by HTS primers and subjected for HTS analysis. Detailed analysis of the undesired TDs revealed that the majority of deletions and insertions occurred at the boundaries of the duplications. The blue curved arrows pointed to the next copy of the tandem duplication. Deletion, insertion and base substitution were indicated by black dash, red box and green letters respectively. The insertion sequence of the red box was consistent with its 5' upstream sequence. Note that to provide an overview of the TDs, we copied the sequences of 2×TD from Fig. 2E to this section.

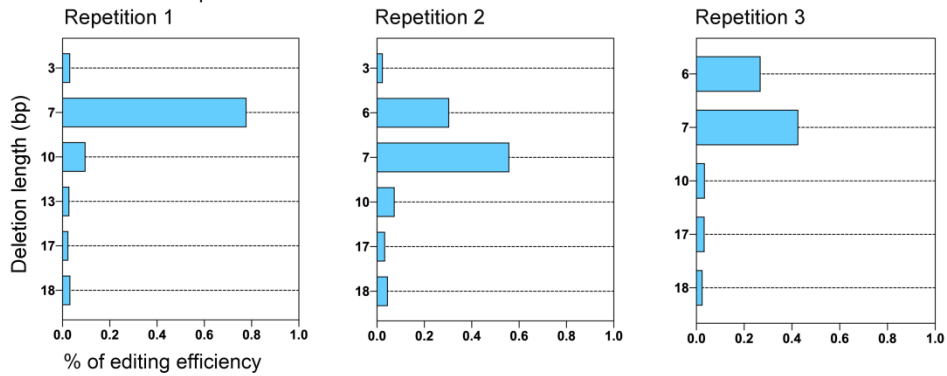
	<div> <div>PAM1</div> <div>nick</div> <div>PAM2</div> </div>	
Wild-type sequence :	<div> <div> <div>AACCGGAGGACAAAGTACAAACGGGCAGAAAGCTGGAGGAGGAAGGGCCTGAGTCCGAGCAGAAGAA</div> <div>GAA</div> <div>GGGCTCCCATCA</div> </div> </div>	<div> <div>----</div> Cleavage <div>-</div> Deletion <div>□</div> Insertion </div>
2 × TD (Accurate) :	<div> <div> <div>AACCGGAGGACAAAGTACAAACGGGCAGAAAGCTGGAGGAGGAAGGGCCTGAGTCCGAGCAGAAGAA</div> <div>GACAAAGTACAAACGGCAGAAAGCTGGAGGAGGAAGGGCCTGAGTCCGAGCAGAAGAA</div> <div>GAA</div> <div>GGGCTCCCATCA</div> </div> </div>	reads / total reads 64435 / 389912 (16.53%)
2 × TD with deletion :	<div> <div> <div>AACCGGAGGACAAAGTACAAACGGGCAGAAAGCTGGAGGAGGAAGGGCCTGAGTCCGAGCAGAAGAA</div> <div>GACAAAGTACAAACGGCAGAAAGCTGGAGGAGGAAGGGCCTGAGTCCGAGCAGAAGAA</div> <div>GAA</div> <div>GGGCTCCCATCA</div> </div> </div>	256 / 389912 (0.066%)
2 × TD with insertion :	<div> <div> <div>AACCGGAGGACAAAGTACAAACGGGCAGAAAGCTGGAGGAGGAAGGGCCTGAGTCCGAGCAGAAGAA</div> <div>GACAAAGTACAAACGGCAGAAAGCTGGAGGAGGAAGGGCCTGAGTCCGAGCAGAAGAA</div> <div>GAA</div> <div>GGGCTCCCATCA</div> </div> </div>	95 / 389912 (0.024%)
3 × TD (Accurate) :	<div> <div> <div>AACCGGAGGACAAAGTACAAACGGGCAGAAAGCTGGAGGAGGAAGGGCCTGAGTCCGAGCAGAAGAA</div> <div>GACAAAGTACAAACGGCAGAAAGCTGGAGGAGGAAGGGCCTGAGTCCGAGCAGAAGAA</div> <div>GAA</div> <div>GGGCTCCCATCA</div> </div> </div>	11889 / 389912 (3.05%)
4 × TD (Accurate) :	<div> <div> <div>AACCGGAGGACAAAGTACAAACGGGCAGAAAGCTGGAGGAGGAAGGGCCTGAGTCCGAGCAGAAGAA</div> <div>GACAAAGTACAAACGGCAGAAAGCTGGAGGAGGAAGGGCCTGAGTCCGAGCAGAAGAA</div> <div>GAA</div> <div>GGGCTCCCATCA</div> </div> </div>	404 / 389912 (0.10%)
5 × TD :	<div> <div> <div>AACCGGAGGACAAAGTACAAACGGGCAGAAAGCTGGAGGAGGAAGGGCCTGAGTCCGAGCAGAAGAA</div> <div>GACAAAGTACAAACGGCAGAAAGCTGGAGGAGGAAGGGCCTGAGTCCGAGCAGAAGAA</div> <div>GAA</div> <div>GGGCTCCCATCA</div> </div> </div>	113 / 389912 (0.029%)
Deletion without edit :	<div> <div> <div>AACCGGAGGACAAAGTACAAACGGGCAGAAAGCTGGAGGAGGAAGGGCCTGAGTCCGAGCAGAAGAA</div> <div>GAA</div> <div>GGGCTCCCATCA</div> </div> </div>	118 / 389912 (0.030%)

Supplemental Fig. S3 HTS sequences showing the editing result of TD-PE at the *EMX1* site. The fragments containing tandem duplication were amplified by HTS primers, purified and then subjected for HTS. The blue curved arrows pointed to the next copy of the tandem duplication. Deletion, insertion and base substitution were indicated by black dash, red box or green letters respectively. The insertion sequence of the red box was consistent with its 5' upstream sequence.

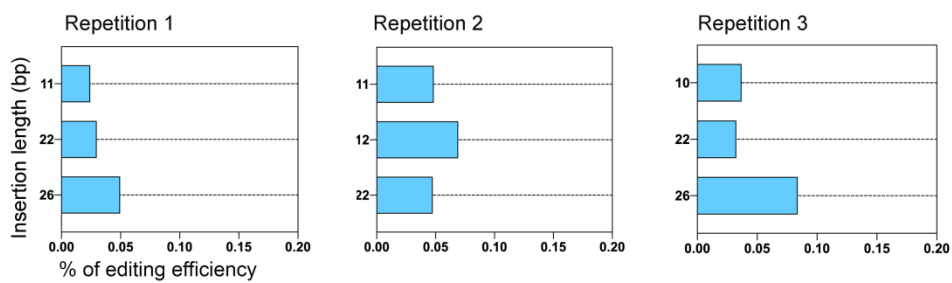
FANCF TD 50-bp 2×TD insertion



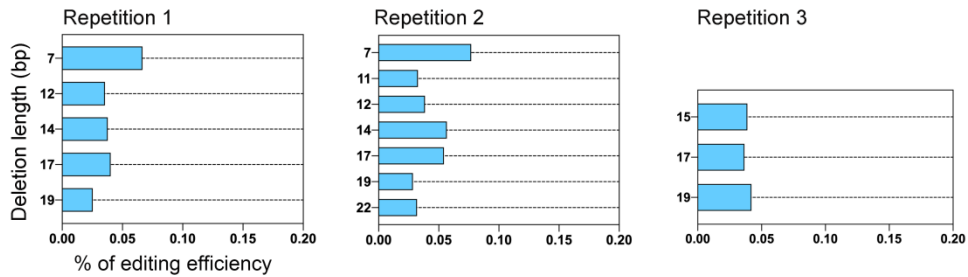
FANCF TD 50-bp 2×TD deletion



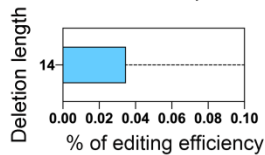
FANCF TD 50-bp 3×TD insertion



FANCF TD 50-bp 3×TD deletion

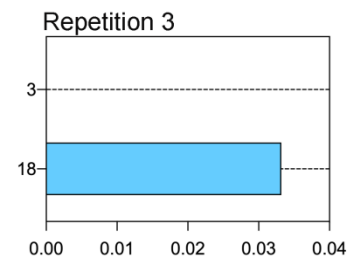
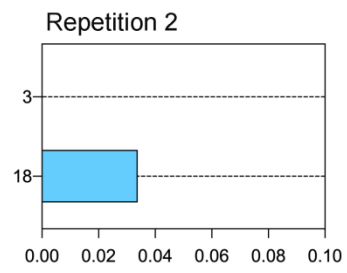
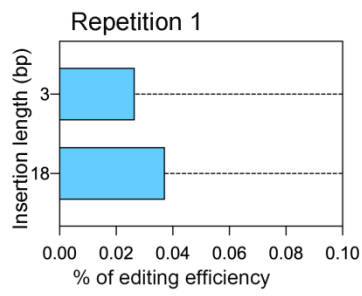


FANCF TD 50-bp 4×TD deletion

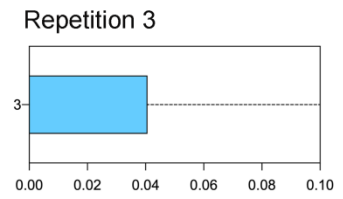
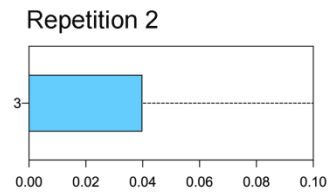
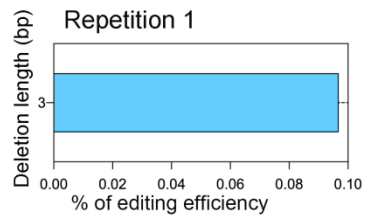


Supplemental Fig. S4 Distribution of the insertion or deletion lengths at *FANCF* site. HTS analysis of the frequencies of undesired edits induced by TD-PE at *FANCF* site (TD 50-bp). The experiment was repeated three times, and the indel data from each repetition were presented separately. Note that the deletion in 4×TD was detectable in only one out of three repetitions.

EMX1 TD 57-bp 2×TD insertion



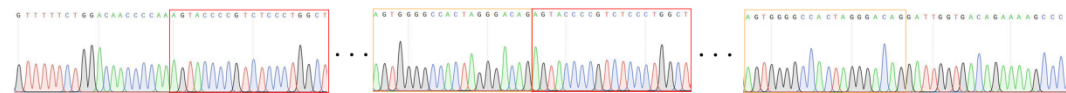
EMX1 TD 57-bp 2×TD deletion



Supplemental Fig. S5 Distribution of the insertion or deletion lengths at *EMXI* site. HTS analysis of the frequencies of undesired edits induced by TD-PE at *EMXI* site (TD 57-bp). The experiment was repeated three times, and the indel data from each repetition were presented separately.

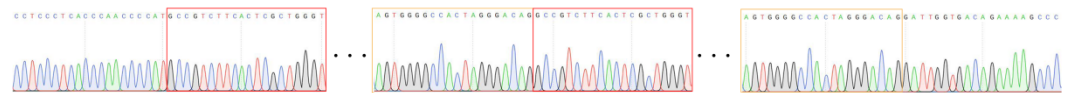
AAVS1 TD 376-bp 2×TD

TD reference sequence: GTTTTCTGGACAACCCCAAAGTACCCCGTCTCCCTGGCT...AGTGGGGCCACTAGGGACAGAGTACCCCGTCTCCCTGGCT...AGTGGGGCCACTAGGGACAGGATTGGTGACAGAAAAGCCC



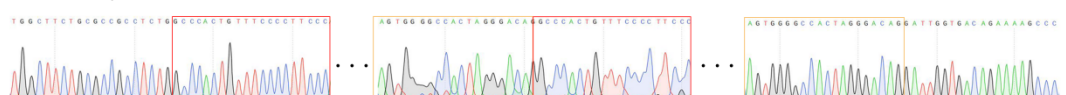
AAVS1 TD 532-bp 2×TD

TD reference sequence: CCTCCCTCACCCAACCCCATGCCGTCTTCACTCGCTGGGT...AGTGGGGCCACTAGGGACAGGCCGTCTTCACTCGCTGGGT...AGTGGGGCCACTAGGGACAGGATTGGTGACAGAAAAGCCC



AAVS1 TD 846-bp 2×TD

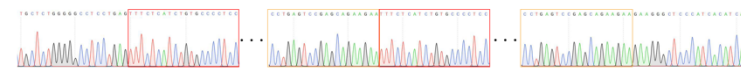
TD reference sequence: TGGCTTCTGCGCCGCCTCTGGCCACTGTTTCCCCTGCC...AGTGGGGCCACTAGGGACAGGCCACTGTTTCCCCTGCC...AGTGGGGCCACTAGGGACAGGATTGGTGACAGAAAAGCCC



Supplemental Fig. S6 Representative Sanger sequencing results of large fragment TDs at the *AAVS1* site (related to Fig. 3A). Representative Sanger sequencing results of TD containing amplicons derived from 3 targeted TDs at the *AAVS1* site. The amplicons containing targeted TDs were gel-purified and subjected directly to Sanger sequencing. TD reference sequence was shown on top of each sequencing chromatogram, with sequences corresponding to each end of the TD boxed in red and yellow respectively.

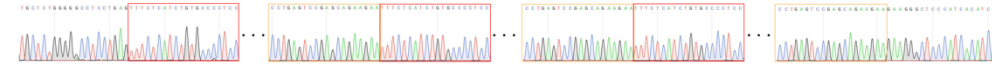
EMX1 TD 116-bp 2×TD

TD reference sequence: TGCTCTGGGGGCTCCTGAGTTTCTCATCTGTGCCCTCC--CCTGAGTCCGAGCAGAAGATTCTCATCTGTGCCCTCC--CCTGAGTCCGAGCAGAAGAAAGAGGGCTCCATCACATCA



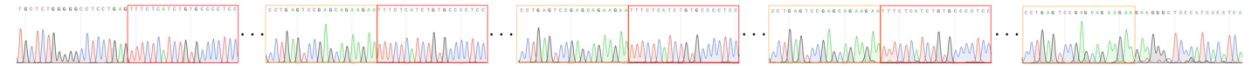
EMX1 TD 116-bp 3×TD

TD reference sequence: TGCTCTGGGGGCTCCTGAGTTTCTCATCTGTGCCCTCC--CCTGAGTCCGAGCAGAAGATTCTCATCTGTGCCCTCC--CCTGAGTCCGAGCAGAAGAAAGAGGGCTCCATCACATCA



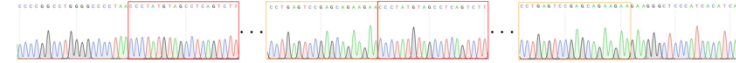
EMX1 TD 116-bp 4×TD

TD reference sequence: TGCTCTGGGGGCTCCTGAGTTTCTCATCTGTGCCCTCC--CCTGAGTCCGAGCAGAAGATTCTCATCTGTGCCCTCC--CCTGAGTCCGAGCAGAAGAAAGAGGGCTCCATCACATCA



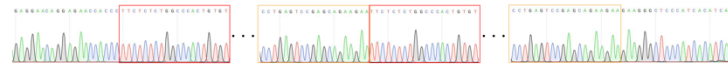
EMX1 TD 201-bp 2×TD

TD reference sequence: GCCCGGCTGGGGGCTTAACCTATGAGCTCAGTCTT--CCTGAGTCCGAGCAGAAGAAAGAGGGCTCCATCACATCA



EMX1 TD 307-bp 2×TD

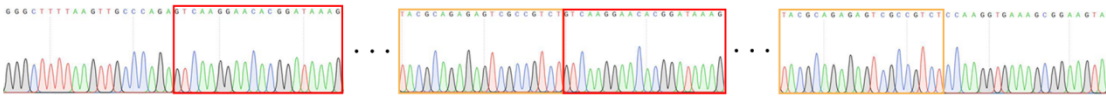
TD reference sequence: GAGGAACAGGAAACACCTTCTCTGTGCCCACTGT--CCTGAGTCCGAGCAGAAGATTCTCTGTGCCCACTGT--CCTGAGTCCGAGCAGAAGAAAGAGGGCTCCATCACATCA



Supplemental Fig. S7 Representative Sanger sequencing results of large fragment TDs at the *EMXI* site (related to Fig. 3A). Representative Sanger sequencing results of TD containing amplicons derived from 3 targeted TDs at *EMXI* site. The amplicons containing targeted TDs were gel-purified and subjected to Sanger sequencing. TD reference sequence was shown on top of each sequencing chromatogram, with sequences corresponding to each end of the TD boxed in red and yellow respectively. *EMXI* TD 116-bp 3×TD and 4×TD were the sequencing results of the bands marked red stars (*) at the *EMXI* site in Fig. 3A.

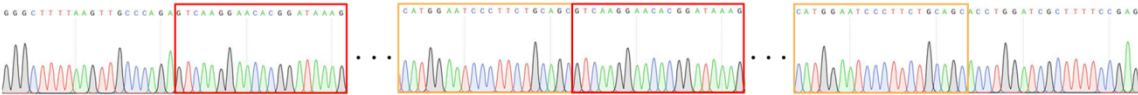
FANCF TD 115-bp 2×TD

TD reference sequence: GGGCTTTTAAAGTTGCCCAGAGTCAAGGAACACGGATAAAG...TACGCAGAGAGTCGCCGTCTGTCAAGGAACACGGATAAAG...TACGCAGAGAGTCGCCGTCTCCAAGGTGAAAGCGGAAGTA



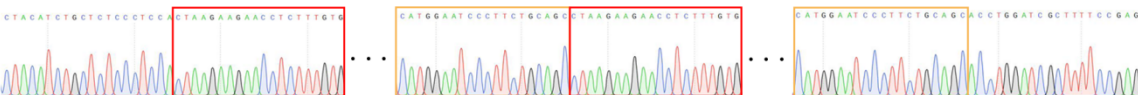
FANCF TD 171-bp 2×TD

TD reference sequence: GGGCTTTTAAAGTTGCCCAGAGTCAAGGAACACGGATAAAG...CATGGAATCCCTTCTGCAGCGTCAAGGAACACGGATAAAG...CATGGAATCCCTTCTGCAGCACCTGGATCGCTTTTCCGAG



FANCF TD 231-bp 2×TD

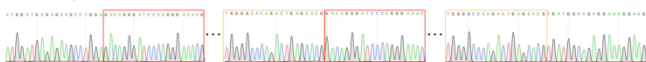
TD reference sequence: CTACATCTGCTCTCCCTCCACTAAGAAGAACCTCTTTGTG...CATGGAATCCCTTCTGCAGCGTCAAGAAGAACCTCTTTGTG...CATGGAATCCCTTCTGCAGCACCTGGATCGCTTTTCCGAG



Supplemental Fig. S8 Representative Sanger sequencing results of large fragment TDs at the *FANCF* site (related to Fig. 3A). Representative Sanger sequencing results of TD containing amplicons derived from 3 targeted TDs at *FANCF* site. The amplicons containing targeted TDs were gel-purified and subjected to Sanger sequencing. TD reference sequence was shown on top of each sequencing chromatogram, with sequences corresponding to each end of the TD boxed in red and yellow.

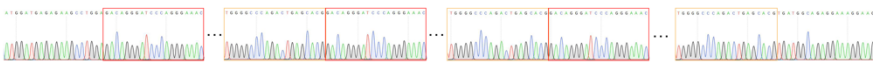
HEK3 TD 136-bp 2×TD

TD reference sequence: ATGGATGAGAGAAGCCTGGA**GACAGGGATCCAGGGAACG**~TGGGGCCAGACTGAGCAC**GACAGGGATCCAGGGAACG**~TGGGGCCAGACTGAGCAC**TGATGGCAGAGAAAGGAAG**



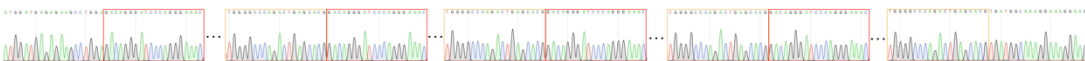
HEK3 TD 136-bp 3×TD

TD reference sequence: ATGGATGAGAGAAGCCTGGA**GACAGGGATCCAGGGAACG**~TGGGGCCAGACTGAGCAC**GACAGGGATCCAGGGAACG**~TGGGGCCAGACTGAGCAC**GACAGGGATCCAGGGAACG**~TGGGGCCAGACTGAGCAC**TGATGGCAGAGAAAGGAAG**



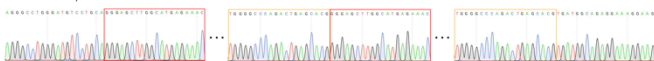
HEK3 TD 136-bp 4×TD

TD reference sequence: ATGGATGAGAGAAGCCTGGA**GACAGGGATCCAGGGAACG**~TGGGGCCAGACTGAGCAC**GACAGGGATCCAGGGAACG**~TGGGGCCAGACTGAGCAC**GACAGGGATCCAGGGAACG**~TGGGGCCAGACTGAGCAC**GACAGGGATCCAGGGAACG**~TGGGGCCAGACTGAGCAC**TGATGGCAGAGAAAGGAAG**



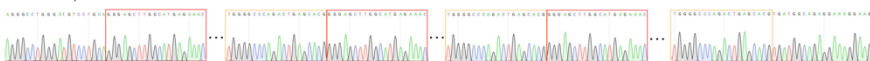
HEK3 TD 222-bp 2×TD

TD reference sequence: AGGGCTGGGATGTCCTGCA**GGGAGCTTGGCATGAGAAAC**~TGGGGCCAGACTGAGCAC**GGGAGCTTGGCATGAGAAAC**~TGGGGCCAGACTGAGCAC**TGATGGCAGAGAAAGGAAG**



HEK3 TD 222-bp 3×TD

TD reference sequence: AGGGCTGGGATGTCCTGCA**GGGAGCTTGGCATGAGAAAC**~TGGGGCCAGACTGAGCAC**GGGAGCTTGGCATGAGAAAC**~TGGGGCCAGACTGAGCAC**GGGAGCTTGGCATGAGAAAC**~TGGGGCCAGACTGAGCAC**TGATGGCAGAGAAAGGAAG**



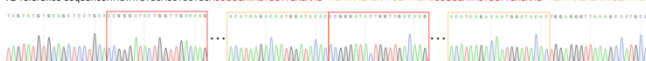
HEK3 TD 397-bp 2×TD

TD reference sequence: AGGAGGGAGCCTGGCCCTTCA**GGTCTGAGCTCAACAGAGGA**~ACATAAGACAATGGATAC**GGTCTGAGCTCAACAGAGGA**~ACATAAGACAATGGATAC**TGGAGGGTTAAGCCATGCC**



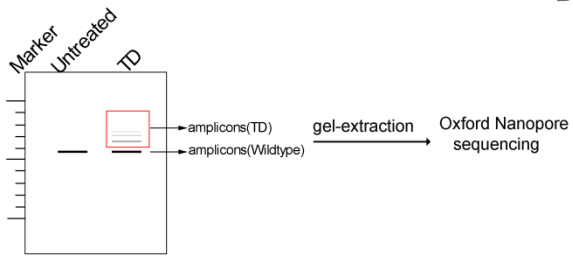
HEK3 TD 499-bp 2×TD

TD reference sequence: TAGTATGTGCAGCTCTGCA**CCGGGATCTGGTTGACAAG**~ACATAAGACAATGGATAC**CCGGGATCTGGTTGACAAG**~ACATAAGACAATGGATAC**TGGAGGGTTAAGCCATGCC**

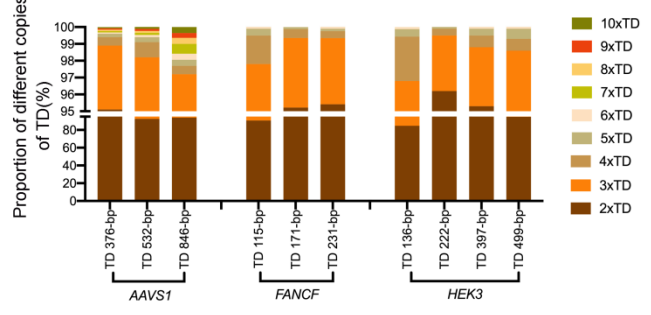


Supplemental Fig. S9 Representative Sanger sequencing results of large fragment TDs at the *HEK3* site (related to Fig. 3A). Representative Sanger sequencing results of TD containing amplicons derived from 4 targeted TDs at *HEK3* site. The amplicons containing targeted TDs were gel-purified and subjected to Sanger sequencing. TD reference sequence was shown on top of each sequencing chromatogram, with sequences corresponding to each end of the TD boxed in red and yellow respectively. *HEK3* TD 136-bp 3×TD and 4×TD and *HEK3* TD 222-bp 3×TD were the sequencing results of the bands marked by red stars (*) at the *HEK3* site in Fig. 3A.

A

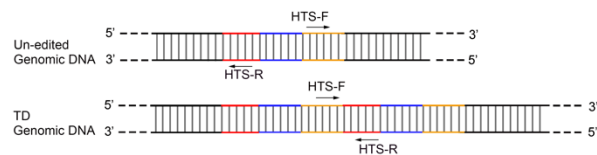


B



Supplemental Fig. S10 Proportions of different copies of TDs were analyzed by Oxford Nanopore sequencing. (A) Diagram showing the procedure to prepare Oxford Nanopore sequencing Samples. The amplicons with size equivalent to or larger than $2\times$ TD were collected and gel extracted for Nanopore sequencing. (B) The proportions of each type of TDs were analyzed.

A



B

AAVS1 TD 376-bp

TD reference sequence:TCCCTCCACCCACAGTGGGGCCACTAGGGACAGGTACCCCGTCTCCCTGGCTTTAGCCACCTCTCCA reads / total reads

2 × TD (Accurate) :TCCCTCCACCCACAGTGGGGCCACTAGGGACAGGTACCCCGTCTCCCTGGCTTTAGCCACCTCTCCA 321892/604583 (99.98%)

2 × TD with insertion :TCCCTCCACCCACAGTGGGGCCACTAGGGACAGGTACCCCGTCTCCCTGGCTTTAGCCACCTCTCCA**CTCTCCCTGGCTTTAGCCACCTCTCCA**CTCTGGCT 72/604583 (0.02%)

32 bp

AAVS1 TD 532-bp

TD reference sequence:TCCCTCCACCCACAGTGGGGCCACTAGGGACAGGCGCTTCACTCGCTGGGTCCCTTTCTCTCCCTCTCTCTG reads / total reads

2 × TD (Accurate) :TCCCTCCACCCACAGTGGGGCCACTAGGGACAGGCGCTTCACTCGCTGGGTCCCTTTCTCTCCCTCTCTCTG 448929/447308 (99.92%)

2 × TD with insertion :TTATCTGTCCCTCCACCCACAGTGGGGCCACTAGGGACAGGCGCTTCACTCGCTGGGTCCCTTTCTCTCTCTCTG**ATCTGTCCCTCCACCCACAGTGGGGCCACTAGGGACAGGCGCTTCACTCGCTGGGTCCCTTTCTCTCTCTG**GGG 273/447308 (0.06%)

82 bp

106/447308 (0.02%)

AAVS1 TD 846-bp

TD reference sequence:TTATCTGTCCCTCCACCCACAGTGGGGCCACTAGGGACAGGCGCTTCACTCGCTGGGTCCCTTTCTCTCTCTG reads / total reads

2 × TD (Accurate) :TTATCTGTCCCTCCACCCACAGTGGGGCCACTAGGGACAGGCGCTTCACTCGCTGGGTCCCTTTCTCTCTCTG 604109/604583 (99.92%)

2 × TD with insertion :TTATCTGTCCCTCCACCCACAGTGGGGCCACTAGGGACAGGCGCTTCACTCGCTGGGTCCCTTTCTCTCTCTG**ATCTGTCCCTCCACCCACAGTGGGGCCACTAGGGACAGGCGCTTCACTCGCTGGGTCCCTTTCTCTCTCTG**GGG 47/604583 (0.08%)

82 bp

Supplemental Fig.S11 Undesired edits of the *AAVSI* site were determined by HTS. (A) PCR strategy for amplifying the junction of TDs. The primers were designed to specifically amplify the boundary of targeted TDs. The HTS primers were listed in Supplemental Table S5.

(B) Undesired edits in indicated TDs of the *AAVSI* site. Sequence corresponding to accurate TD allele was shown on top of each analysis. The sequences corresponding to each end of the TD were showed in red and yellow respectively. The red arrowhead pointed to the boundary of the duplication. Insertion and base substitution were indicated by red box and green letters respectively. Note that the inserted sequence was same as its 5' upstream sequence. Sequences with a ratio over 0.02% were shown.

EMX1 TD 116-bp

TD reference sequence: CTGGAGGAGGAAGGGCCTGAGTCCGAGCAGAAGAA		TTCTCATCTGTGCCCTCCCTCCCTGGCCACAGGT	reads / total reads
2 × TD (Accurate) :	CTGGAGGAGGAAGGGCCTGAGTCCGAGCAGAAGAA	TTCTCATCTGTGCCCTCCCTCCCTGGCCACAGGT	39852/ 41510 (96.01%)
2 × TD with insertion :	CTGGAGGAGGAAGGGCCTGAGTCCGAGCAGAAGAA	TTCTCATCTGTGCCCTCCCTCCCTGGCCACAGGT	786/41510 (1.90%)
	CTGGAGGAGGAAGGGCCTGAGTCCGAGCAGAAGAA	TTCTCATCTGTGCCCTCCCTCCCTGGCCACAGGT	480/41510 (1.16%)
	CTGGAGGAGGAAGGGCCTGAGTCCGAGCAGAAGAA	TTCTCATCTGTGCCCTCCCTCCCTGGCCACAGGT	704/41510 (0.17%)
	CTGGAGGAGGAAGGGCCTGAGTCCGAGCAGAAGAA	TTCTCATCTGTGCCCTCCCTCCCTGGCCACAGGT	65/41510 (0.16%)
	CTGGAGGAGGAAGGGCCTGAGTCCGAGCAGAAGAA	TTCTCATCTGTGCCCTCCCTCCCTGGCCACAGGT	51/41510 (0.12%)
	CTGGAGGAGGAAGGGCCTGAGTCCGAGCAGAAGAA	TTCTCATCTGTGCCCTCCCTCCCTGGCCACAGGT	47/41510 (0.11%)
	CTGGAGGAGGAAGGGCCTGAGTCCGAGCAGAAGAA	TTCTCATCTGTGCCCTCCCTCCCTGGCCACAGGT	46/41510 (0.11%)
	CTGGAGGAGGAAGGGCCTGAGTCCGAGCAGAAGAA	TTCTCATCTGTGCCCTCCCTCCCTGGCCACAGGT	36/41510 (0.087%)
	CTGGAGGAGGAAGGGCCTGAGTCCGAGCAGAAGAA	TTCTCATCTGTGCCCTCCCTCCCTGGCCACAGGT	35/41510 (0.084%)
	CTGGAGGAGGAAGGGCCTGAGTCCGAGCAGAAGAA	TTCTCATCTGTGCCCTCCCTCCCTGGCCACAGGT	22/41510 (0.053%)
	CTGGAGGAGGAAGGGCCTGAGTCCGAGCAGAAGAA	TTCTCATCTGTGCCCTCCCTCCCTGGCCACAGGT	22/41510 (0.053%)
	CTGGAGGAGGAAGGGCCTGAGTCCGAGCAGAAGAA	TTCTCATCTGTGCCCTCCCTCCCTGGCCACAGGT	21/41510 (0.051%)
	CTGGAGGAGGAAGGGCCTGAGTCCGAGCAGAAGAA	TTCTCATCTGTGCCCTCCCTCCCTGGCCACAGGT	20/41510 (0.048%)
	CTGGAGGAGGAAGGGCCTGAGTCCGAGCAGAAGAA	TTCTCATCTGTGCCCTCCCTCCCTGGCCACAGGT	20/41510 (0.048%)
	CTGGAGGAGGAAGGGCCTGAGTCCGAGCAGAAGAA	TTCTCATCTGTGCCCTCCCTCCCTGGCCACAGGT	20/41510 (0.048%)
	CTGGAGGAGGAAGGGCCTGAGTCCGAGCAGAAGAA	TTCTCATCTGTGCCCTCCCTCCCTGGCCACAGGT	19/41510 (0.046%)
	CTGGAGGAGGAAGGGCCTGAGTCCGAGCAGAAGAA	TTCTCATCTGTGCCCTCCCTCCCTGGCCACAGGT	17/41510 (0.041%)
	CTGGAGGAGGAAGGGCCTGAGTCCGAGCAGAAGAA	TTCTCATCTGTGCCCTCCCTCCCTGGCCACAGGT	16/41510 (0.039%)
	CTGGAGGAGGAAGGGCCTGAGTCCGAGCAGAAGAA	TTCTCATCTGTGCCCTCCCTCCCTGGCCACAGGT	16/41510 (0.039%)
	CTGGAGGAGGAAGGGCCTGAGTCCGAGCAGAAGAA	TTCTCATCTGTGCCCTCCCTCCCTGGCCACAGGT	14/41510 (0.034%)
	CTGGAGGAGGAAGGGCCTGAGTCCGAGCAGAAGAA	TTCTCATCTGTGCCCTCCCTCCCTGGCCACAGGT	14/41510 (0.034%)
2 × TD with deletion :	CTGGAGGAGGAAGGGCCTGAGTCCGAGCAGAAGAA	TTCTCATCTGTGCCCTCCCTCCCTGGCCACAGGT	26/41510 (0.063%)

Supplemental Fig. S12 Undesired edits of *EMXI* TD 116-bp were determined by HTS. Sequence corresponding to accurate TD allele was shown on top of the analysis. The sequences corresponding to each end of the TD were showed in red and yellow respectively. The red arrowhead pointed to the boundary of the duplication. Deletion and insertion were indicated by black dash and red box. Note that the inserted sequence was the same as its 5' upstream sequence. The inserted sequences that were likely derived from sgRNA scaffold were highlighted in blue. Sequences with a ratio over 0.02% were shown.

EMX1 TD 201-bp

		Insertion	
TD reference sequence:.....CTGGAGGAGGAAGGGCCTGAGTCCGAGCAGAAGAA			
2 × TD (Accurate) :		CCCTATGTAGCCTCAGTCTTCCCATCAGGCTCTCA.....	reads / total reads
2 × TD with insertion :		CCCTATGTAGCCTCAGTCTTCCCATCAGGCTCTCA.....	519489/ 524263 (99.09%)
.....CTGGAGGAGGAAGGGCCTGAGTCCGAGCAGAAGAA		CCCTATGTAGCCTCAGTCTTCCCATCAGGCTCTCA.....	1620/524263 (0.31%)
.....CTGGAGGAGGAAGGGCCTGAGTCCGAGCAGAAGAA		CCCTATGTAGCCTCAGTCTTCCCATCAGGCTCTCA.....	1408/524263 (0.27%)
.....CTGGAGGAGGAAGGGCCTGAGTCCGAGCAGAAGAA		CCCTATGTAGCCTCAGTCTTCCCATCAGGCTCTCA.....	327/524263 (0.062%)
.....CTGGAGGAGGAAGGGCCTGAGTCCGAGCAGAAGAA		CCCTATGTAGCCTCAGTCTTCCCATCAGGCTCTCA.....	308/524263 (0.059%)
.....CTGGAGGAGGAAGGGCCTGAGTCCGAGCAGAAGAA		CCCTATGTAGCCTCAGTCTTCCCATCAGGCTCTCA.....	203/524263 (0.039%)
.....CTGGAGGAGGAAGGGCCTGAGTCCGAGCAGAAGAA		CCCTATGTAGCCTCAGTCTTCCCATCAGGCTCTCA.....	146/524263 (0.028%)
.....CTGGAGGAGGAAGGGCCTGAGTCCGAGCAGAAGAA		CCCTATGTAGCCTCAGTCTTCCCATCAGGCTCTCA.....	143/524263 (0.027%)
.....CTGGAGGAGGAAGGGCCTGAGTCCGAGCAGAAGAA		CCCTATGTAGCCTCAGTCTTCCCATCAGGCTCTCA.....	136/524263 (0.026%)
.....CTGGAGGAGGAAGGGCCTGAGTCCGAGCAGAAGAA		CCCTATGTAGCCTCAGTCTTCCCATCAGGCTCTCA.....	132/524263 (0.025%)
.....CTGGAGGAGGAAGGGCCTGAGTCCGAGCAGAAGAA		CCCTATGTAGCCTCAGTCTTCCCATCAGGCTCTCA.....	127/524263 (0.024%)
.....CTGGAGGAGGAAGGGCCTGAGTCCGAGCAGAAGAA		CCCTATGTAGCCTCAGTCTTCCCATCAGGCTCTCA.....	115/524263 (0.022%)
.....CTGGAGGAGGAAGGGCCTGAGTCCGAGCAGAAGAA		CCCTATGTAGCCTCAGTCTTCCCATCAGGCTCTCA.....	109/524263 (0.021%)

Supplemental Fig. S13 Undesired edits of *EMXI* TD 201-bp were determined by HTS.

Sequences corresponding to accurate TD allele was shown on top of the analysis. The sequences corresponding to each end of the TD were showed in red and yellow respectively. The red arrowhead pointed to the boundary of the duplication. Insertions was indicated by red boxes. Note that most inserted sequences were same as their 5' upstream sequences. A small group of insertions that were likely derived from sgRNA scaffold were highlighted in blue. Sequences with a ratio over 0.02% were shown.

- Deletion Insertion

D reference sequence:	reads / total reads
2 × TD (Accurate) :	207814 / 210165 (98.88%)
2 × TD with insertion :	
CTGGAGGAGGAAGGGCCTGAGTCCGAGCAGAAGAATTCCTCTG GCCCACTGTGCTCTTCTG GCGCTG	7432/210165 (0.35%)
CTGGAGGAGGAAGGGCCTGAGTCCGAGCAGAAGAATTCCTCTG GCCCACTGTGCTCTTCTG GCGCTG	142/210165 (0.068%)
CTGGAGGAGGAAGGGCCTGAGTCCGAGCAGAAGAATTCCTCTG GCCCACTGTGCTCTTCTG GCGCTG	82/210165 (0.039%)
CTGGAGGAGGAAGGGCCTGAGTCCGAGCAGAAGAATTCCTCTG GCCCACTGTGCTCTTCTG GCGCTG	69/210165 (0.033%)
CTGGAGGAGGAAGGGCCTGAGTCCGAGCAGAAGAATTCCTCTG GCCCACTGTGCTCTTCTG GCGCTG	60/210165 (0.029%)
CTGGAGGAGGAAGGGCCTGAGTCCGAGCAGAAGAATTCCTCTG GCCCACTGTGCTCTTCTG GCGCTG	58/210165 (0.028%)
CTGGAGGAGGAAGGGCCTGAGTCCGAGCAGAAGAATTCCTCTG GCCCACTGTGCTCTTCTG GCGCTG	51/210165 (0.024%)
2 × TD with deletion :	
CTGGAGGAGGAAGGGCCTGAGTCCGA - CAGAAGAATTCCTCTG GCCCACTGTGCTCTTCTG GCGCTG	337/210165 (0.16%)
CTGGAGGAGGAAGGGCCT - AGTCCGAGCAGAAGAATTCCTCTG GCCCACTGTGCTCTTCTG GCGCTG	153/210165 (0.073%)
CTGGAGGAGGAAGGGCCTGAGTCCGAGCAGAAGAATTCCTCT - GGCCCACTGTGCTCTTCTG GCGCTG	125/210165 (0.059%)
CTGGAGGAGGAAGGGCCTGAGTCCG - GCAGAAGAATTCCTCTG GCCCACTGTGCTCTTCTG GCGCTG	125/210165 (0.059%)
CTGGAGGAGGA - GCGCTGAGTCCGAGCAGAAGAATTCCTCTG GCCCACTGTGCTCTTCTG GCGCTG	124/210165 (0.059%)
CTGGAGGAG - AGCGGCTGAGCAGAAGAAGAATTCCTCTG GCCCACTGTGCTCTTCTG GCGCTG	94/210165 (0.045%)
CTGGAGGAG - AGCGGCTGAGCAGAAGAAGAATTCCTCTG GCCCACTGTGCTCTTCTG GCGCTG	79/210165 (0.038%)
CTGGAGGA - GGCGCTGAGTCCGAGCAGAAGAATTCCTCTG GCCCACTGTGCTCTTCTG GCGCTG	58/210165 (0.028%)
CTGG - AGGAAGGGCCTGAGTCCGAGCAGAAGAATTCCTCTG GCCCACTGTGCTCTTCTG GCGCTG	51/210165 (0.024%)

Supplemental Fig. S14 Undesired edits of *EMXI* TD 307-bp were determined by HTS.

Sequence corresponding to accurate TD allele was shown on top of each analysis. The sequences corresponding to each end of the TD were showed in red and yellow respectively. The red arrowhead pointed to the boundary of the duplication. Deletion, insertion and base substitution were indicated by black dash, red box and green letters respectively. The inserted sequence in the red box was same as with its 5' upstream sequence. The sequences with a ratio over 0.02% were shown.

[illegible]

Supplemental Fig. S15 Undesired edits of *FANCF* TD 115-bp were determined by HTS.

Sequence corresponding to accurate TD allele was shown on top of the analysis. The sequences corresponding to each end of the TD were showed in red and yellow respectively. The red arrowhead pointed to the boundary of the duplication. Deletion, insertion and base substitution were indicated by black dash, red box and green letters respectively. Note that the inserted sequence in red box was same as its 5' upstream sequence. A small group of insertions that were likely derived from sgRNA scaffold were highlighted in blue. The sequences with a ratio over 0.02% were shown.

- Deletion Insertion

Reference sequence: GGCCCTTCGGCGACCTCATGGAATCCCTCTTCGCAGCGTCAAGGAACACGGGATAAAGACGCTGGGAGATTGA.	reads / total reads
2 × TD (Accurate) : GGCCCTTCGGCGACCTCATGGAATCCCTCTTCGCAGCGTCAAGGAACACGGGATAAAGACGCTGGGAGATTGA.	197338/208725 (94.54%)
2 × TD with insertion : GGCCCTTCGGCGACCTCATGGAATCCCTCTTCGCAGCGTCAAGGAACACGGGATAAAGACGCTGGGAGATTGA.	971/208725 (0.47%)
. GGCCCTTCGGCGACCTCATGGAATCCCTCTTCGCAGCGTCAAGGAACACGGGATAAAGACGCTGGGAGATTGA.	829/208725 (0.40%)
. GGCCCTTCGGCGACCTCATGGAATCCCTCTTCGCAGCGTCAAGGAACACGGGATAAAGACGCTGGGAGATTGA.	399/208725 (0.19%)
. GGCCCTTCGGCGACCTCATGGAATCCCTCTTCGCAGCGTCAAGGAACACGGGATAAAGACGCTGGGAGATTGA.	341/208725 (0.16%)
. GGCCCTTCGGCGACCTCATGGAATCCCTCTTCGCAGCGTCAAGGAACACGGGATAAAGACGCTGGGAGATTGA.	298/208725 (0.14%)
. GGCCCTTCGGCGACCTCATGGAATCCCTCTTCGCAGCGTCAAGGAACACGGGATAAAGACGCTGGGAGATTGA.	233/208725 (0.11%)
. GGCCCTTCGGCGACCTCATGGAATCCCTCTTCGCAGCGTCAAGGAACACGGGATAAAGACGCTGGGAGATTGA.	212/208725 (0.10%)
. GGCCCTTCGGCGACCTCATGGAATCCCTCTTCGCAGCGTCAAGGAACACGGGATAAAGACGCTGGGAGATTGA.	182/208725 (0.087%)
. GGCCCTTCGGCGACCTCATGGAATCCCTCTTCGCAGCGTCAAGGAACACGGGATAAAGACGCTGGGAGATTGA.	128/208725 (0.062%)
. GGCCCTTCGGCGACCTCATGGAATCCCTCTTCGCAGCGTCAAGGAACACGGGATAAAGACGCTGGGAGATTGA.	128/208725 (0.061%)
. GGCCCTTCGGCGACCTCATGGAATCCCTCTTCGCAGCGTCAAGGAACACGGGATAAAGACGCTGGGAGATTGA.	122/208725 (0.058%)
. GGCCCTTCGGCGACCTCATGGAATCCCTCTTCGCAGCGTCAAGGAACACGGGATAAAGACGCTGGGAGATTGA.	112/208725 (0.054%)
. GGCCCTTCGGCGACCTCATGGAATCCCTCTTCGCAGCGTCAAGGAACACGGGATAAAGACGCTGGGAGATTGA.	102/208725 (0.049%)
. GGCCCTTCGGCGACCTCATGGAATCCCTCTTCGCAGCGTCAAGGAACACGGGATAAAGACGCTGGGAGATTGA.	94/208725 (0.045%)
. GGCCCTTCGGCGACCTCATGGAATCCCTCTTCGCAGCGTCAAGGAACACGGGATAAAGACGCTGGGAGATTGA.	86/208725 (0.041%)
. GGCCCTTCGGCGACCTCATGGAATCCCTCTTCGCAGCGTCAAGGAACACGGGATAAAGACGCTGGGAGATTGA.	82/208725 (0.039%)
. GGCCCTTCGGCGACCTCATGGAATCCCTCTTCGCAGCGTCAAGGAACACGGGATAAAGACGCTGGGAGATTGA.	82/208725 (0.039%)
. GGCCCTTCGGCGACCTCATGGAATCCCTCTTCGCAGCGTCAAGGAACACGGGATAAAGACGCTGGGAGATTGA.	71/208725 (0.034%)
. GGCCCTTCGGCGACCTCATGGAATCCCTCTTCGCAGCGTCAAGGAACACGGGATAAAGACGCTGGGAGATTGA.	69/208725 (0.033%)
. GGCCCTTCGGCGACCTCATGGAATCCCTCTTCGCAGCGTCAAGGAACACGGGATAAAGACGCTGGGAGATTGA.	67/208725 (0.032%)
. GGCCCTTCGGCGACCTCATGGAATCCCTCTTCGCAGCGTCAAGGAACACGGGATAAAGACGCTGGGAGATTGA.	66/208725 (0.032%)
. GGCCCTTCGGCGACCTCATGGAATCCCTCTTCGCAGCGTCAAGGAACACGGGATAAAGACGCTGGGAGATTGA.	64/208725 (0.031%)
. GGCCCTTCGGCGACCTCATGGAATCCCTCTTCGCAGCGTCAAGGAACACGGGATAAAGACGCTGGGAGATTGA.	63/208725 (0.030%)
. GGCCCTTCGGCGACCTCATGGAATCCCTCTTCGCAGCGTCAAGGAACACGGGATAAAGACGCTGGGAGATTGA.	62/208725 (0.030%)
. GGCCCTTCGGCGACCTCATGGAATCCCTCTTCGCAGCGTCAAGGAACACGGGATAAAGACGCTGGGAGATTGA.	61/208725 (0.029%)
. GGCCCTTCGGCGACCTCATGGAATCCCTCTTCGCAGCGTCAAGGAACACGGGATAAAGACGCTGGGAGATTGA.	54/208725 (0.026%)
2 × TD with deletion : GGCCCTTCGGCGACCTCATGGAATCCCTCTTCGCAGCGTCAAGGAACACGGGATAAAGACGCTGGGAGATTGA.	5977/208725 (2.86%)
. GGCCCTTCGGCGACCTCATGGAATCCCTCTTCGCAGCGTCAAGGAACACGGGATAAAGACGCTGGGAGATTGA.	154/208725 (0.074%)
. GGCCCTTCGGCGACCTCATGGAATCCCTCTTCGCAGCGTCAAGGAACACGGGATAAAGACGCTGGGAGATTGA.	146/208725 (0.070%)
. GGCCCTTCGGCGACCTCATGGAATCCCTCTTCGCAGCGTCAAGGAACACGGGATAAAGACGCTGGGAGATTGA.	89/208725 (0.043%)
. GGCCCTTCGGCGACCTCATGGAATCCCTCTTCGCAGCGTCAAGGAACACGGGATAAAGACGCTGGGAGATTGA.	75/208725 (0.036%)
. GGCCCTTCGGCGACCTCATGGAATCCCTCTTCGCAGCGTCAAGGAACACGGGATAAAGACGCTGGGAGATTGA.	54/208725 (0.026%)
. GGCCCTTCGGCGACCTCATGGAATCCCTCTTCGCAGCGTCAAGGAACACGGGATAAAGACGCTGGGAGATTGA.	51/208725 (0.024%)

Supplemental Fig. S16 Undesired edits of *FANCF* TD 171-bp were determined by HTS.

Sequence corresponding to accurate TD allele was shown on top of the analysis. The sequences corresponding to each end of the TD were showed in red and yellow respectively. The red arrowhead pointed to the boundary of the duplication. Deletion, insertion and base substitution were indicated by black dash, red box and green letters respectively. Note that most inserted sequences were same as their 5' upstream sequences. A small group of insertions that were likely derived from sgRNA scaffold were highlighted in blue. The sequences with a ratio over 0.02% were shown.

TD reference sequence:	reads / total reads
2 × TD (Accurate) :	317231/330088 (96.10%)
2 × TD with insertion :	989/330088 (0.30%)
.....	762/330088 (0.23%)
.....	684/330088 (0.21%)
.....	677/330088 (0.21%)
.....	780/330088 (0.24%)
.....	553/330088 (0.17%)
.....	358/330088 (0.11%)
.....	350/330088 (0.11%)
.....	319/330088 (0.10%)
.....	295/330088 (0.089%)
.....	255/330088 (0.077%)
.....	252/330088 (0.076%)
.....	244/330088 (0.074%)
.....	223/330088 (0.068%)
.....	219/330088 (0.065%)
.....	212/330088 (0.064%)
.....	203/330088 (0.061%)
.....	198/330088 (0.060%)
.....	175/330088 (0.053%)
.....	172/330088 (0.052%)
.....	167/330088 (0.051%)
.....	164/330088 (0.050%)
.....	161/330088 (0.049%)
.....	157/330088 (0.048%)
.....	146/330088 (0.044%)
.....	145/330088 (0.044%)
.....	136/330088 (0.041%)
.....	135/330088 (0.041%)
.....	135/330088 (0.041%)
.....	134/330088 (0.041%)
.....	124/330088 (0.038%)
.....	123/330088 (0.037%)
.....	121/330088 (0.037%)
.....	119/330088 (0.036%)
.....	115/330088 (0.035%)
.....	109/330088 (0.033%)
.....	107/330088 (0.032%)
.....	106/330088 (0.032%)
.....	104/330088 (0.032%)
.....	102/330088 (0.031%)
.....	101/330088 (0.031%)
.....	97/330088 (0.029%)
.....	95/330088 (0.028%)
.....	93/330088 (0.028%)
.....	92/330088 (0.028%)
.....	89/330088 (0.027%)
.....	87/330088 (0.026%)
.....	86/330088 (0.026%)
.....	85/330088 (0.026%)
.....	84/330088 (0.025%)
.....	83/330088 (0.025%)
.....	81/330088 (0.025%)
.....	76/330088 (0.023%)
.....	74/330088 (0.022%)
.....	73/330088 (0.022%)
2 × TD with deletion :	170/330088 (0.052%)
.....	161/330088 (0.049%)
.....	148/330088 (0.045%)
.....	138/330088 (0.041%)
.....	115/330088 (0.035%)
.....	84/330088 (0.025%)

Supplemental Fig. S17 Undesired edits of *FANCF* TD 231-bp were determined by HTS.

Sequence corresponding to accurate TD allele was shown on top of the analysis. The sequences corresponding to each end of the TD were showed in red and yellow respectively. The red arrowhead pointed to the boundary of the duplication. Deletion, insertion and base substitution were indicated by black dash, red box and green letters respectively. Note that most inserted sequences were same as their 5' upstream sequences. A small group of insertions that were likely derived from sgRNA scaffold were highlighted in blue. The sequences with a ratio over 0.02% were shown.

 Insertion

Supplemental Fig. S18 Undesired edits of *HEK3* TD 136-bp were determined by HTS.

Sequence corresponding to accurate TD allele was shown on top of the analysis. The sequences corresponding to each end of the TD were showed in red and yellow respectively. The red arrowhead pointed to the boundary of the duplication. Insertion and base substitution were indicated by red box and green letters respectively. Note that most inserted sequences were same as their 5' upstream sequences. A small group of insertions that were likely derived from sgRNA scaffold were highlighted in blue. The sequences with a ratio over 0.02% were shown.

HEK3 TD 222-bp

			- Deletion Insertion	
TD reference sequence:.....CCCTGGCCTGGGTCAATCCTTGGGGCCAGACTGAGCACGGGGAGCTTGGCATGAGAAACCTTGGAGAGTTTAAAGCAAG.....			reads / total reads	
2 × TD (Accurate) :CCCTGGCCTGGGTCAATCCTTGGGGCCAGACTGAGCACGGGGAGCTTGGCATGAGAAACCTTGGAGAGTTTAAAGCAAG.....		149318/ 151942 (98.27%)	
2 × TD with insertion :CCCTGGCCTGGGTCAATCCTTGGGGCCAGACTGAGCA TCCCTTGGGGCCAGACTGAGCA CGGGAGCTTGGCATGAGAAACCTTGGAGAGTTTAAAGCAAG.....	22-bp	618/151942 (0.41%)	
CCCTGGCCTGGGTCAATCCTTGGGGCCAGACTGAGCACGGGGAGCTTGGCATGAGAA TCCCTTGGGGCCAGACTGAGCACGGGGAGCTTGGCATGAGAA ACCTTGGAGAGTTTAAAGCAAG.....	42-bp	452/151942 (0.30%)	
CCCTGGCCTGGGTCAATCCTTGGGGCCAGACTGAGCACGGGGAGCTTGGCATGAGAAACCTTGG GGCCAGACTGAGCACGGGGAGCTTGGCATGAGAAACCTTGG AGAGTTTAAAGCAAG.....	42-bp	394/151942 (0.26%)	
CCCTGGCCTGGGTCAATCCTTGGGGCCAGACTGAGCACGGGGAGCTTGGCATGAGAAACCTTGG ATCCTTGGGGCCAGACTGAGCACGGGGAGCTTGGCATGAGAAACCTTGG SAGAGTTT.....	50-bp	173/151942 (0.11%)	
CCCTGGCCTGGGTCAATCCTTGGGGCCAGACT TGGGGCCAGACT 3AGCACGGGGAGCTTGGCATGAGAAACCTTGGAGAGTTTAAAGCAAG.....	13-bp	114/151942 (0.075%)	
CCCTGGCCTGGGTCAATCCTTGGGGCCAGACTGAGCA ATCCTTGGGGCCAGACTGAGCA CGGGAGCTTGGCATGAGAAACCTTGGAGAGTTTAAAGCAAG.....	23-bp	93/151942 (0.061%)	
CCCTGGCCTGGGTCAATCCTTGGGGCCAGACTGAGCAC GACTGAGCACG GGAGCTTGGCATGAGAAACCTTGGAGAGTTTAAAGCAAG.....	12-bp	93/151942 (0.061%)	
CCCTGGCCTGGGTCAATCCTTGGGGCCAGACTGAGCACGGGGAGCTTGG T GAGCACGGGGAGCTTGG CAT GAGAAACCTTGGAGAGTTTAAAGCAAG.....	18-bp	75/151942 (0.049%)	
CCCTGGCCTGGGTCAATCCTTGGGGCCAGACTGAGCACGGGGAGCTTGGCAT GGGCCAGACTGAGCACGGGGAGCTTGGCATG AGAAACCTTGGAGAGTTTAAAGCAAG.....	32-bp	72/151942 (0.047%)	
CCCTGGCCTGGGTCAATCCTTGGGGCCAGACTGAGCACGGGGAGCTTGGCATG ATCCTTGGGGCCAGACTGAGCACGGGGAGCTTGGCATG AAACCTTGGAGAGTTTAAAGCAAG.....	42-bp	67/151942 (0.044%)	
CCCTGGCCTGGGTCAATCCTTGGGGCCAGACT ATCCTTGGGGCC CA G AGCTGAGCACGGGGAGCTTGGCATGAGAAACCTTGGAGAGTTTAAAGCAAG.....	11-bp	65/151942 (0.043%)	
CCCTGGCCTGGGTCAATCCTTGGGGCCAGACT TTGGGGCCAGACTGAGC ACGGGGAGCTTGGCATGAGAAACCTTGGAGAGTTTAAAGCAAG.....	18-bp	64/151942 (0.042%)	
CCCTGGCCTGGGTCAATCCTTGGGGCCAGACT ATCCTTGGGGCCAGACT TGAGCACGGGGAGCTTGGCATGAGAAACCTTGGAGAGTTTAAAGCAAG.....	17-bp	62/151942 (0.041%)	
CCCTGGCCTGGGTCAATCCTTGGGGCCAGACT GGTGCATCCTTGGGGCCAGACT TGAGCACGGGGAGCTTGGCATGAGAAACCTTGGAGAGTTTAAAGCAAG.....	22-bp	50/151942 (0.033%)	
CCCTGGCCTGGGTCAATCCTTGGGGCCAGACTGAGCACGGGGAGCTTGGCATGAG GAGCTTGGCATGAG AAACCTTGGAGAGTTTAAAGCAAG.....	14-bp	49/151942 (0.032%)	
CCCTGGCCTGGGTCAATCCTTGGGGCCAGACTGAGCA TCATCCTTGGGGCCAGACTGAGCA CGGGAGCTTGGCATGAGAAACCTTGGAGAGTTTAAAGCAAG.....	24-bp	48/151942 (0.030%)	
CCCTGGCCTGGGTCAATCCTTGGGGCCAGACTGAGCACGGGGAGCTTGGCAT CCTTGGGGCCAGACTGAGCACGGGGAGCTTGGCAT GAGAAACCTTGGAGAGTTTAAAGCAAG.....	36-bp	43/151942 (0.028%)	
CCCTGGCCTGGGTCAATCCTTGGGGCCAGACTGAGCACGGGGAGCTTGG GGCCAGACTGAGCACGGGGAGCTTGG CATGAGAAACCTTGGAGAGTTTAAAGCAAG.....	27-bp	42/151942 (0.028%)	
2 × TD with deletion :CCCTGGCCTGGGTCAATCCTTGGGGCCAGACT-----GAGCTTGGCATGAGAAACCTTGGAGAGTTTAAAGCAAG.....		52/151942 (0.034%)	

Supplemental Fig. S19 Undesired edits of *HEK3* TD 222-bp were determined by HTS.

Sequence corresponding to accurate TD allele was shown on top of the analysis. The sequences corresponding to each end of the TD were showed in red and yellow respectively. The red arrowhead pointed to the boundary of the duplication. Deletion, insertion and base substitution were indicated by black dash, red box and green letters respectively. Note that most inserted sequences were the same as their 5' upstream sequences. A small group of insertions that were likely derived from sgRNA scaffold were highlighted in blue. The sequences with a ratio over 0.02% were shown.

HEK3 TD 397-bp

		- Deletion Insertion	
TD reference sequence:TCTCCCCCATGTCCTACATAAGACAATGGATACACGGGTCGAGCTCAACAGAGGAAAAGATCTCAGGGCA.		reads / total reads	
2 × TD (Accurate) :TCTCCCCCATGTCCTACATAAGACAATGGATACACGGGTCGAGCTCAACAGAGGAAAAGATCTCAGGGCA.	366436/370811 (98.82%)	
2 × TD with insertion :TATCTCCCCCATGTCCTACATAAGACAAT TCTCCCCCATGTCCTACATAAGACAAT GGATACACGGGTCGAGCTCAACAGAGGAAAAGATCTCAGGGCA.	427/370811 (0.12%)	
TCTCCCCCATGTCCTACATAAGACAATGGATACACGGGTCGAGCTCAACAGAGGAAAAGATCTCAGGGCA.	225/370811 (0.061%)	
TCTCCCCCATGTCCTACATAAGACAATGGATAC TCTCCCCCATGTCCTACATAAGACAATGGATAC CGGTCGAGCTCAACAGAGGAAAAGATCTCAGGGCA.	180/370811 (0.049%)	
TCTCCCCCATGTCCTACATAAGACAATGGATAC TCTCCCCCATGTCCTACATAAGACAATGGATAC CGGTCGAGCTCAACAGAGGAAAAGATCTCAGGGCA.	149/370811 (0.040%)	
TCTCCCCCATGTCCTACATAAGACAATGGATAC TCTCCCCCATGTCCTACATAAGACAATGGATAC CGGTCGAGCTCAACAGAGGAAAAGATCTCAGGGCA.	121/370811 (0.033%)	
TCTCCCCCATGTCCTACATAAGACAATGGATAC TCTCCCCCATGTCCTACATAAGACAATGGATAC CGGTCGAGCTCAACAGAGGAAAAGATCTCAGGGCA.	107/370811 (0.029%)	
TCTCCCCCATGTCCTACATAAGACAATGGATAC TCTCCCCCATGTCCTACATAAGACAATGGATAC CGGTCGAGCTCAACAGAGGAAAAGATCTCAGGGCA.	104/370811 (0.028%)	
2 × TD with deletion :TCTCCCCCATGTCCTACATAAGACAATGGATACACGGGTCGAGCTCAACAGAGGAAAAGATCTCAGGGCA.	641/370811 (0.17%)	
TCTCCCCCATGTCCTACATAAGACAATGGATACACGGGTCGAGCTCAACAGAGGAAAAGATCTCAGGGCA.	578/370811 (0.16%)	
TCTCCCCCATGTCCTACATAAGACAATGGATACACGGGTCGAGCTCAACAGAGGAAAAGATCTCAGGGCA.	391/370811 (0.11%)	
TCTCCCCCATGTCCTACATAAGACAATGGATACACGGGTCGAGCTCAACAGAGGAAAAGATCTCAGGGCA.	269/370811 (0.073%)	
TCTCCCCCATGTCCTACATAAGACAATGGATACACGGGTCGAGCTCAACAGAGGAAAAGATCTCAGGGCA.	257/370811 (0.069%)	
TCTCCCCCATGTCCTACATAAGACAATGGATACACGGGTCGAGCTCAACAGAGGAAAAGATCTCAGGGCA.	255/370811 (0.069%)	
TCTCCCCCATGTCCTACATAAGACAATGGATACACGGGTCGAGCTCAACAGAGGAAAAGATCTCAGGGCA.	232/370811 (0.063%)	
TCTCCCCCATGTCCTACATAAGACAATGGATACACGGGTCGAGCTCAACAGAGGAAAAGATCTCAGGGCA.	227/370811 (0.061%)	
TCTCCCCCATGTCCTACATAAGACAATGGATACACGGGTCGAGCTCAACAGAGGAAAAGATCTCAGGGCA.	212/370811 (0.057%)	

Supplemental Fig. S20 Undesired edits of *HEK3* TD 397-bp were determined by HTS.

Sequence corresponding to accurate TD allele was shown on top of the analysis. The sequences corresponding to each end of the TD were showed in red and yellow respectively. The red arrowhead pointed to the boundary of the duplication. Deletion and insertion were indicated by black dash and red box respectively. Note that most inserted sequences were same as their 5' upstream sequences. The sequences with a ratio over 0.02% were shown.

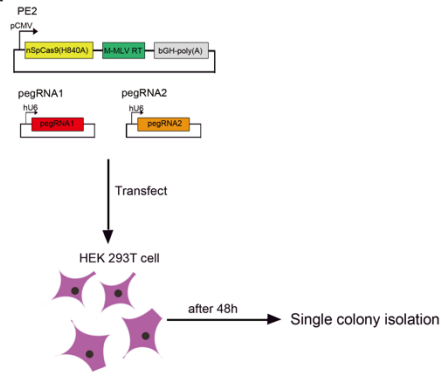
HEK3 TD 499-bp

TD reference sequence:ATCTCCCCCATGTCCTACATAAGACAATGGATACACCGGGGATACTGGTTGACAAGTTTGGCTGGGCTGGA.....	reads / total reads
2 × TD (Accurate) :ATCTCCCCCATGTCCTACATAAGACAATGGATACACCGGGGATACTGGTTGACAAGTTTGGCTGGGCTGGA.....	410401/410732 (99.92%)
2 × TD with insertion :ATCTCCCCCATGTCCTACATAAGACAATATCTCCCCCATGTCCTACATAAGACAATGGATACACCGGGGATACTGGTTGACAAGTTTGGCTGGGCTGGA.....	172/410732 (0.042%)
ATCTCCCCCATGTCCTACATAAGACAATGTCCTACATAAGACAATGGATACACCGGGGATACTGGTTGACAAGTTTGGCTGGGCTGGA.....	159/410732 (0.038%)

Supplemental Fig. S21 Undesired edits of *HEK3* TD 499-*bp* were determined by HTS.

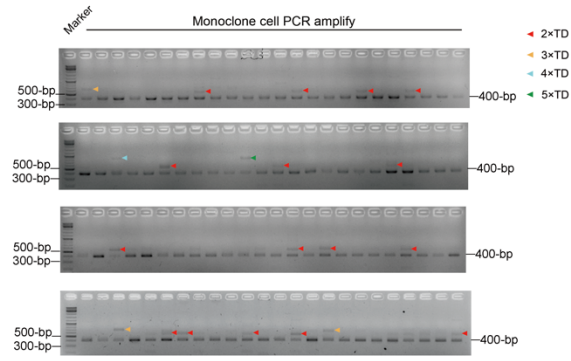
Sequence corresponding to accurate TD allele was shown on top of the analysis. The sequences corresponding to each end of the TD were showed in red and yellow respectively. The red arrowhead pointed to the boundary of the duplication. Insertion was indicated by red box. Note that most inserted sequences were same as their 5' upstream sequences. The sequences with a ratio over 0.02% were shown.

A



B

HEK3 TD 136-bp



C

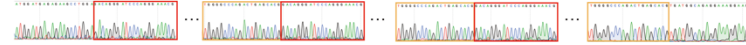
HEK3 TD 136-bp 2xTD

TD reference sequence: ATGGATGAGAGAAGCCTGGAGACAGGGATCCCAGGGAACG--TGGGGCCAGACTGAGCACGGACAGGGATCCCAGGGAACG--TGGGGCCAGACTGAGCACGTGATGGCAGAGAAAGGAAG



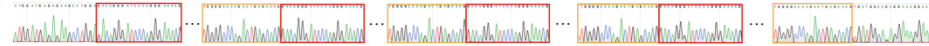
HEK3 TD 136-bp 3xTD

TD reference sequence: ATGGATGAGAGAAGCCTGGAGACAGGGATCCCAGGGAACG--TGGGGCCAGACTGAGCACGGACAGGGATCCCAGGGAACG--TGGGGCCAGACTGAGCACGTGATGGCAGAGAAAGGAAG



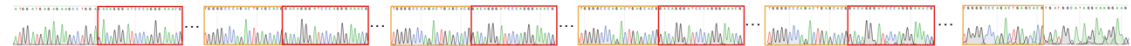
HEK3 TD 136-bp 4xTD

TD reference sequence: ATGGATGAGAGAAGCCTGGAGACAGGGATCCCAGGGAACG--TGGGGCCAGACTGAGCACGGACAGGGATCCCAGGGAACG--TGGGGCCAGACTGAGCACGTGATGGCAGAGAAAGGAAG

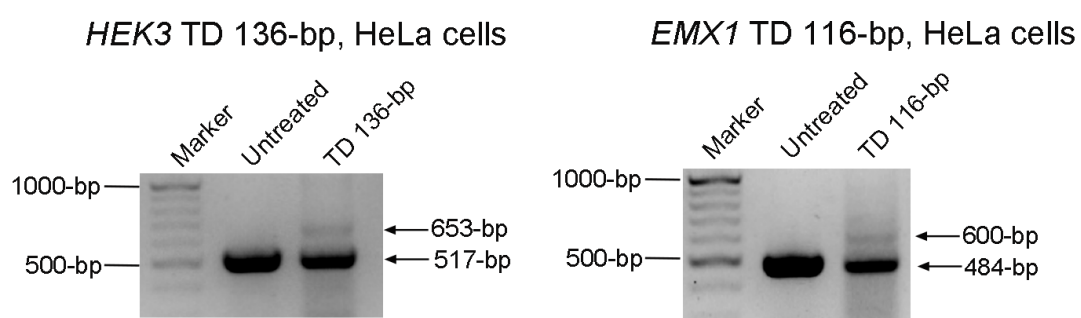
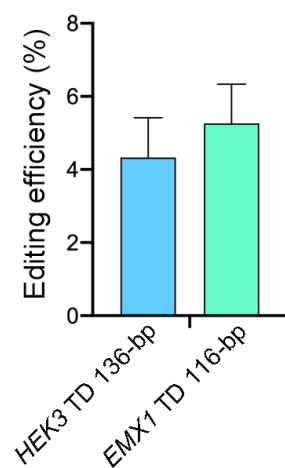


HEK3 TD 136-bp 5xTD

TD reference sequence: ATGGATGAGAGAAGCCTGGAGACAGGGATCCCAGGGAACG--TGGGGCCAGACTGAGCACGGACAGGGATCCCAGGGAACG--TGGGGCCAGACTGAGCACGTGATGGCAGAGAAAGGAAG

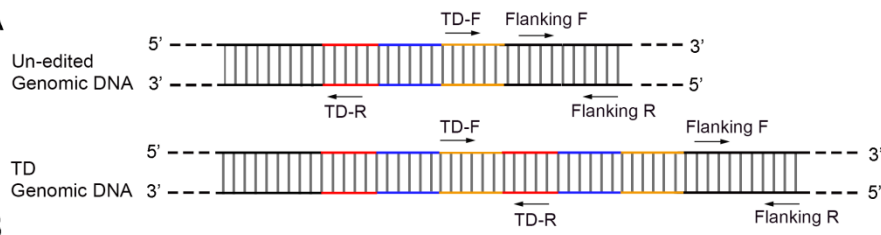


Supplemental Fig. S22 Monoclonal analysis of the *HEK3* TD 136-bp. (A) Diagram showing the procedure of monoclonal analysis. HEK293T cells were transfected with TD-PE plasmids, including PE2 and two pegRNA, and 48 hours post the transfection cells were diluted monoclonal isolation. (B) Agarose gel electrophoresis showed the amplicons derived from each monoclonal genome. The colonies containing different copies of TDs were highlighted with triangles of different colors. (C) Sanger sequencing results of representative TDs derived from monoclonal genomes. Reference sequences of each type of TDs were shown on top of the sequence chromatogram, and the duplicated sequences corresponding to each end of the TD were marked by yellow and red boxes respectively.

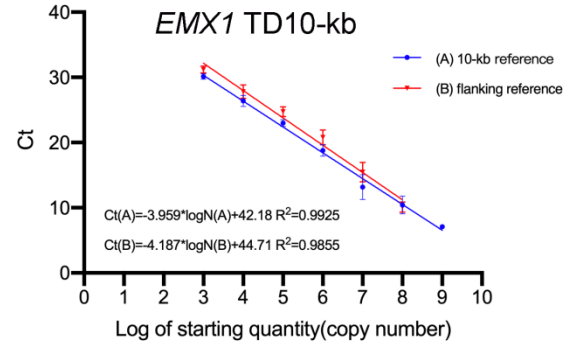
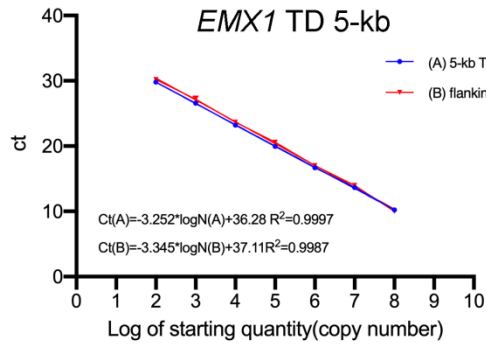
A**B**

Supplemental Fig. S23 Verification of TD-PE strategy in HeLa cells. (A) Agarose gel analysis of the presence of targeted tandem duplication at indicated loci. (B) Quantification of the efficiency of targeted tandem duplication in (A) using Image J. Values and error bars reflect mean \pm s.d. of n=3 independent biological replicates.

A

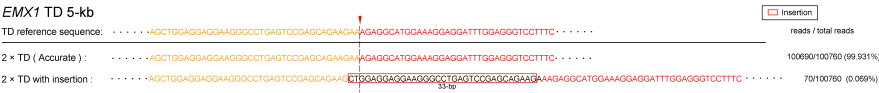


B

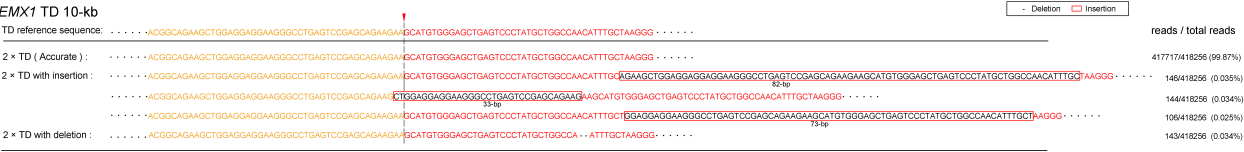


Supplemental Fig. S24 Standard curves for absolute quantification by real-time PCR of *EMXI* TD 5-kb and 10-kb. (A) Schematic diagram showing the design of qPCR primers. The flanking fragment of the target site or the one containing tandem duplication obtained by PCR were ligated to the blunt vector to construct reference plasmids. qPCR primers were designed inside the amplified fragment. The primers used to construct the reference plasmids and qPCR primers are listed in Supplemental Table S4. (B) The standard curves for the reference plasmids were determined by CT values against log-transformed concentrations of serial tenfold dilutions (2×10^2 , 10^3 , 10^4 , 10^5 , 10^6 , 10^7 , 10^8 and 10^9 copies per $1 \mu\text{l}$).

EMX1 TD 5-kb



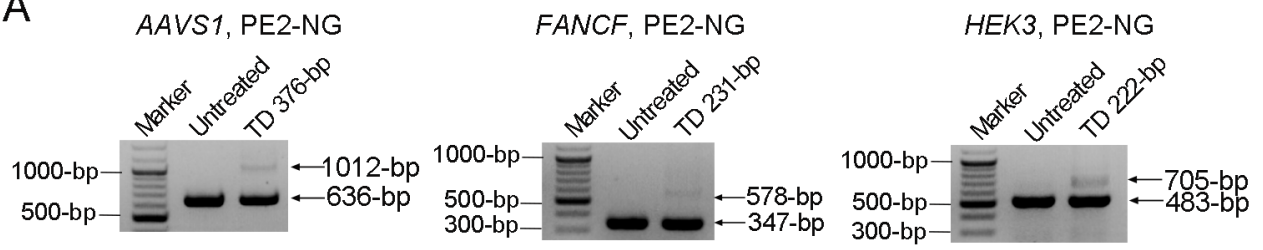
EMX1 TD 10-kb



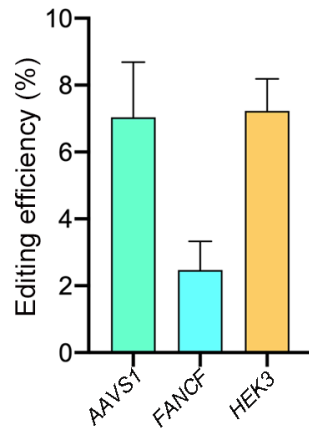
Supplemental Fig. S25 Undesired edits of *EMXI* TD 5-kb/10-kb were determined by HTS.

Sequence corresponding to accurate TD allele was shown on top of the analysis. The sequences corresponding to each end of the TD were showed in red and yellow respectively. The red arrowhead pointed to the boundary of the duplication. Deletion and insertion were indicated by black dash and red box respectively. Note that most inserted sequences were the same as their 5' upstream sequences. The sequences with a ratio over 0.02% were shown.

A



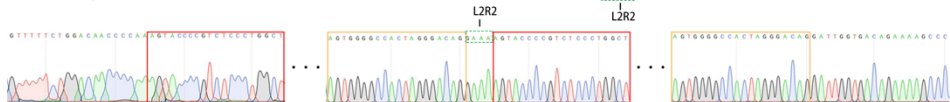
B



Supplemental Fig. S26 Targeted TDs produced by PE2-NG. (A) Targeted TDs produced by PE2-NG. Agarose gel analysis of the presence of targeted tandem duplication at indicated loci. B, Quantification of the efficiency of targeted tandem duplication in (B) using Image J. Values and error bars reflect mean \pm s.d. of n=3 independent biological replicates.

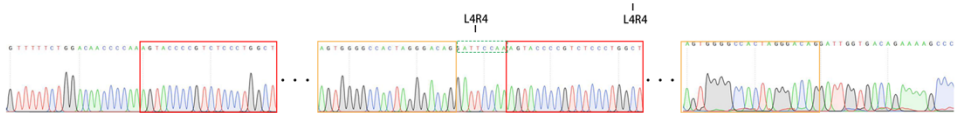
AAVS1 TD 376-bp L2R2

TD reference sequence: GTTTTCTGGACAACCCCAAAGTACCCCGTCTCCCTGGCT...AGTGGGGCCACTAGGGACAGGAAAAAGTACCCCGTCTCCCTGGCT...AGTGGGGCCACTAGGGACAGGATTGGTGACAGAAAAGCCC



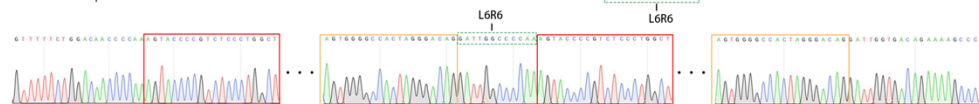
AAVS1 TD 376-bp L4R4

TD reference sequence: GTTTTCTGGACAACCCCAAAGTACCCCGTCTCCCTGGCT...AGTGGGGCCACTAGGGACAGGATTCCAAAGTACCCCGTCTCCCTGGCT...AGTGGGGCCACTAGGGACAGGATTGGTGACAGAAAAGCCC



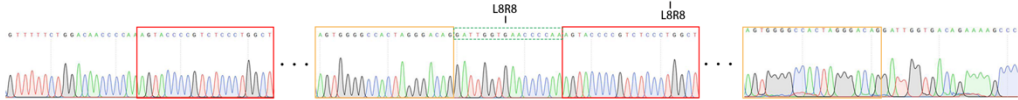
AAVS1 TD 376-bp L6R6

TD reference sequence: GTTTTCTGGACAACCCCAAAGTACCCCGTCTCCCTGGCT...AGTGGGGCCACTAGGGACAGGATTGGGCCCAAGTACCCCGTCTCCCTGGCT...AGTGGGGCCACTAGGGACAGGATTGGTGACAGAAAAGCCC



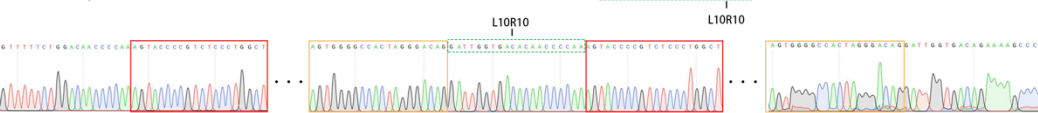
AAVS1 TD 376-bp L8R8

TD reference sequence: GTTTTCTGGACAACCCCAAAGTACCCCGTCTCCCTGGCT...AGTGGGGCCACTAGGGACAGGATTGGTGAACCCCAAAGTACCCCGTCTCCCTGGCT...AGTGGGGCCACTAGGGACAGGATTGGTGACAGAAAAGCCC



AAVS1 TD 376-bp L10R10

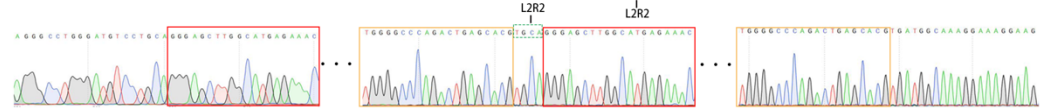
TD reference sequence: GTTTTCTGGACAACCCCAAAGTACCCCGTCTCCCTGGCT...AGTGGGGCCACTAGGGACAGGATTGGTGACACAACCCCAAAGTACCCCGTCTCCCTGGCT...AGTGGGGCCACTAGGGACAGGATTGGTGACAGAAAAGCCC



Supplemental Fig.S27 Representative Sanger sequencing results of the additive fragment containing TDs at the *AAVSI* site (related to Fig. 4B). The PCR amplicons containing targeted TDs were gel-purified and subjected directly to Sanger sequencing. TD reference sequences were shown on top of each Sequencing chromatogram. The sequences corresponding to each end of the TDs were marked by yellow and red boxes respectively. Sequences in green dashed boxes represent additive fragments.

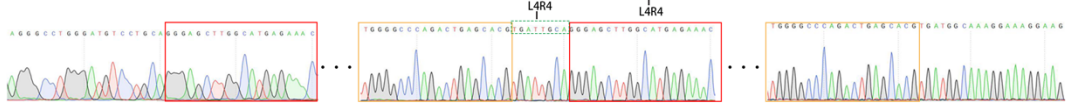
HEK3 TD 222-bp L2R2

TD reference sequence: AGGGCCTGGGATGTCCTGCA**GGGAGCTTGGCATGAGAAAC**...**TGGGGCCAGACTGAGCACCTGATGGCAGAGGAAAGGAAG**



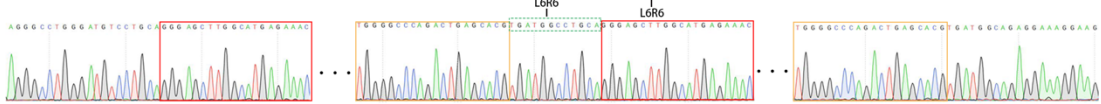
HEK3 TD 222-bp L4R4

TD reference sequence: AGGGCCTGGGATGTCCTGCA**GGGAGCTTGGCATGAGAAAC**...**TGGGGCCAGACTGAGCACCTGATGGCAGAGGAAAGGAAG**



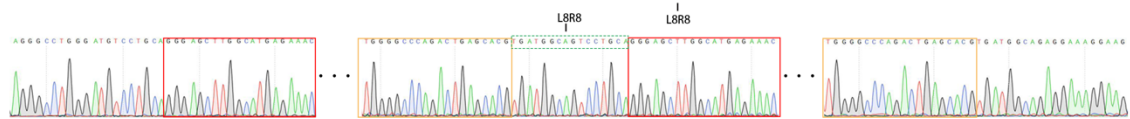
HEK3 TD 222-bp L6R6

TD reference sequence: AGGGCCTGGGATGTCCTGCA**GGGAGCTTGGCATGAGAAAC**...**TGGGGCCAGACTGAGCACCTGATGGCAGAGGAAAGGAAG**



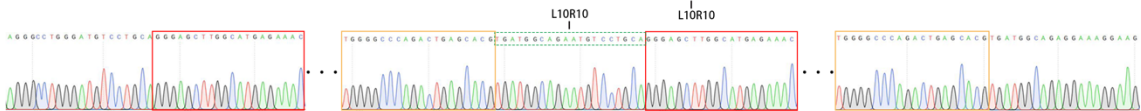
HEK3 TD 222-bp L8R8

TD reference sequence: AGGGCCTGGGATGTCCTGCA**GGGAGCTTGGCATGAGAAAC**...**TGGGGCCAGACTGAGCACCTGATGGCAGAGGAAAGGAAG**



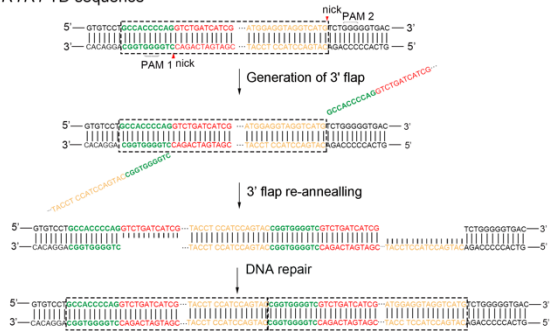
HEK3 TD 222-bp L10R10

TD reference sequence: AGGGCCTGGGATGTCCTGCA**GGGAGCTTGGCATGAGAAAC**...**TGGGGCCAGACTGAGCACCTGATGGCAGAGGAAAGGAAG**

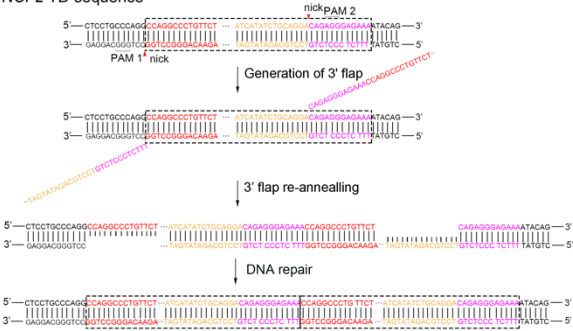


Supplemental Fig.S28 Representative Sanger sequencing results of the additive fragment containing TDs at the *HEK3* site (related to Fig. 4B). The PCR amplicons containing targeted TDs were gel-purified and subjected directly to Sanger sequencing. TD reference sequences were shown on top of each Sequencing chromatogram. The sequences corresponding to each end of the TDs were marked by yellow and red boxes respectively. Sequences in green dashed boxes represent additive fragments.

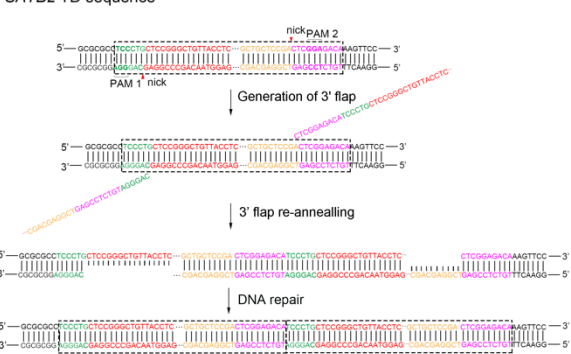
RYR1 TD sequence



NCF2 TD sequence



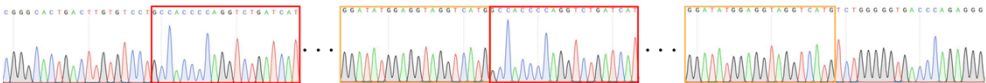
SATB2 TD sequence



Supplemental Fig.S29 PegRNAs design at three disease loci and the putative editing process of TD-PE strategy. PAM1 and PAM2 are the upstream and downstream PAMs respectively, and the red arrows indicate the positions of the nicks. Black dashed box indicates the sequences to be duplicated. Green and pink sequences denote the upstream and downstream additive fragments respectively.

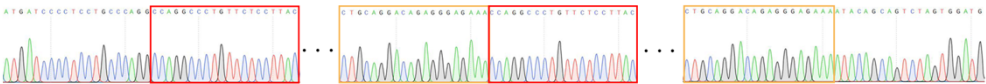
RYR1 TD 85-bp

TD reference sequence:CGGGCACTGACTTGTGTCCTGCCACCCCAAGGTCTGATCAT---GGATATGGAGGTAGGTCATGGCCACCCCAAGGTCTGATCAT---GGATATGGAGGTAGGTCATGCTGGGGGTGACCCAGAGGG



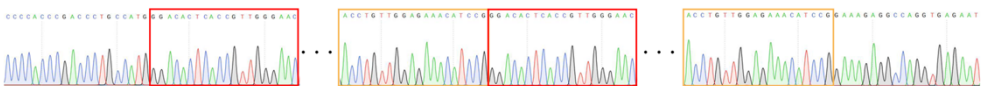
NCF2 TD 149-bp

TD reference sequence:ATGATCCCTCCTGCCAGGCCAGGCCCTGTTCTCCTTAC---CTGCAGGACAGAGGGAGAAACAGGCCCTGTTCTCCTTAC---CTGCAGGACAGAGGGAGAAATACAGCAGTCTAGTGGATG



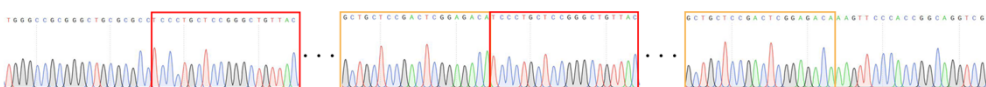
ENG TD 180-bp

TD reference sequence:CCCCACCGACCTGCCATGGGACACTCACCCTTGGGAAC---ACCTGTTGGAGAAACATCCGGACACTCACCCTTGGGAAC---ACCTGTTGGAGAAACATCCGGAAAGAGGCCAGGTGAGAAT



SATB2 TD 209-bp

TD reference sequence:TGGGCCGCGGGCTGCGCGCCTCCCTGCTCCGGGCTGTAC---GCTGCTCCGACTCGGAGACATCCCTGCTCCGGGCTGTAC---GCTGCTCCGACTCGGAGACAAGTTCCACCGGCAGGTGCG



Supplemental Fig.S30 Representative Sanger sequencing results of 4 disease-related TDs at indicated loci (related to Fig. 5B). The PCR amplicons containing targeted tandem duplication were gel-purified and subjected directly to Sanger sequencing. TD reference sequences were shown on top of each Sequencing result. The sequences corresponding to each end of the TDs were marked by red and yellow boxes respectively.

Supplemental Table 1. List of the targets tested in this study.

sgRNA	Target sequence	Oligo-F	Oligo-R	Reference
<i>FANCF</i>	GGAATCCCTTCTG CAGCACC TGG	CACCGGAATCCCTT CTGCAGCACC	AAACGGTGCTGCA GAAGGGATTCC	(Anzalone et al. 2019)
<i>FANCF</i> -50	GCCCTACTTCCGC TTTCACCT TGG	CACCGCCCTACTTC CGCTTTCACCT	AAACAGGTGAAAG CGGAAGTAGGG	(Anzalone et al. 2019)
<i>EMXI</i>	AGTCCGAGCAGA AGAAGAA GGG	CACCGAGTCCGAG CAGAAGAAGAA	AAACTTCTTCTTCT GCTCGGACTC	(Anzalone et al. 2019)
<i>EMXI</i> -57	CCGTTTGTACTTT GTCCTC CGG	CACCGCCGTTTGTA CTTTGTCCTC	AAACGAGGACAA AGTACAAACGGC	(Anzalone et al. 2019)
<i>FANCF</i> +115	GCAGAGAGTCGC CGTCTCCA AGG	CACCGCAGAGAGT CGCCGTCTCCA	AAACTGGAGACGG CGACTCTCTGC	This study
<i>FANCF</i> -115	TATCCGTGTTCCCT TGACTCT GGG	CACCGTATCCGTGT TCCTTGACTCT	AAACAGAGTCAAG GAACACGGATAC	This study
<i>FANCF</i> -171	TATCCGTGTTCCCT TGACTCT GGG	CACCGTATCCGTGT TCCTTGACTCT	AAACAGAGTCAAG GAACACGGATAC	This study
<i>FANCF</i> -231	AAAGAGGTTCTT CTTAGTGG AGG	CACCGAAAGAGGT TCTTCTTAGTGG	AAACCCACTAAGA AGAACCTCTTTC	This study
<i>EMXI</i> -116	GGGGCACAGATG AGAAACTC AGG	CACCGGGGCACAG ATGAGAAACTC	AAACGAGTTTCTC ATCTGTGCCCC	(Anzalone et al. 2019)
<i>EMXI</i> -201	ACTGAGGCTACA TAGGGTTA GGG	CACCGACTGAGGCT ACATAGGGTTA	AAACTAACCCTAT GTAGCCTCAGTC	This study
<i>EMXI</i> -307	CAGTGGGCCAGA GAGAAGGG TGG	CACCGCAGTGGGC CAGAGAGAAGGG	AAACCCCTTCTCT CTGGCCCACTGC	This study
<i>AAVSI</i>	GGGGCCACTAGG GACAGGAT TGG	CACCGGGGCCACT AGGGACAGGAT	AAACATCCTGTCC CTAGTGGCCCC	This study
<i>AAVSI</i> -376	CAGGGAGACGGG GTACTTTG GGG	CACCGCAGGGAGA CGGGGTACTTTG	AAACCAAAGTACC CCGTCTCCCTGC	This study
<i>AAVSI</i> -532	CAGCGAGTGAAG ACGGCATG GGG	CACCGCAGCGAGT GAAGACGGCATG	AAACCATGCCGTC TTCACCTCGCTGC	This study
<i>AAVSI</i> -846	AAGGGGAAACAG TGGGCCAG AGG	CACCGAAGGGGAA ACAGTGGGCCAG	AAACCTGGCCCAC TGTTTCCCCTTC	This study
<i>HEK3</i>	GGCCCAGACTGA GCACGTGA TGG	CACCGGCCAGACT GAGCACGTGA	AAACTCACGTGCT CAGTCTGGGCC	(Anzalone et al. 2019)
<i>HEK3</i> -136	TCCCTGGGATCCC TGTCTCC AGG	CACCGTCCCTGGGA TCCCTGTCTCC	AAACGGAGACAG GGATCCCAGGGAC	This study
<i>HEK3</i> -222	TCTCATGCCAAGC TCCCTGC AGG	CACCGTCTCATGCC AAGCTCCCTGC	AAACGCAGGGAGC TTGGCATGAGAC	This study

<i>HEK3</i> +1	TAAGACAATGGA TACACTGG AGG	CACCGTAAGACAA TGGATACACTGG	AAACCCAGTGTAT CCATTGTCTTAC	This study
<i>HEK3</i> -397	TCTGTTGAGCTCG ACCCTGA AGG	CACCGTCTGTTGAG CTCGACCCTGA	AAACTCAGGGTCG AGCTCAACAGAC	This study
<i>HEK3</i> -499	GTCAACCAGTAT CCCGGTGC AGG	CACCGTCAACCAGT ATCCCGGTGC	AAACGCACCGGGA TACTGGTTGAC	This study
<i>ENG</i> +1	GTTTCTGCAAGAC CTGTTGG AGA	CACCGTTTCTGCAA GACCTGTTGG	AAACCCAACAGGT CTTGCAGAAAC	This study
<i>ENG</i> -180	CTTCCTGGAGTTC CCAACGG TGA	CACCGCTTCCTGGA GTTCCCAACGG	AAACCCGTTGGGA ACTCCAGGAAG C	This study
<i>SATB2</i> +1	GCTCCATGCTGCT CCGACTC GGA	CACCGCTCCATGCT GCTCCGACTC	AAACGAGTCGGAG CAGCATGGAGC	This study
<i>SATB2</i> -209	GAGGTAACAGCC CGGAGCAG GGA	CACCGAGGTAACA GCCCCGAGCAG	AAACCTGCTCCGG GCTGTTACCTC	This study
<i>NCF2</i> +1	AAATCATATCTGC AGGACAG AGG	CACCGAAATCATAT CTGCAGGACAG	AAACCTGTCCTGC AGATATGATTTC	This study
<i>NCF2</i> -149	AGGAGAACAGGG CCTGGCCT GGG	CACCGAGGAGAAC AGGGCCTGGCCT	AAACAGGCCAGGC CCTGTTCTCCTC	This study
<i>RYR1</i> +1	TATGGAGGTAGG TCATGTCT GGG	CACCGTATGGAGGT AGGTCATGTCT	AAACAGACATGAC CTACCTCCATAC	This study
<i>RYR1</i> -85	AGCGTCGATGAT CAGACCTG GGG	CACCGAGCGTCGAT GATCAGACCTG	AAACCAGGTCTGA TCATCGACGCTC	This study

Supplemental Table 2. Sequences of pegRNAs used in mammalian cell experiments.

pegRNA	spacer	3' extension (5' to 3')	PBS length (nt)	HA length (nt)
<i>FANCF</i> +TD50 pegRNA	GGAATCCCTTCTGCAGCACC	CGCGAAGGCCCTACTTCCGCTTTCAGCT GCAGAAGGGA	13	25
<i>FANCF</i> -TD50 pegRNA	GCCCTACTTCCGCTTTCACCT	CACCTCATGGAATCCCTTCTGCAGCTGA AAGCGGAAGT	13	25
<i>EMX1</i> +TD57 pegRNA	GAGTCCGAGCAGAAGAAGA A	CTCCAGCTTCTGCCGTTTGTACTTTGTCT TCTTCTGCTCGG	13	28
<i>EMX1</i> -TD57 pegRNA	CCGTTTGTACTTTGTCCTC	GAGGAAGGGCCTGAGTCCGAGCAGAAG AAGACAAAGTACAAA	13	29
<i>FANCF</i> +TD115 pegRNA	GCAGAGAGTCGCCGTCTCCA	AGCGTCTTTATCCGTGTTCTTGACAGAC GGCGACTC	12	25
<i>FANCF</i> -TD115 pegRNA	TATCCGTGTTTCCTTGACTCT	ATCAGTACGCAGAGAGTCGCCGTCTGTC AAGGAACAC	12	25
<i>FANCF</i> +TD171 pegRNA	GGAATCCCTTCTGCAGCACC	AGCGTCTTTATCCGTGTTCTTGACGCTG CAGAAGGGA	13	25
<i>FANCF</i> -TD171 pegRNA	TATCCGTGTTTCCTTGACTCT	ACCTCATGGAATCCCTTCTGCAGCGTCA AGGAACACG	13	24
<i>FANCF</i> +TD231 pegRNA	GGAATCCCTTCTGCAGCACC	GCCACACAAAGAGGTTCTTCTTAGGCTG CAGAAGGGA	13	24
<i>FANCF</i> -TD231 pegRNA	AAAGAGGTTCTTCTTAGTGG	ACCTCATGGAATCCCTTCTGCAGC CTAAGAAGAACCT	13	24
<i>EMX1</i> +TD116 pegRNA	GAGTCCGAGCAGAAGAAGA A	GGGAGGGAGGGGCACAGATGAGAAATT CTTCTGCTCGG	13	25
<i>EMX1</i> -TD116 pegRNA	GGGGCACAGATGAGAAACTC	AAGGGCCTGAGTCCGAGCAGAAGAATTT CTCATCTGTG	13	25
<i>EMX1</i> +TD201 pegRNA	GAGTCCGAGCAGAAGAAGA A	ATGGGAAGACTGAGGCTACATAGGGTTC TTCTGCTCGG	13	25
<i>EMX1</i> -TD201 pegRNA	ACTGAGGCTACATAGGGTTA	AAGGGCCTGAGTCCGAGCAGAAGAACC CTATGTAGCCT	13	25
<i>EMX1</i> +TD307 pegRNA	GAGTCCGAGCAGAAGAAGA A	GATGGCAGGGCAGGAAGAGGACACAGT GGGCCAGAGAGAATTCTTCTGCTCGG	13	40
<i>EMX1</i> -TD307 pegRNA	CAGTGGGCCAGAGAGAAGG G	AGAAGCTGGAGGAGGAAGGGCCTGAGT CCGAGCAGAAGAATTCTCTTGCCCC	13	40
<i>AAVSI</i> +TD376 pegRNA	GGGGCCACTAGGGACAGGAT	GAGGATGGAGAGGTGGCTAAAGCCAGG GAGACGGGGTACTCTGTCCCTAGTGG	13	40
<i>AAVSI</i> -TD376 pegRNA	CAGGGAGACGGGGTACTTTG	ATCTGTCCCCTCCACCCACAGTGGGGC CACTAGGGACAGAGTACCCCGTCTC	13	40
<i>AAVSI</i> +TD532 pegRNA	GGGGCCACTAGGGACAGGAT	AGAAGGAGAAGGAAAAGGGAACCCAGC GAGTGAAGACGGCCTGTCCCTAGTGG	13	40
<i>AAVSI</i> -TD532 pegRNA	CAGCGAGTGAAGACGGCATG	ATCTGTCCCCTCCACCCACAGTGGGGC CACTAGGGACAG GCCGTCTTCACTC	13	40

<i>AAVSI</i> +TD846 pegRNA	GGGGCCACTAGGGACAGGAT	AGAGAAAGCAGGACCTGCCTGGGAAGG GGAAACAGTGGGCCTGTCCCTAGTGG	13	40
<i>AAVSI</i> -TD846 pegRNA	AAGGGGAAACAGTGGGGCA G	ATCTGTCCCCTCCACCCCACAGTGGGGC CACTAGGGACAGGGCCACTGTTTCC	13	40
<i>HEK3</i> +TD136 pegRNA	GGCCCAGACTGAGCACGTGA	TGGGCGTTTCCCTGGGATCCCTGTCCGTG CTCAGTCTG	13	25
<i>HEK3</i> -TD136 pegRNA	GTCCCTGGGATCCCTGTCTC C	ATCCTTGGGGCCCAGACTGAGCACGGAC AGGGATCCCA	13	23
<i>HEK3</i> +TD222 pegRNA	GGCCCAGACTGAGCACGTGA	AAGGTTTCTCATGCCAAGCTCCCCGTGC TCAGTCTG	13	23
<i>HEK3</i> -TD222 pegRNA	GTCTCATGCCAAGCTCCCTG C	ATCCTTGGGGCCCAGACTGAGCACGGGG AGCTTGGCAT	13	25
<i>HEK3</i> +TD397 pegRNA	TAAGACAATGGATACACTGG	TGGGTGCCCTGAGATCTTTTCTCTGTGTG AGCTCGACCCGTGTATCCATTGT	13	39
<i>HEK3</i> -TD397 pegRNA	TCTGTTGAGCTCGACCCTGA	TAATATCTCCCCATGTCCTACATAAGA CAATGGATACACGGGTCGAGCTCAA	13	40
<i>HEK3</i> +TD499 pegRNA	GTCAACCAGTATCCCGGTGC	TAATATCTCCCCATGTCCTACATAAGA CAATGGATACACCCGGGATACTGGT	13	40
<i>HEK3</i> -TD499 pegRNA	TAAGACAATGGATACACTGG	TGGCTTCCAGCCCAGCCAAACTTGTCAA CCAGTATCCCGGGTGTATCCATTGT	13	40
<i>EMX1</i> +TD5K pegRNA	GAGTCCGAGCAGAAGAAGA A	AGCAGGAAAGGACCCTCCAAATCCTCCT TTCCATGCCTCTTCTTCTGCTCGG	13	40
<i>EMX1</i> -TD5K pegRNA	CTCCTTTCCATGCCTCTGAG	AGAAGCTGGAGGAGGAAGGGCCTGAGT CCGAGCAGAAGAAAGAGGCATGGAAA	13	40
<i>EMX1</i> +TD10K pegRNA	GAGTCCGAGCAGAAGAAGA A	AGCAAATGTTGGCCAGCATAGGGACTCA GCTCCCACATGCTTCTTCTGCTCGG	13	40
<i>EMX1</i> -TD10K pegRNA	ACTCAGCTCCCACATGCAGG	AGAAGCTGGAGGAGGAAGGGCCTGAGT CCGAGCAGAAGAAGCATGTGGGAGCT	13	40
<i>AAVSI</i> +TD L2R2 pegRNA	GGGGCCACTAGGGACAGGAT	GAGGATGGAGAGGTGGCTAAAGCCAGG GAGACGGGGTACTTTTCTGTCCCTAGT GG	13	40
<i>AAVSI</i> -TD L2R2 pegRNA	CAGGGAGACGGGGTACTTTG	ATCTGTCCCCTCCACCCCACAGTGGGGC CACTAGGGACAGGAAAAGTACCCCGTCT C	13	40
<i>AAVSI</i> +TD L4R4 pegRNA	GGGGCCACTAGGGACAGGAT	GAGGATGGAGAGGTGGCTAAAGCCAGG GAGACGGGGTACTTTGGAATCCTGTCCC TAGTGG	13	40
<i>AAVSI</i> -TD L4R4 pegRNA	CAGGGAGACGGGGTACTTTG	ATCTGTCCCCTCCACCCCACAGTGGGGC CACTAGGGACAGGATTCCAAAGTACCCC GTCTC	13	40
<i>AAVSI</i> +TD L6R6 pegRNA	GGGGCCACTAGGGACAGGAT	GAGGATGGAGAGGTGGCTAAAGCCAGG GAGACGGGGTACTTTGGGGCCAATCCTG TCCCTAGTGG	13	40

<i>AAVSI</i> -TD L6R6 pegRNA	CAGGGAGACGGGGTACTTTG	ATCTGTCCCCTCCACCCCACAGTGGGGC CACTAGGGACAGGATTGGCCCCAAAGTA CCCCGTCTC	13	40
<i>AAVSI</i> +TD L8R8 pegRNA	GGGGCCACTAGGGACAGGAT	GAGGATGGAGAGGTGGCTAAAGCCAGG GAGACGGGGTACTTTGGGGTTACCAAT CCTGTCCCTAGTGG	13	40
<i>AAVSI</i> -TD L8R8 pegRNA	CAGGGAGACGGGGTACTTTG	ATCTGTCCCCTCCACCCCACAGTGGGGC CACTAGGGACAGGATTGGTGAACCCCAA AGTACCCCGTCTC	13	40
<i>AAVSI</i> +TD L10R10 pegRNA	GGGGCCACTAGGGACAGGAT	GAGGATGGAGAGGTGGCTAAAGCCAGG GAGACGGGGTACTTTGGGGTTGTGTCAC CAATCCTGTCCCTAGTGG	13	40
<i>AAVSI</i> -TD L10R10 pegRNA	CAGGGAGACGGGGTACTTTG	ATCTGTCCCCTCCACCCCACAGTGGGGC CACTAGGGACAGGATTGGTGACACAACC CCAAAGTACCCCGTCTC	13	40
<i>HEK3</i> +TD L2R2 pegRNA	GGCCCAGACTGAGCACGTGA	AAGGTTTCTCATGCCAAGCTCCCTGCAC GTGCTCAGTCTG	13	23
<i>HEK3</i> -TD L2R2 pegRNA	GTCTCATGCCAAGCTCCCTG C	ATCCTTGGGGCCCAGACTGAGCACGTGC AGGGAGCTTGGCAT	13	25
<i>HEK3</i> +TD L4R4 pegRNA	GGCCCAGACTGAGCACGTGA	AAGGTTTCTCATGCCAAGCTCCCTGCAA TCACGTGCTCAGTCTG	13	23
<i>HEK3</i> -TD L4R4 pegRNA	GTCTCATGCCAAGCTCCCTG C	ATCCTTGGGGCCCAGACTGAGCACGTGA TTGCAGGGAGCTTGGCAT	13	25
<i>HEK3</i> +TD L6R6 pegRNA	GGCCCAGACTGAGCACGTGA	AAGGTTTCTCATGCCAAGCTCCCTGCAG GCCATCACGTGCTCAGTCTG	13	23
<i>HEK3</i> -TD L6R6 pegRNA	GTCTCATGCCAAGCTCCCTG C	ATCCTTGGGGCCCAGACTGAGCACGTGA TGGCCTGCAGGGAGCTTGGCAT	13	25
<i>HEK3</i> +TD L8R8 pegRNA	GGCCCAGACTGAGCACGTGA	AAGGTTTCTCATGCCAAGCTCCCTGCAG GAGCCATCACGTGCTCAGTCTG	13	23
<i>HEK3</i> -TD L8R8 pegRNA	GTCTCATGCCAAGCTCCCTG C	ATCCTTGGGGCCCAGACTGAGCACGTGA TGGCAGTCCTGCAGGGAGCTTGGCAT	13	25
<i>HEK3</i> +TD L10R10 pegRNA	GGCCCAGACTGAGCACGTGA	AAGGTTTCTCATGCCAAGCTCCCTGCAG GACATTCTGCCATCACGTGCTCAGTCTG	13	23
<i>HEK3</i> -TD L10R10 pegRNA	GTCTCATGCCAAGCTCCCTG C	ATCCTTGGGGCCCAGACTGAGCACGTGA TGGCAGAATGTCCTGCAGGGAGCTTGGC AT	13	25
<i>ENG</i> +180 pegRNA	GTTTCTGCAAGACCTGTTGG	CATGTCCTCTTCCTGGAGTTCCCAACGGT GAGTGTCCCGGATGTTTCTCCAACAGGT CTTGCA	13	25
<i>ENG</i> -180 pegRNA	CTTCCTGGAGTTCCCAACGG	AATGGACTGTTTCTGCAAGACCTGTTGG AGAAACATCCGGGACACTACCGTTGGG AACTCCAG	13	25
<i>SATB2</i> +209 pegRNA	GCTCCATGCTGCTCCGACTC	GGCCGTGGGAGGTAACAGCCCGGAGCA GGGATGTCTCCGAGTCGGAGCAGCATG	13	25

<i>SATB2</i> -209	GAGGTAACAGCCCGGAGCA	GCTCCGCCGCTCCATGCTGCTCCGACTC	13	25
pegRNA	G	GGAGACATCCCTGCTCCGGGCTGTTA		
<i>NCF2</i> +149	AGGAGAACAGGGCCTGGCCT	GTGAGGTAAGGAGAACAGGGCCTGGTTT	13	25
pegRNA		CTCCCTCTGTCCTGCAGATATG		
<i>NCF2</i> -149	AAATCATATCTGCAGGACAG	TGATAGCCAAATCATATCTGCAGGACAG	13	25
pegRNA		AGGGAGAAACCAGGCCCTGTTC		
<i>RYR1</i> +85	TATGGAGGTAGGTCATGTCT	TCACCAAAGCGTCGATGATCAGACCTG	13	25
pegRNA		GGGTGGCCATGACCTACCTC		
<i>RYR1</i> -85	AGCGTCGATGATCAGACCTG	AAGGAGGATATGGAGGTAGGTCATGGC	13	25
pegRNA		CACCCCAGGTCTGATCATCGA		

Supplemental Table 3. Summary of primers for amplification of each target sites.

Target site	Forward primer	Reverse primer	Figures
<i>FANCF</i>	AAGACGCTGGGAGATTGACA	CAAAGCGCCGATGGATGTG	Fig. 1B
<i>EMX1</i>	TGGGGGCCTCCTGAGTTT	CTCTGCCCTCGTGGGTTTGT	Fig. 1B
<i>FANCF</i>	CATCATCTCGCACGTGGTTC	TGCTGACGTAGGTAGTGCTT	Fig. 2B
<i>EMX1</i>	GCTTCCAGAGCCTGCACTCC	TCGATGTCCTCCCCATTGGC	Fig. 2B
<i>AAVS1</i>	GAACTCTGCCCTCTAACGCT	AGAGGTTCTGGCAAGGAGAG	Fig. 2B
<i>HEK3</i>	GTGAGCTGCTGTCCCAGAGG	ATGCAGGTGCTGAAAGCCAC	Fig. 2B
<i>HEK3</i>	CACGTGATGGCAGAGGAAAG	GGCATTGTTATTTCCCTCCCA	Fig. 2B
<i>HEK3</i>	TTGATGAATCAGTGCTGGAG	GAGCTGCACATACTAGCC	Supplemental
TD136-bp			Fig. S22B
<i>EMX1</i> TD 5- kb	TGGGGGCCTCCTGAGTTT	AGCTCCTCACAGAACCACTC	Fig. 2D
<i>EMX1</i> TD 10- kb	TGGGGGCCTCCTGAGTTT	CAACACACACACTGGAGACT	Fig. 2D
<i>AAVS1</i>	CCTGTGCCATCTCTCGTTTC	AGAGGTTCTGGCAAGGAGAG	Fig. 3B
<i>ENG</i>	CTAACGAGGACTCAGCCACTG	AGTCCCTAGGAGACGTTTGGA	Fig. 3C
<i>SATB2</i>	TAATCCTACACCGCGACAGC	TGGCCTGAACGTCTATCACG	Fig. 3C
<i>NCF2</i>	CATGGTCCTTTGCGCTTCAC	GGAAGGGAGAAGCAGACTC	Fig. 3C
<i>RYR1</i>	GCAAGCCCTGGAGGTAGGTA	CACCCCCTTTCCTAACTGGC	Fig. 3C

Supplemental Table S4. Primers used for qPCR to detect the copy number of target DNA fragments.

Target site	Forward primer	Reverse primer
<i>EMXI</i> TD 5-kb/10-kb flanking (plasmid)	AGGGCTCCCATCACATCAA	ACGCGTTTGCTCTACCAGCC
<i>EMXI</i> TD 5-kb (plasmid)	CTGTGACCCTTTGTTTGAGA	CACCGTGAGCTAGGAACAAA
<i>EMXI</i> TD 10-kb (plasmid)	CTGCCATCCCCTTCTGTGAA	CTTCCCCTTTCTCCACCTCT
<i>EMXI</i> TD 5-kb/10-kb flanking	GGCCAATGGGGAGGACATC	TTGTCCCTCTGTCAATGGCG
<i>EMXI</i> TD 5-kb	GGCCTGAGTCCGAGCAGAA	TCCCCCTCTTATCCATCCCTC
<i>EMXI</i> TD 10-kb	CGAGCAGAAGAAGCATGTGGG	GTTTCTGCATGGGCGTGTG

Supplemental Table S5. Sequences of primers used for HTS.

sample name	Primer Name	Sequence
<i>FANCF</i> TD 50-bp	<i>FANCF</i> -HTS-1-forward	acacagtAGGCGTATCATTTGCGGGAT
	<i>FANCF</i> -HTS-reverse	AAAAGCGATCCAGGTGCTGC
<i>FANCF</i> TD 50-bp	<i>FANCF</i> -HTS-2-forward	acagtcaAGGCGTATCATTTGCGGGAT
	<i>FANCF</i> -HTS-reverse	AAAAGCGATCCAGGTGCTGC
<i>FANCF</i> TD 50-bp	<i>FANCF</i> -HTS-3-forward	actctgaAGGCGTATCATTTGCGGGAT
	<i>FANCF</i> -HTS-reverse	AAAAGCGATCCAGGTGCTGC
<i>FANCF</i> -CTR	<i>FANCF</i> -HTS-4-forward	gtagtctAGGCGTATCATTTGCGGGAT
	<i>FANCF</i> -HTS-reverse	AGGTAGCGCGCCCACTGCAA
<i>FANCF</i> -CTR	<i>FANCF</i> -HTS-5-forward	gtcacatAGGCGTATCATTTGCGGGAT
	<i>FANCF</i> -HTS-reverse	AGGTAGCGCGCCCACTGCAA
<i>FANCF</i> -CTR	<i>FANCF</i> -HTS-6-forward	gtctagtAGGCGTATCATTTGCGGGAT
	<i>FANCF</i> -HTS-reverse	AGGTAGCGCGCCCACTGCAA
<i>EMX1</i> TD 57-bp	<i>EMX1</i> -HTS-1-forward	catcagaTGGGGGCCTCCTGAGTTT
	<i>EMX1</i> -HTS-reverse	TGATGTGATGGGAGCCCTTC
<i>EMX1</i> TD 57-bp	<i>EMX1</i> -HTS-2-forward	catgtctTGGGGGCCTCCTGAGTTT
	<i>EMX1</i> -HTS-reverse	TGATGTGATGGGAGCCCTTC
<i>EMX1</i> TD 57-bp	<i>EMX1</i> -HTS-3-forward	cacaagtTGGGGGCCTCCTGAGTTT
	<i>EMX1</i> -HTS-reverse	TGATGTGATGGGAGCCCTTC
<i>EMX1</i> -CTR	<i>EMX1</i> -HTS-4-forward	acacgtaTGGGGGCCTCCTGAGTTT
	<i>EMX1</i> -HTS-reverse	TGATGTGATGGGAGCCCTTC
<i>EMX1</i> -CTR	<i>EMX1</i> -HTS-5-forward	acagatcTGGGGGCCTCCTGAGTTT
	<i>EMX1</i> -HTS-reverse	TGATGTGATGGGAGCCCTTC
<i>EMX1</i> -CTR	<i>EMX1</i> -HTS-6-forward	acagcatTGGGGGCCTCCTGAGTTT
	<i>EMX1</i> -HTS-reverse	TGATGTGATGGGAGCCCTTC
<i>AAVSI</i> TD 376-bp	<i>AAVSI</i> TD-376 -HTS -1-forward	cttgagtATTCCCAGGGCCGGTTAATG
	<i>AAVSI</i> TD-376 -reverse	GTGACCCGAATCCACAGGAG
<i>AAVSI</i> TD 376-bp	<i>AAVSI</i> TD-376 -HTS -2-forward	ctcacgaATTCCCAGGGCCGGTTAATG
	<i>AAVSI</i> TD-376 -reverse	GTGACCCGAATCCACAGGAG
<i>AAVSI</i> TD 376-bp	<i>AAVSI</i> TD-376 -HTS -3-forward	ctcagacATTCCCAGGGCCGGTTAATG
	<i>AAVSI</i> TD-376 -reverse	GTGACCCGAATCCACAGGAG
<i>AAVSI</i> TD 532-bp	<i>AAVSI</i> TD-532 -HTS -1-forward	ctctcagATTCCCAGGGCCGGTTAATG
	<i>AAVSI</i> TD-532 -reverse	GAGACATCCGTCGGAGAAGG
<i>AAVSI</i> TD 532-bp	<i>AAVSI</i> TD-532 -HTS -2-forward	ctctgtaATTCCCAGGGCCGGTTAATG
	<i>AAVSI</i> TD-532 -reverse	GAGACATCCGTCGGAGAAGG
<i>AAVSI</i> TD 532-bp	<i>AAVSI</i> TD-532 -HTS -3-forward	ctgatgtATTCCCAGGGCCGGTTAATG
	<i>AAVSI</i> TD-532 -reverse	GAGACATCCGTCGGAGAAGG
<i>AAVSI</i> TD 846-bp	<i>AAVSI</i> TD-846-HTS -1-forward	ctgtagaATTCCCAGGGCCGGTTAATG
	<i>AAVSI</i> TD-846 -reverse	CAAGCTGCAGACAGAAAGCG
<i>AAVSI</i> TD 846-bp	<i>AAVSI</i> TD-846 -HTS -2-forward	gaacactATTCCCAGGGCCGGTTAATG
	<i>AAVSI</i> TD-846 -reverse	CAAGCTGCAGACAGAAAGCG
<i>AAVSI</i> TD 846-bp	<i>AAVSI</i> TD-846 -HTS -3-forward	gatctcaATTCCCAGGGCCGGTTAATG
	<i>AAVSI</i> TD-846-reverse	CAAGCTGCAGACAGAAAGCG

<i>EMXI</i> TD 116-bp	<i>EMXI</i> TD-116-HTS -1-forward	cacactgTCCAGAACCGGAGGACAAAG
	<i>EMXI</i> TD-116-reverse	ACCACACCTTCACCTGGG
<i>EMXI</i> TD 116-bp	<i>EMXI</i> TD-116-HTS -2-forward	cacttgaTCCAGAACCGGAGGACAAAG
	<i>EMXI</i> TD-116-reverse	ACCACACCTTCACCTGGG
<i>EMXI</i> TD 116-bp	<i>EMXI</i> TD-116-HTS -3-forward	cactgatTCCAGAACCGGAGGACAAAG
	<i>EMXI</i> TD-116-reverse	ACCACACCTTCACCTGGG
<i>EMXI</i> TD 201-bp	<i>EMXI</i> TD-201-HTS -1-forward	cagatcaTGGGGGCCTCCTGAGTTT
	<i>EMXI</i> TD-201-reverse	CCTCAACACTCAGGCTGAGC
<i>EMXI</i> TD 201-bp	<i>EMXI</i> TD-201-HTS -2-forward	cagacatTGGGGGCCTCCTGAGTTT
	<i>EMXI</i> TD-201-reverse	CCTCAACACTCAGGCTGAGC
<i>EMXI</i> TD 201-bp	<i>EMXI</i> TD-201-HTS -3-forward	cagagtcTGGGGGCCTCCTGAGTTT
	<i>EMXI</i> TD-201-reverse	CCTCAACACTCAGGCTGAGC
<i>EMXI</i> TD 307-bp	<i>EMXI</i> TD-307-HTS -1-forward	cagtactGTTTCTCATCTGTGCCCTCC
	<i>EMXI</i> TD-307-reverse	GCTCCCATGGGTCTAACATTC
<i>EMXI</i> TD 307-bp	<i>EMXI</i> TD-307-HTS -2-forward	cagtctaGTTTCTCATCTGTGCCCTCC
	<i>EMXI</i> TD-307-reverse	GCTCCCATGGGTCTAACATTC
<i>EMXI</i> TD 307-bp	<i>EMXI</i> TD-307-HTS -3-forward	ctagacaGTTTCTCATCTGTGCCCTCC
	<i>EMXI</i> TD-307-reverse	GCTCCCATGGGTCTAACATTC
<i>FANCF</i> TD 115-bp	<i>FANCF</i> TD-115-HTS -1-forward	cagtactATAGCATTGCAGAGAGGCGT
	<i>FANCF</i> TD-115-reverse	GGTCGAAATGCATGTCAATC
<i>FANCF</i> TD 115-bp	<i>FANCF</i> TD-115-HTS -1-forward	cagtactATAGCATTGCAGAGAGGCGT
	<i>FANCF</i> TD-115-reverse	GGTCGAAATGCATGTCAATC
<i>FANCF</i> TD 115-bp	<i>FANCF</i> TD-115-HTS -1-forward	cagtactATAGCATTGCAGAGAGGCGT
	<i>FANCF</i> TD-115-reverse	GGTCGAAATGCATGTCAATC
<i>FANCF</i> TD 171-bp	<i>FANCF</i> TD-171-HTS -1-forward	tgtcagtATAGCATTGCAGAGAGGCGT
	<i>FANCF</i> TD-171-reverse	GGTCGAAATGCATGTCAATC
<i>FANCF</i> TD 171-bp	<i>FANCF</i> TD-171-HTS -1-forward	tggtgcaATAGCATTGCAGAGAGGCGT
	<i>FANCF</i> TD-171-reverse	GGTCGAAATGCATGTCAATC
<i>FANCF</i> TD 171-bp	<i>FANCF</i> TD-171-HTS -1-forward	caactgtATAGCATTGCAGAGAGGCGT
	<i>FANCF</i> TD-171-reverse	GGTCGAAATGCATGTCAATC
<i>FANCF</i> TD 231-bp	<i>FANCF</i> TD-231-HTS -1-forward	catcagaCGCTGGGAGATTGACATGCA
	<i>FANCF</i> TD-231-reverse	CCGTGTTTCCTTGACTCTGGG
<i>FANCF</i> TD 231-bp	<i>FANCF</i> TD-231-HTS -1-forward	catgtctCGCTGGGAGATTGACATGCA
	<i>FANCF</i> TD-231-reverse	CCGTGTTTCCTTGACTCTGGG
<i>FANCF</i> TD 231-bp	<i>FANCF</i> TD-231-HTS -1-forward	cacaagtCGCTGGGAGATTGACATGCA
	<i>FANCF</i> TD-231-reverse	CCGTGTTTCCTTGACTCTGGG
<i>HEK3</i> TD 136-bp	<i>HEK3</i> TD-136-HTS -1-forward	gacatctGCATTTGTAGGCTTGATGCT
	<i>HEK3</i> TD-136-reverse	ATGCTTACTTGCAGCAGAAA
<i>HEK3</i> TD 136-bp	<i>HEK3</i> TD-136-HTS -2-forward	gacagtaGCATTTGTAGGCTTGATGCT
	<i>HEK3</i> TD-136-reverse	ATGCTTACTTGCAGCAGAAA
<i>HEK3</i> TD 136-bp	<i>HEK3</i> TD-136-HTS -3-forward	gactacaGCATTTGTAGGCTTGATGCT
	<i>HEK3</i> TD-136-reverse	ATGCTTACTTGCAGCAGAAA
<i>HEK3</i> TD 222-bp	<i>HEK3</i> TD-222-HTS -1-forward	gactcacGCATTTGTAGGCTTGATGCT
	<i>HEK3</i> TD-222-reverse	ATGCTTACTTGCAGCAGAAA
<i>HEK3</i> TD 222-bp	<i>HEK3</i> TD-222-HTS -2-forward	gagtcgtGCATTTGTAGGCTTGATGCT

	<i>HEK3</i> TD-222-reverse	ATGCTTACTTGCAGCAGAAA
<i>HEK3</i> TD 222-bp	<i>HEK3</i> TD-222-HTS -3-forward	gtacagaGCATTGTAGGCTTGATGCT
	<i>HEK3</i> TD-222-reverse	ATGCTTACTTGCAGCAGAAA
<i>HEK3</i> TD 397-bp	<i>HEK3</i> TD-397-HTS -1-forward	gtagtctTGAGCACTTGTTACACGCAG
	<i>HEK3</i> TD-397-reverse	CTGATTTTCATGCAGGTGCT
<i>HEK3</i> TD 397-bp	<i>HEK3</i> TD-397-HTS -2-forward	gtcacatTGAGCACTTGTTACACGCAG
	<i>HEK3</i> TD-397-reverse	CTGATTTTCATGCAGGTGCT
<i>HEK3</i> TD 397-bp	<i>HEK3</i> TD-397-HTS -3-forward	gtctagtTGAGCACTTGTTACACGCAG
	<i>HEK3</i> TD-397-reverse	CTGATTTTCATGCAGGTGCT
<i>HEK3</i> TD 499-bp	<i>HEK3</i> TD-499-HTS -1-forward	gtcgtgaGTTACACGCAGGGCACTATG
	<i>HEK3</i> TD-499-reverse	TGTTGAGCTCGACCCTGAAG
<i>HEK3</i> TD 499-bp	<i>HEK3</i> TD-499-HTS -2-forward	gtgaacaGTTACACGCAGGGCACTATG
	<i>HEK3</i> TD-499-reverse	TGTTGAGCTCGACCCTGAAG
<i>HEK3</i> TD 499-bp	<i>HEK3</i> TD-499-HTS -3-forward	gtgactcGTTACACGCAGGGCACTATG
	<i>HEK3</i> TD-499-reverse	TGTTGAGCTCGACCCTGAAG
<i>EMX1</i> TD 5-kb	<i>EMX1</i> TD 5kb-HTS-1-forward	atctgctTCCAGAACCGGAGGACAAAG
	<i>EMX1</i> TD 5kb-reverse	TCCCCCTCTTATCCATCCCTC
<i>EMX1</i> TD 5-kb	<i>EMX1</i> TD 5kb-HTS-2-forward	atgtgacTCCAGAACCGGAGGACAAAG
	<i>EMX1</i> TD 5kb-reverse	TCCCCCTCTTATCCATCCCTC
<i>EMX1</i> TD 5-kb	<i>EMX1</i> TD 5kb-HTS-3-forward	atgcactTCCAGAACCGGAGGACAAAG
	<i>EMX1</i> TD 5kb-reverse	TCCCCCTCTTATCCATCCCTC
<i>EMX1</i> TD 10-kb	<i>EMX1</i> TD 10kb-HTS-1-forward	acaactgGTTCCAGAACCGGAGGACAAA
	<i>EMX1</i> TD 10kb-reverse	CCAGCCTCCCTACTGTCTTTTT
<i>EMX1</i> TD 10-kb	<i>EMX1</i> TD 10kb-HTS-2-forward	acatcgaGTTCCAGAACCGGAGGACAAA
	<i>EMX1</i> TD 10kb-reverse	CCAGCCTCCCTACTGTCTTTTT
<i>EMX1</i> TD 10-kb	<i>EMX1</i> TD 10kb-HTS-3-forward	acatgagGTTCCAGAACCGGAGGACAAA
	<i>EMX1</i> TD 10kb-reverse	CCAGCCTCCCTACTGTCTTTTT

Supplemental Reference

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