

Supplemental information

Targeted, programmable and precise tandem duplication in mammalian genome

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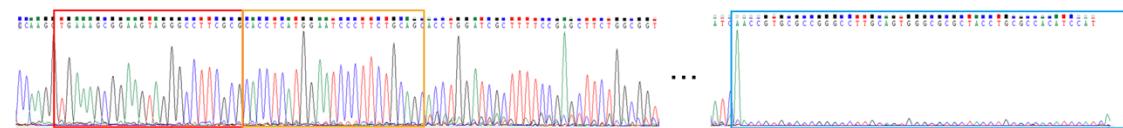
FANCF TD 50-bp

Wild-type sequence:

PAM1 nick PAM2
CCAAGG **TGAAAGCGGAAGTAGGGCCTCGCGCACCTCATGGAATCCCTCTGCAGC** ACCTGGATCGCTTTTC

Reference sequence:

CCAAGG **TGAAAGCGGAAGTAGGGCCTCGCGCACCTCATGGAATCCCTCTGCAGCTGAAAGCGGAAGTAGGGCCTCGCGCACCTCATGGAATCCCTCTGCAGCACCTGGATCGCTTTTC**



Supplemental Fig. S1 Representative Sanger sequencing results of TD-PE mediated 50-bp tandem duplication at *FANCF* site. PCR amplicons containing targeted tandem duplication was purified and directly subjected to Sanger sequencing. Wild-type sequence and TD reference sequence were shown on top of the sequencing chromatogram. PAM1 and PAM2 of the two pegRNAs were shown in red and yellow respectively. Red arrows indicated the positions of the nicks. The sequences in-between two nicks are the duplicated sequences, with the upstream end shown in red and the downstream end shown in yellow. The peaks in the sequencing chromatogram corresponding to each end were boxed in red and yellow respectively. The blue box marked the fragment that is located at the end of the double peaks. The length of the fragment is the same as that of the duplicated region, and the sequence corresponds to the 3' end of the amplicons that contain TDs.

FANCF TD 50-bp

Wild-type sequence :	PAM1	nick	PAM2	----- Cleavage - Deletion □ Insertion
	CTCCA	AGGTGAAAGCGGAAGTAGGGCCTTCGGCACCTCATGGAA	TCTCGGACCCCTGGATCGCTTT	reads / total reads
2 × TD (Accurate) :	CTCCA	AGGTGAAAGCGGAAGTAGGGCCTTCGGCACCTCATGGAA	TCTCGGACCCCTGGATCGCTTT	56314 / 221396 (25.44%)
2 × TD with insertion :	CTCCA	AGGTGAAAGCGGAAGTAGGGCGGAAGTAGGGCCTTCGGCACCTCATGGAA	TCTCGGACCCCTGGATCGCTTT	1171 / 221396 (0.53%)
		11-bp		
	CTCCA	AGGTGAAAGCGGAAGTAGGGCCTTCGGCACCTCATGGAA	TCTCGGACCCCTGGATCGCTTT	841 / 221396 (0.38%)
		13-bp		
	CTCCA	AGGTGAAAGCGGAAGTAGGGCCTTCGGCACCTCATGGAA	TCTCGGACCCCTGGATCGCTTT	405 / 221396 (0.18%)
		7-bp		
	CTCCA	AGGTGAAAGCGGAAGTAGGGCCTTCGGCACCTCATGGAA	TCTCGGACCCCTGGATCGCTTT	340 / 221396 (0.15%)
		11-bp		
	CTCCA	AGGTGAAAGCGGAAGTAGGGCCTTCGGCACCTCATGGAA	TCTCGGACCCCTGGATCGCTTT	319 / 221396 (0.14%)
		12-bp		
	CTCCA	AGGTGAAAGCGGAAGTAGGGCCTTCGGCACCTCATGGAA	TCTCGGACCCCTGGATCGCTTT	282 / 221396 (0.13%)
		6-bp		
	CTCCA	AGGTGAAAGCGGAAGTAGGGCCTTCGGCACCTCATGGAA	TCTCGGACCCCTGGATCGCTTT	218 / 221396 (0.098%)
		12-bp		
	CTCCA	AGGTGAAAGCGGAAGTAGGGCCTTCGGCACCTCATGGAA	TCTCGGACCCCTGGATCGCTTT	213 / 221396 (0.096%)
		6-bp		
	CTCCA	AGGTGAAAGCGGAAGTAGGGCCTTCGGCACCTCATGGAA	TCTCGGACCCCTGGATCGCTTT	173 / 221396 (0.078%)
		10-bp		
	CTCCA	AGGTGAAAGCGGAAGTAGGGCCTTCGGCACCTCATGGAA	TCTCGGACCCCTGGATCGCTTT	160 / 221396 (0.072%)
		7-bp		
	CTCCA	AGGTGAAAGCGGAAGTAGGGCCTTCGGCACCTCATGGAA	TCTCGGACCCCTGGATCGCTTT	129 / 221396 (0.058%)
		6-bp		
	CTCCA	AGGTGAAAGCGGAAGTAGGGCCTTCGGCACCTCATGGAA	TCTCGGACCCCTGGATCGCTTT	111 / 221396 (0.050%)
		9-bp		
	CTCCA	AGGTGAAAGCGGAAGTAGGGCCTTCGGCACCTCATGGAA	TCTCGGACCCCTGGATCGCTTT	102 / 221396 (0.046%)
		7-bp		
	CTCCA	AGGTGAAAGCGGAAGTAGGGCCTTCGGCACCTCATGGAA	TCTCGGACCCCTGGATCGCTTT	99 / 221396 (0.045%)
		10-bp		
	CTCCA	AGGTGAAAGCGGAAGTAGGGCCTTCGGCACCTCATGGAA	TCTCGGACCCCTGGATCGCTTT	96 / 221396 (0.043%)
		17-bp		
	CTCCA	AGGTGAAAGCGGAAGTAGGGCCTTCGGCACCTCATGGAA	TCTCGGACCCCTGGATCGCTTT	92 / 221396 (0.042%)
		7-bp		
	CTCCA	AGGTGAAAGCGGAAGTAGGGCCTTCGGCACCTCATGGAA	TCTCGGACCCCTGGATCGCTTT	88 / 221396 (0.040%)
		10-bp		
	CTCCA	AGGTGAAAGCGGAAGTAGGGCCTTCGGCACCTCATGGAA	TCTCGGACCCCTGGATCGCTTT	80 / 221396 (0.036%)
		7-bp		
	CTCCA	AGGTGAAAGCGGAAGTAGGGCCTTCGGCACCTCATGGAA	TCTCGGACCCCTGGATCGCTTT	56 / 221396 (0.025%)
2 × TD with deletion :	CTCCA	AGGTGAAAGCGGAAGTAGGGCCTTCGGCACCTCATGGAA	TCTCGGACCCCTGGATCGCTTT	945 / 221396 (0.43%)

	CTCCA	AGGTGAAAGCGGAAGTAGGGCCTTCGGCACCTCATGGAA	TCTCGGACCCCTGGATCGCTTT	596 / 221396 (0.27%)

	CTCCA	AGGTGAAAGCGGAAGTAGGGCCTTCGGCACCTCATGGAA	TCTCGGACCCCTGGATCGCTTT	79 / 221396 (0.036%)

	CTCCA	AGGTGAAAGCGGAAGTAGGGCCTTCGGCACCTCATGGAA	TCTCGGACCCCTGGATCGCTTT	76 / 221396 (0.034%)

	CTCCA	AGGTGAAAGCGGAAGTAGGGCCTTCGGCACCTCATGGAA	TCTCGGACCCCTGGATCGCTTT	58 / 221396 (0.026%)
3 × TD (Accurate) :	CTCCA	AGGTGAAAGCGGAAGTAGGGCCTTCGGCACCTCATGGAA	TCTCGGACCCCTGGATCGCTTT	18718 / 221396 (8.45%)
		11-bp		
	CTCCA	AGGTGAAAGCGGAAGTAGGGCCTTCGGCACCTCATGGAA	TCTCGGACCCCTGGATCGCTTT	110 / 221396 (0.050%)
		13-bp		
	CTCCA	AGGTGAAAGCGGAAGTAGGGCCTTCGGCACCTCATGGAA	TCTCGGACCCCTGGATCGCTTT	108 / 221396 (0.049%)
		11-bp		
	CTCCA	AGGTGAAAGCGGAAGTAGGGCCTTCGGCACCTCATGGAA	TCTCGGACCCCTGGATCGCTTT	66 / 221396 (0.030%)
		22-bp		
	CTCCA	AGGTGAAAGCGGAAGTAGGGCCTTCGGCACCTCATGGAA	TCTCGGACCCCTGGATCGCTTT	54 / 221396 (0.024%)
3 × TD with deletion :	CTCCA	AGGTGAAAGCGGAAGTAGGGCCTTCGGCACCTCATGGAA	TCTCGGACCCCTGGATCGCTTT	103 / 221396 (0.047%)

	CTCCA	AGGTGAAAGCGGAAGTAGGGCCTTCGGCACCTCATGGAA	TCTCGGACCCCTGGATCGCTTT	93 / 221396 (0.042%)

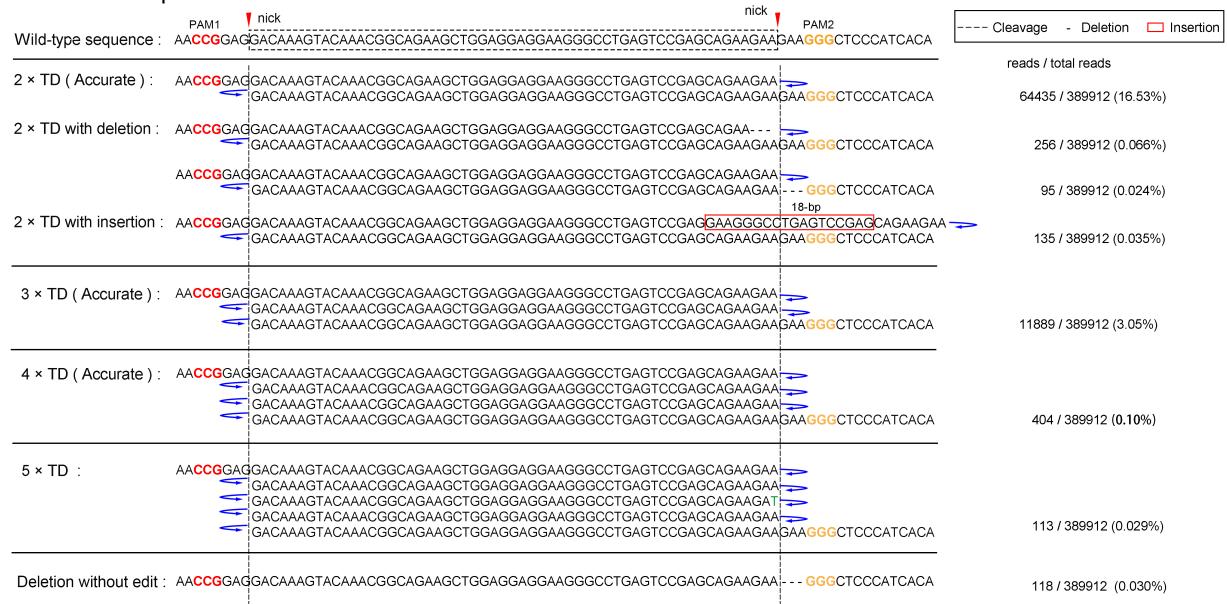
	CTCCA	AGGTGAAAGCGGAAGTAGGGCCTTCGGCACCTCATGGAA	TCTCGGACCCCTGGATCGCTTT	81 / 221396 (0.037%)
4 × TD (Accurate) :	CTCCA	AGGTGAAAGCGGAAGTAGGGCCTTCGGCACCTCATGGAA	TCTCGGACCCCTGGATCGCTTT	4559 / 221396 (2.06%)

	CTCCA	AGGTGAAAGCGGAAGTAGGGCCTTCGGCACCTCATGGAA	TCTCGGACCCCTGGATCGCTTT	77 / 221396 (0.035%)

6 × TD :	CTCCA	AGGTGAAAGCGGAAGTAGGGCCTTCGGCACCTCATGGAA	TCTCGGACCCCTGGATCGCTTT	53 / 221396 (0.024%)

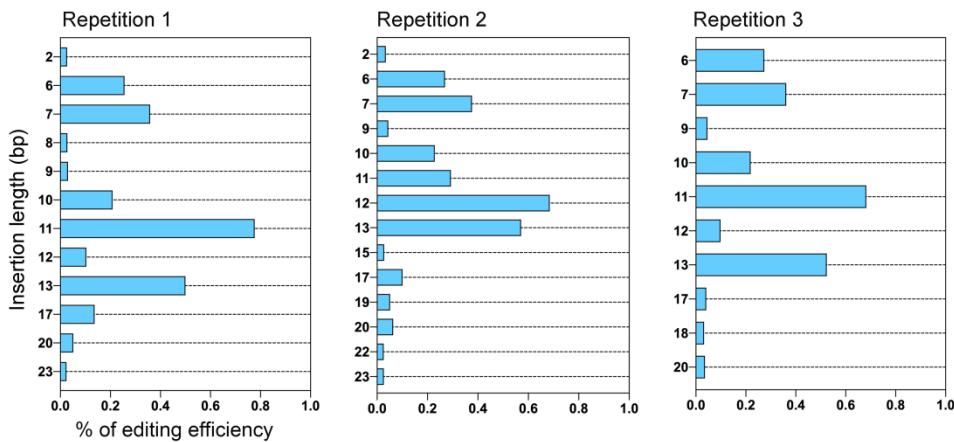
Supplemental Fig. S2 HTS sequences showing the editing result of TD-PE at the *FANCF* site. The fragments containing tandem duplications were amplified by HTS primers and subjected for HTS analysis. Detailed analysis of the undesired TDs revealed that the majority of deletions and insertions occurred at the boundaries of the duplications. The blue curved arrows pointed to the next copy of the tandem duplication. Deletion, insertion and base substitution were indicated by black dash, red box and green letters respectively. The insertion sequence of the red box was consistent with its 5' upstream sequence. Note that to provide an overview of the TDs, we copied the sequences of 2×TD from Fig. 2E to this section.

EMX1 TD 57-bp

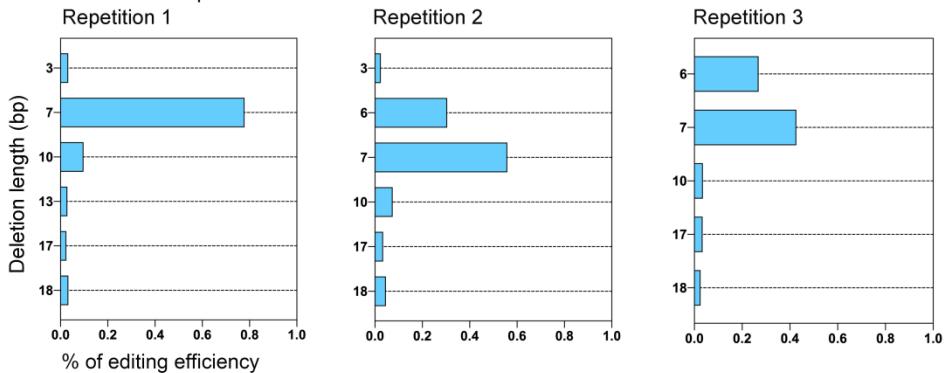


Supplemental Fig. S3 HTS sequences showing the editing result of TD-PE at the *EMX1* site. The fragments containing tandem duplication were amplified by HTS primers, purified and then subjected for HTS. The blue curved arrows pointed to the next copy of the tandem duplication. Deletion, insertion and base substitution were indicated by black dash, red box or green letters respectively. The insertion sequence of the red box was consistent with its 5' upstream sequence.

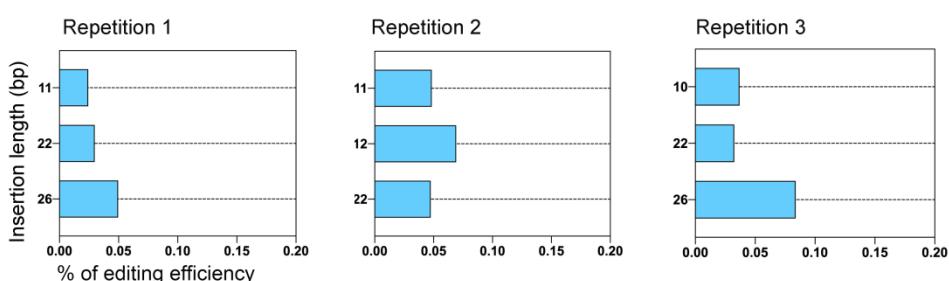
FANCF TD 50-bp 2×TD insertion



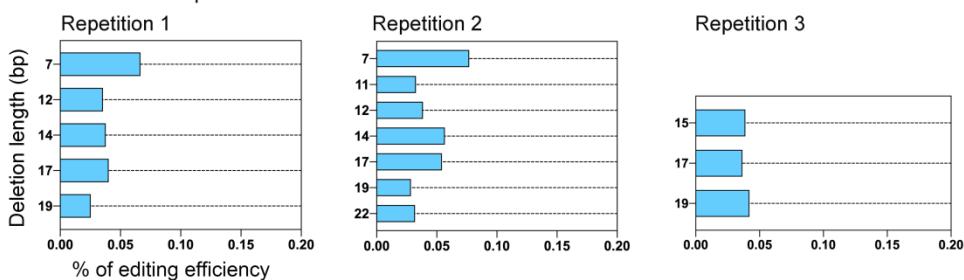
FANCF TD 50-bp 2×TD deletion



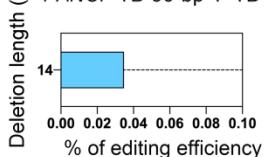
FANCF TD 50-bp 3×TD insertion



FANCF TD 50-bp 3×TD deletion

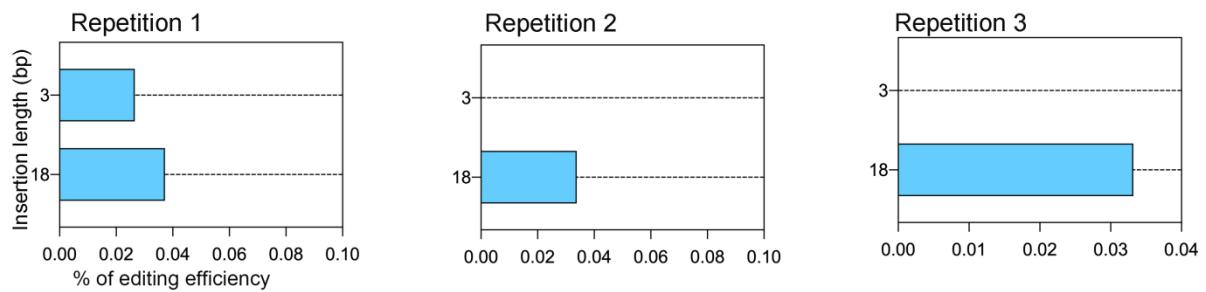


FANCF TD 50-bp 4×TD deletion

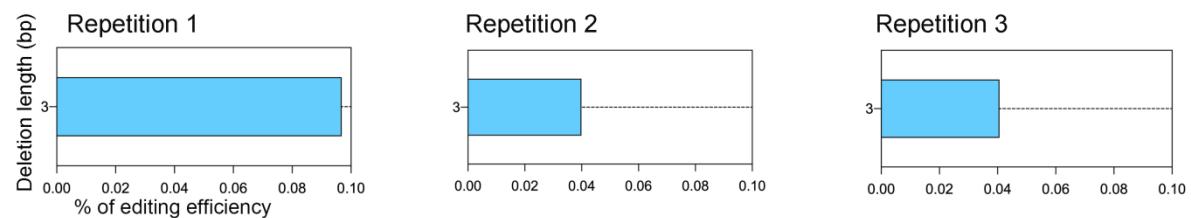


Supplemental Fig. S4 Distribution of the insertion or deletion lengths at *FANCF* site. HTS analysis of the frequencies of undesired edits induced by TD-PE at *FANCF* site (TD 50-bp). The experiment was repeated three times, and the indel data from each repetition were presented separately. Note that the deletion in 4×TD was detectable in only one out of three repetitions.

EMX1 TD 57-bp 2×TD insertion



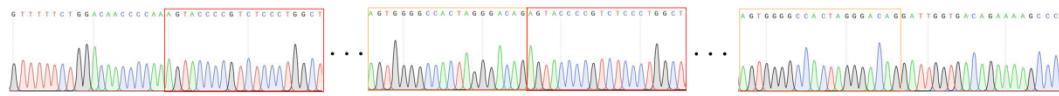
EMX1 TD 57-bp 2×TD deletion



Supplemental Fig. S5 Distribution of the insertion or deletion lengths at *EMX1* site. HTS analysis of the frequencies of undesired edits induced by TD-PE at *EMX1* site (TD 57-bp). The experiment was repeated three times, and the indel data from each repetition were presented separately.

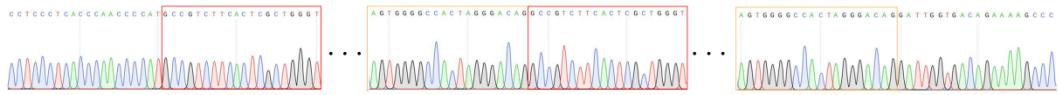
AAVS1 TD 376-bp 2×TD

TD reference sequence: GTTTTCTGGACAACCCAAAGTACCCCGTCTCCCTGGCT...AGTGGGGCCACTAGGGACAGAGTACCCCGTCTCCCTGGCT...AGTGGGGCCACTAGGGACAGGATTGGTACAGAAAAGCCC



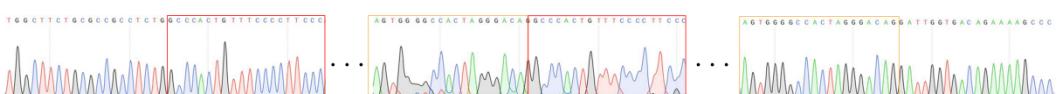
AAVS1 TD 532-bp 2×TD

TD reference sequence: CCTCCCTCACCAACCCATGCCGCTTTCACTCGCTGGT...AGTGGGGCCACTAGGGACAGGCCGCTTCACTCGCTGGT...AGTGGGGCCACTAGGGACAGGATTGGTACAGAAAAGCCC



AAVS1 TD 846-bp 2×TD

TD reference sequence: TGGCTTCTGGCCGCCCTCTGGCCCACTGTTCCCCCTCCC...AGTGGGGCCACTAGGGACAGGCCACTGTTCCCCCTCCC...AGTGGGGCCACTAGGGACAGGATTGGTACAGAAAAGCCC



Supplemental Fig. S6 Representative Sanger sequencing results of large fragment TDs at the *AAVS1* site (related to Fig. 3A). Representative Sanger sequencing results of TD containing amplicons derived from 3 targeted TDs at the *AAVS1* site. The amplicons containing targeted TDs were gel-purified and subjected directly to Sanger sequencing. TD reference sequence was shown on top of each sequencing chromatogram, with sequences corresponding to each end of the TD boxed in red and yellow respectively.

EMX1 TD 116-bp 2×TD

TD reference sequence: TGCTCTGGGGGCCCTCGAGTTCTCATCTGTGCCCTCC--CCTGAGTCGAGCAGAAGAA

TTTCTCATCTGTGCCCTCC--CCTGAGTCGAGCAGAAGAA

GA

AGGGCTCCCATCACATCA

EMX1 TD 116-bp 3×TD

TD reference sequence: TGCTCTGGGGGCCCTCGAGTTCTCATCTGTGCCCTCC--CCTGAGTCGAGCAGAAGAA

TTTCTCATCTGTGCCCTCC--CCTGAGTCGAGCAGAAGAA

GA

AGGGCTCCCATCACATCA

EMX1 TD 116-bp 4×TD

TD reference sequence: TGCTCTGGGGGCCCTCGAGTTCTCATCTGTGCCCTCC--CCTGAGTCGAGCAGAAGAA

TTTCTCATCTGTGCCCTCC--CCTGAGTCGAGCAGAAGAA

GA

AGGGCTCCCATCACATCA

EMX1 TD 201-bp 2×TD

TD reference sequence: CCCGGCCCTGGGCCCTAAC

CCCTATGTAGCTCAAGCTT--CCTGAGTCGAGCAGAAGAA

GA

AGGGCTCCCATCACATCA

EMX1 TD 307-bp 2×TD

TD reference sequence: GAGGAACAAGAAACACCC

TTCTCTCTGGCCCACTGT--CCTGAGTCGAGCAGAAGAA

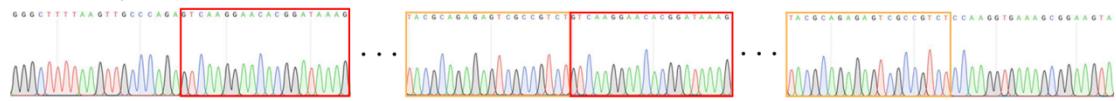
GA

AGGGCTCCCATCACATCA

Supplemental Fig. S7 Representative Sanger sequencing results of large fragment TDs at the *EMX1* site (related to Fig. 3A). Representative Sanger sequencing results of TD containing amplicons derived from 3 targeted TDs at *EMX1* site. The amplicons containing targeted TDs were gel-purified and subjected to Sanger sequencing. TD reference sequence was shown on top of each sequencing chromatogram, with sequences corresponding to each end of the TD boxed in red and yellow respectively. *EMX1* TD 116-bp 3×TD and 4×TD were the sequencing results of the bands marked red stars (*) at the *EMX1* site in Fig. 3A.

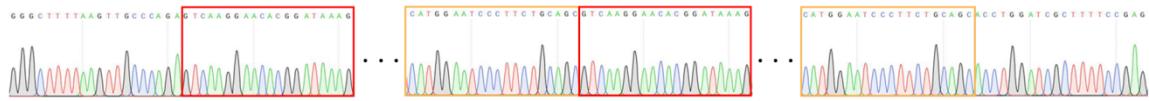
FANCF TD 115-bp 2×TD

TD reference sequence: GGGCTTTAAGTTGCCAGA**GTCAAGGAACACGGATAAG**...TACGCAGAGTCGCCGT**GTCAAGGAACACGGATAAG**...TACGCAGAGTCGCCGT**GTCAAGGTGAAAGCGGAAGTA**



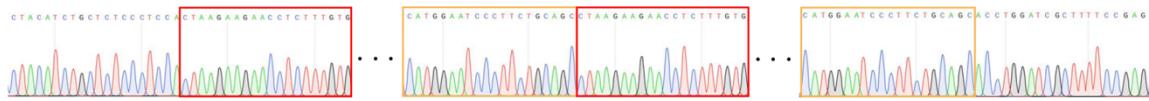
FANCF TD 171-bp 2×TD

TD reference sequence: GGGCTTTAAGTTGCCAGA**GTCAAGGAACACGGATAAG**...CATGGAATCCCTCTGCAGC**GTCAAGGAACACGGATAAG**...CATGGAATCCCTCTGCAGC**ACCTGGATCGCTTCCGAG**



FANCF TD 231-bp 2×TD

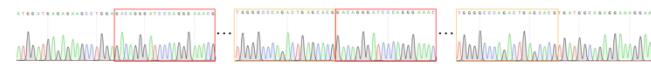
TD reference sequence: CTACATCTGCTCTCCCTCCA**CTAAGAAGAACCTCTTGTG**...CATGGAATCCCTCTGCAGC**CTAAGAAGAACCTCTTGTG**...CATGGAATCCCTCTGCAGC**ACCTGGATCGCTTCCGAG**



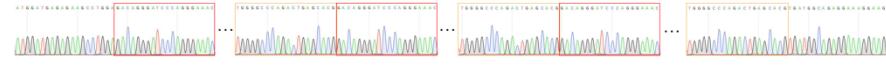
Supplemental Fig. S8 Representative Sanger sequencing results of large fragment TDs at the *FANCF* site (related to Fig. 3A). Representative Sanger sequencing results of TD containing amplicons derived from 3 targeted TDs at *FANCF* site. The amplicons containing targeted TDs were gel-purified and subjected to Sanger sequencing. TD reference sequence was shown on top of each sequencing chromatogram, with sequences corresponding to each end of the TD boxed in red and yellow.

HEK3 TD 136-bp 2xTD

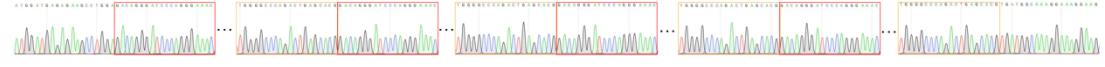
TD reference sequence: ATGGATGAGAGAAGCTGGAGACAGGGATCCCAAGGGAAACG... TGGGCCCCAGACTGAGCACGACAGGGATCCCAAGGGAAACG... TGGGCCCCAGACTGAGCACGACAGGGATCCCAAGGGAAACG...



HEK3 TD 136-bp 3xTD

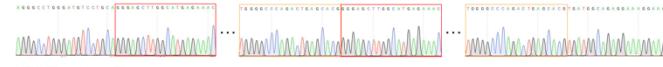


HEK3 TD 136-bp 4xTD



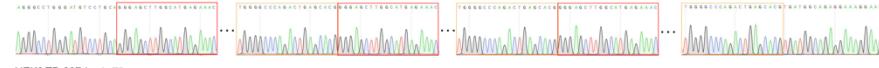
HEK3 TD 222-bp 2xTD

TD reference sequence: AGGGCCCTGGGATGCTCTGCA GGGAGCTCTGGCATGAGAAAC... TGGGGCCCAAGACTGAGGACCGGGAGCTCTGGCATGAGAAAC... TGGGCCCAAGACTGAGGACCGTGTGGCAAGAGGAAAGGAA

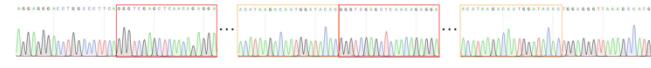


HEK3 TD 222-bp 3xTD

TD reference sequence: AGGGCTGGATGTCCTGCA~~GGGAGCTTGGCATGAGAAC~~...~~TGGGAGCCGACAGTGACCA~~GGGAGCTTGGCATGAGAAC...~~TGGGAGCCGACAGTGACCA~~GGGAGCTTGGCATGAGAAC...~~TGGGAGCCGACAGTGACCA~~TGATGGCAGGAAAGGAAG

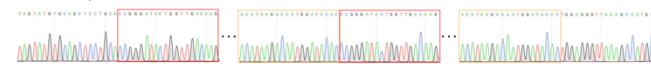


HEK3 TD 397-bp 2×TD

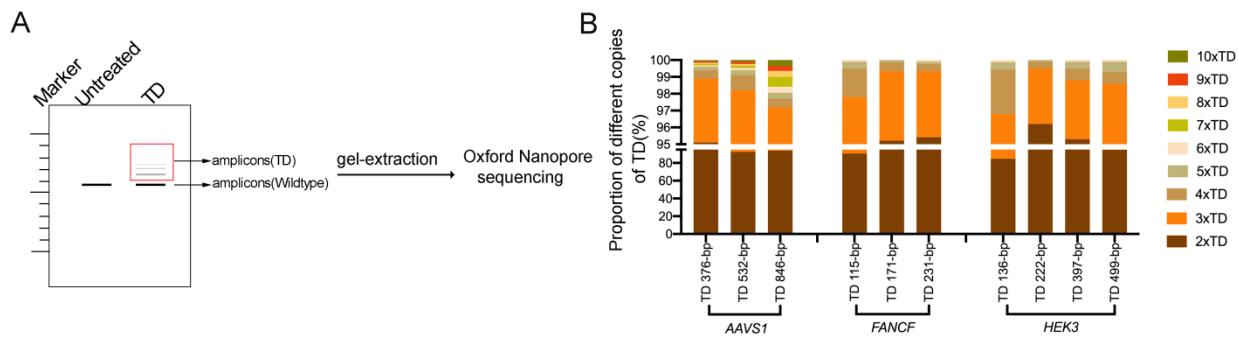


HEK3 TD 499-bp 2×TD

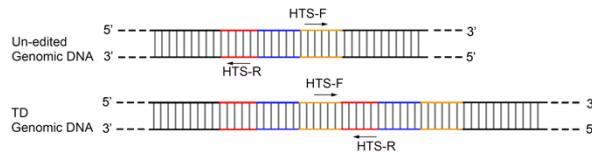
TD reference sequence: TAGTATGTGAGCTCTGAGCGGGATACTGGTTGACAAG...ACATAAGACAATGGTACACCGGGATACTGGTTGACAAG...ACATAAGACAATGGTACACCTGGAGGGTTAACCGCATGCC



Supplemental Fig. S9 Representative Sanger sequencing results of large fragment TDs at the *HEK3* site (related to Fig. 3A). Representative Sanger sequencing results of TD containing amplicons derived from 4 targeted TDs at *HEK3* site. The amplicons containing targeted TDs were gel-purified and subjected to Sanger sequencing. TD reference sequence was shown on top of each sequencing chromatogram, with sequences corresponding to each end of the TD boxed in red and yellow respectively. *HEK3* TD 136-bp 3×TD and 4×TD and *HEK3* TD 222-bp 3×TD were the sequencing results of the bands marked by red stars (*) at the *HEK3* site in Fig. 3A.



Supplemental Fig. S10 Proportions of different copies of TDs were analyzed by Oxford Nanopore sequencing. (A) Diagram showing the procedure to prepare Oxford Nanopore sequencing Samples. The amplicons with size equivalent to or larger than $2 \times$ TD were collected and gel extracted for Nanopore sequencing. (B) The proportions of each type of TDs were analyzed.

A**B****AAVS1 TD 376-bp**

TD reference sequence:**TCCCTCCACCCACAGTGGGCCACTAGGGACAG**GTACCCGCTCCCTGGCTTAGGCCACCTCTCCA.....

2 × TD (Accurate) :**TCCCTCCACCCACAGTGGGCCACTAGGGACAG**GTACCCGCTCCCTGGCTTAGGCCACCTCTCCA..... 321892/604583 (99.99%)

2 × TD with insertion :**TCCCTCCACCCACAGTGGGCCACTAGGGACAG**GTACCCGCTCCCTGGCTTAGGCCACCTCTCCA**TGCT**..... 72/604583 (0.02%)

[] Insertion
reads / total reads**AAVS1 TD 532-bp**

TD reference sequence:**TCCCTCCACCCACAGTGGGCCACTAGGGACAG**GGCGTCTTCACTGGCTGGGTTCCCTTCTCCCTTCTCCCTTCTG.....

2 × TD (Accurate) :**TCCCTCCACCCACAGTGGGCCACTAGGGACAG**GGACAGGGCCGCTTCACTGGCTGGGTTCCCTTCTCCCTTCTG..... 44692/9447308 (99.92%)

2 × TD with insertion :**TATCTGTCCCTCACCCACAGTGGGCCACTAGGGACAG**GGCGTCTTCACTGGCTGGGTTCCCTTCTCCCTTCTG**ATCTGTCCCTCACCCACAGTGGGCCACTAGGGACAGGGCCGCTTCACTGGCTGGGTTCCCTTCTCCCTTCTG**..... 273/447308 (0.06%)

.....**TCCCTCCACCCACAGTGGGCCACTAGGGACAG**GGCGTCTTCACTGGCTGGGTTCCCTTCTCCCTTCTG**TCCCTCCACCCACAGTGGGCCACTAGGGACAGGGCCGCTTCACTGGCTGGGTTCCCTTCTCCCTTCTG**..... 106/447308 (0.02%)

[] Insertion
reads / total reads**AAVS1 TD 846-bp**

TD reference sequence:**TTATCTGTCCCTCACCCACAGTGGGCCACTAGGGACAG**GGCGACTGTTCCCTTCCAGGGAGGTCTGTTCTG.....

2 × TD (Accurate) :**TTATCTGTCCCTCACCCACAGTGGGCCACTAGGGACAG**GGCGACTGTTCCCTTCCAGGGAGGTCTGTTCTG..... 604109/604583 (99.92%)

2 × TD with insertion :**TTATCTGTCCCTCACCCACAGTGGGCCACTAGGGACAG**GGCGACTGTTCCCTTCCAGGGAGGTCTGTTCTG**ATCTGTCCCTCACCCACAGTGGGCCACTAGGGACAGGGCCACTGTTCCCTTCCAGGGAGGTCTGTTCTG**..... 474/604583 (0.08%)

[] Insertion
reads / total reads

Supplemental Fig.S11 Undesired edits of the *AAVS1* site were determined by HTS. (A) PCR strategy for amplifying the junction of TDs. The primers were designed to specifically amplify the boundary of targeted TDs. The HTS primers were listed in Supplemental Table S5. (B) Undesired edits in indicated TDs of the *AAVS1* site. Sequence corresponding to accurate TD allele was shown on top of each analysis. The sequences corresponding to each end of the TD were showed in red and yellow respectively. The red arrowhead pointed to the boundary of the duplication. Insertion and base substitution were indicated by red box and green letters respectively. Note that the inserted sequence was same as its 5' upstream sequence. Sequences with a ratio over 0.02% were shown.

EMX1 TD 116-bp



Supplemental Fig. S12 Undesired edits of *EMX1* TD 116-bp were determined by HTS. Sequence corresponding to accurate TD allele was shown on top of the analysis. The sequences corresponding to each end of the TD were showed in red and yellow respectively. The red arrowhead pointed to the boundary of the duplication. Deletion and insertion were indicated by black dash and red box. Note that the inserted sequence was the same as its 5' upstream sequence. The inserted sequences that were likely derived from sgRNA scaffold were highlighted in blue. Sequences with a ratio over 0.02% were shown.

EMX1 TD 201-bp

□ Insertion

TD reference sequence:	reads / total reads
.....CTGGAGGAGGAAGGGCCTGAGTCGAGCAGAAGA.....	519489/ 524263 (99.09%)
2 x TD (Accurate) :
2 x TD with insertion :
.....CTGGAGGAGGAAGGGCCTGAGTCGAGCAGAAGA.....	1620/524263 (0.31%)
.....CTGGAGGAGGAAGGGCCTGAGTCGAGCAGAAGA.....	1408/524263 (0.27%)
.....CTGGAGGAGGAAGGGCCTGAGTCGAGCAGAAGA.....	327/524263 (0.062%)
.....CTGGAGGAGGAAGGGCCTGAGTCGAGCAGAAGA.....	308/524263 (0.059%)
.....CTGGAGGAGGAAGGGCCTGAGTCGAGCAGAAGA.....	203/524263 (0.039%)
.....CTGGAGGAGGAAGGGCCTGAGTCGAGCAGAAGA.....	146/524263 (0.028%)
.....CTGGAGGAGGAAGGGCCTGAGTCGAGCAGAAGA.....	143/524263 (0.027%)
.....CTGGAGGAGGAAGGGCCTGAGTCGAGCAGAAGA.....	136/524263 (0.026%)
.....CTGGAGGAGGAAGGGCCTGAGTCGAGCAGAAGA.....	132/524263 (0.025%)
.....CTGGAGGAGGAAGGGCCTGAGTCGAGCAGAAGA.....	127/524263 (0.024%)
.....CTGGAGGAGGAAGGGCCTGAGTCGAGCAGAAGA.....	115/524263 (0.022%)
.....CTGGAGGAGGAAGGGCCTGAGTCGAGCAGAAGA.....	109/524263 (0.021%)

Supplemental Fig. S13 Undesired edits of *EMX1* TD 201-bp were determined by HTS.

Sequences corresponding to accurate TD allele was shown on top of the analysis. The sequences corresponding to each end of the TD were showed in red and yellow respectively. The red arrowhead pointed to the boundary of the duplication. Insertions was indicated by red boxes. Note that most inserted sequences were same as their 5' upstream sequences. A small group of insertions that were likely derived from sgRNA scaffold were highlighted in blue. Sequences with a ratio over 0.02% were shown.

EMX1 TD 307-bp

TD reference sequence:CTGGAGGAGGAAGGGCCTGAGTCGAGCAGAAGA.....	- Deletion	□ Insertion	reads / total reads
2 x TD (Accurate) :CTGGAGGAGGAAGGGCCTGAGTCGAGCAGAAGA.....	207814/ 210165 (98.88%)
2 x TD with insertion :CTGGAGGAGGAAGGGCCTGAGTCGAGCAGAAGA.....	CTGGAGGAGGAAGGGCCTGAGTCGAGCAGAAGA.....	743/210165 (0.35%)
CTGGAGGAGGAAGGGCCTGAGTCGAGCAGAAGA.....	33-bp	142/210165 (0.068%)
CTGGAGGAGGAAGGGCCTGAGTCGAGCAGAAGA.....	21-bp	82/210165 (0.039%)
CTGGAGGAGGAAGGGCCTGAGTCGAGCAGAAGA.....	16-bp	69/210165 (0.033%)
CTGGAGGAGGAAGGGCCTGAGTCGAGCAGAAGA.....	26-bp	60/210165 (0.029%)
CTGGAGGAGGAAGGGCCTGAGTCGAGCAGAAGA.....	A.....	58/210165 (0.028%)
CTGGAGGAGGAAGGGCCTGAGTCGAGCAGAAGA.....	18-bp	51/210165 (0.024%)
2 x TD with deletion :CTGGAGGAGGAAGGGCCTGAGTCGAGCAGAAGA.....	337/210165 (0.16%)
CTGGAGGAGGAAGGGCCTGAGTCGAGCAGAAGA.....	153/210165 (0.073%)
CTGGAGGAGGAAGGGCCTGAGTCGAGCAGAAGA.....	125/210165 (0.059%)
CTGGAGGAGGAAGGGCCTGAGTCGAGCAGAAGA.....	125/210165 (0.059%)
CTGGAGGAGGAAGGGCCTGAGTCGAGCAGAAGA.....	124/210165 (0.059%)
CTGGAGGAGGAAGGGCCTGAGTCGAGCAGAAGA.....	94/210165 (0.045%)
CTGGAGGAGGAAGGGCCTGAGTCGAGCAGAAGA.....	79/210165 (0.038%)
CTGGAGGAGGAAGGGCCTGAGTCGAGCAGAAGA.....	58/210165 (0.028%)
CTGGAGGAGGAAGGGCCTGAGTCGAGCAGAAGA.....	51/210165 (0.024%)

Supplemental Fig. S14 Undesired edits of *EMX1* TD 307-bp were determined by HTS.

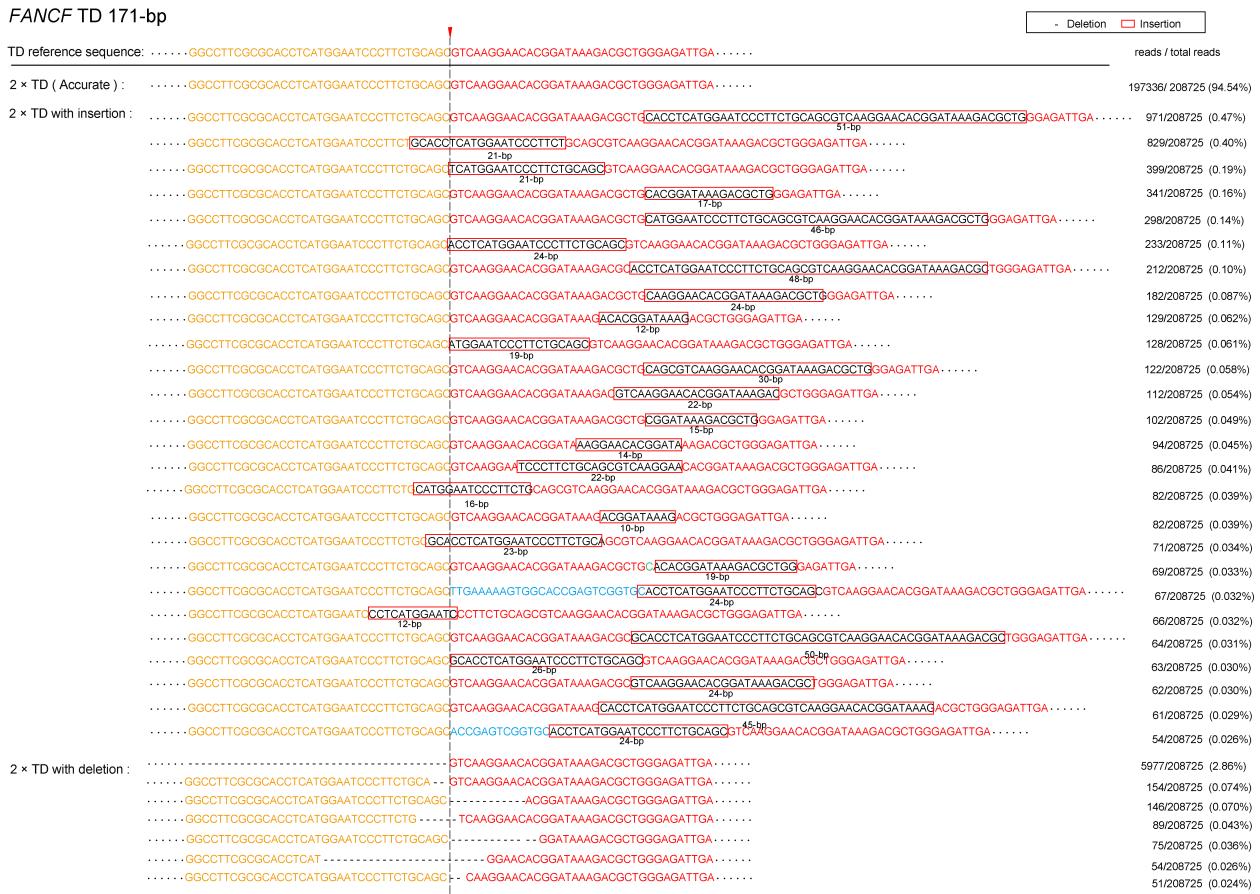
Sequence corresponding to accurate TD allele was shown on top of each analysis. The sequences corresponding to each end of the TD were showed in red and yellow respectively. The red arrowhead pointed to the boundary of the duplication. Deletion, insertion and base substitution were indicated by black dash, red box and green letters respectively. The inserted sequence in the red box was same as with its 5' upstream sequence. The sequences with a ratio over 0.02% were shown.

FANCF TD 115-bp

Supplemental Fig. S15 Undesired edits of *FANCF* TD 115-bp were determined by HTS.

Sequence corresponding to accurate TD allele was shown on top of the analysis. The sequences corresponding to each end of the TD were showed in red and yellow respectively. The red arrowhead pointed to the boundary of the duplication. Deletion, insertion and base substitution were indicated by black dash, red box and green letters respectively. Note that the inserted sequence in red box was same as its 5' upstream sequence. A small group of insertions that were likely derived from sgRNA scaffold were highlighted in blue. The sequences with a ratio over 0.02% were shown.

FANCF TD 171-bp



Supplemental Fig. S16 Undesired edits of *FANCF* TD 171-bp were determined by HTS.

Sequence corresponding to accurate TD allele was shown on top of the analysis. The sequences corresponding to each end of the TD were showed in red and yellow respectively. The red arrowhead pointed to the boundary of the duplication. Deletion, insertion and base substitution were indicated by black dash, red box and green letters respectively. Note that most inserted sequences were same as their 5' upstream sequences. A small group of insertions that were likely derived from sgRNA scaffold were highlighted in blue. The sequences with a ratio over 0.02% were shown.

FANCF TD 231-bp

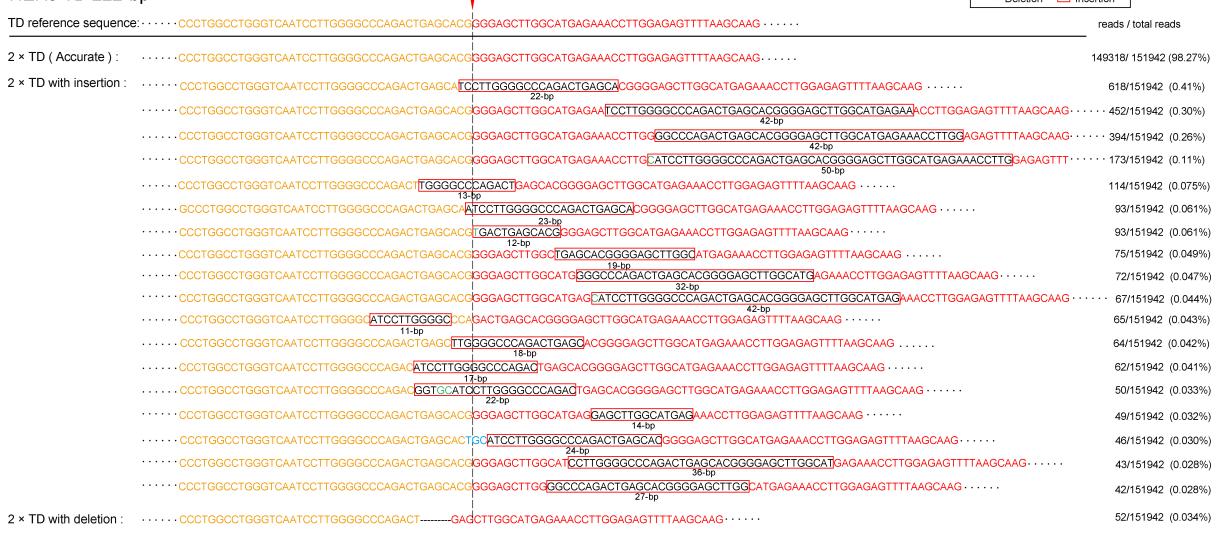
Supplemental Fig. S17 Undesired edits of *FANCF* TD 231-bp were determined by HTS.

Sequence corresponding to accurate TD allele was shown on top of the analysis. The sequences corresponding to each end of the TD were showed in red and yellow respectively. The red arrowhead pointed to the boundary of the duplication. Deletion, insertion and base substitution were indicated by black dash, red box and green letters respectively. Note that most inserted sequences were same as their 5' upstream sequences. A small group of insertions that were likely derived from sgRNA scaffold were highlighted in blue. The sequences with a ratio over 0.02% were shown.

HEK3 TD 136-bp

Supplemental Fig. S18 Undesired edits of *HEK3* TD 136-bp were determined by HTS. Sequence corresponding to accurate TD allele was shown on top of the analysis. The sequences corresponding to each end of the TD were showed in red and yellow respectively. The red arrowhead pointed to the boundary of the duplication. Insertion and base substitution were indicated by red box and green letters respectively. Note that most inserted sequences were same as their 5' upstream sequences. A small group of insertions that were likely derived from sgRNA scaffold were highlighted in blue. The sequences with a ratio over 0.02% were shown.

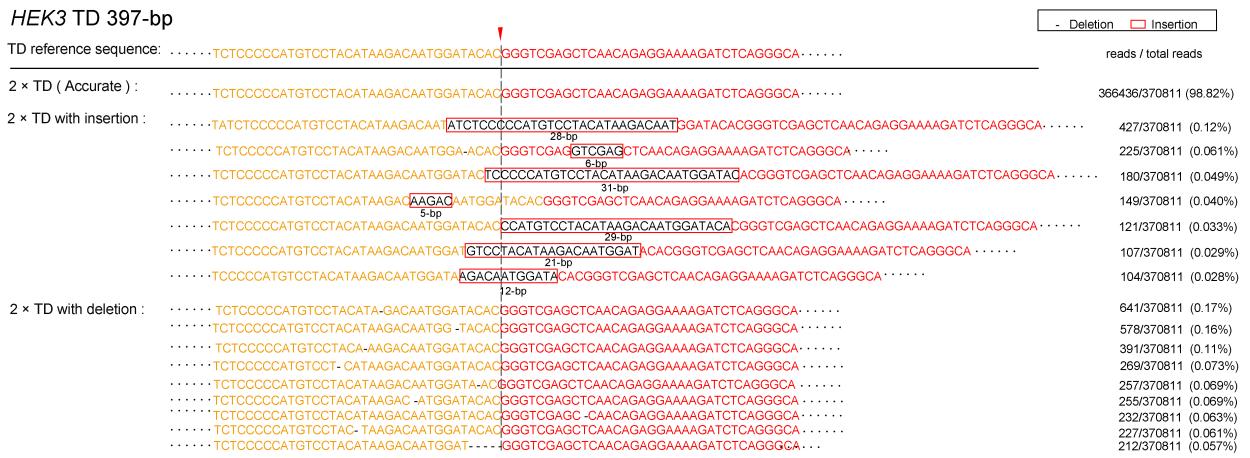
HEK3 TD 222-bp



Supplemental Fig. S19 Undesired edits of *HEK3* TD 222-bp were determined by HTS.

Sequence corresponding to accurate TD allele was shown on top of the analysis. The sequences corresponding to each end of the TD were showed in red and yellow respectively. The red arrowhead pointed to the boundary of the duplication. Deletion, insertion and base substitution were indicated by black dash, red box and green letters respectively. Note that most inserted sequences were the same as their 5' upstream sequences. A small group of insertions that were likely derived from sgRNA scaffold were highlighted in blue. The sequences with a ratio over 0.02% were shown.

HEK3 TD 397-bp



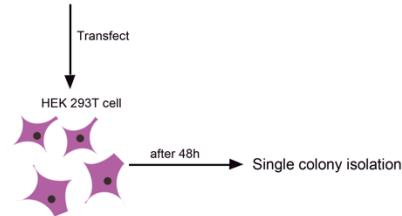
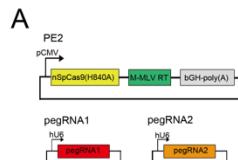
Supplemental Fig. S20 Undesired edits of *HEK3* TD 397-*bp* were determined by HTS. Sequence corresponding to accurate TD allele was shown on top of the analysis. The sequences corresponding to each end of the TD were showed in red and yellow respectively. The red arrowhead pointed to the boundary of the duplication. Deletion and insertion were indicated by black dash and red box respectively. Note that most inserted sequences were same as their 5' upstream sequences. The sequences with a ratio over 0.02% were shown.

HEK3 TD 499-bp

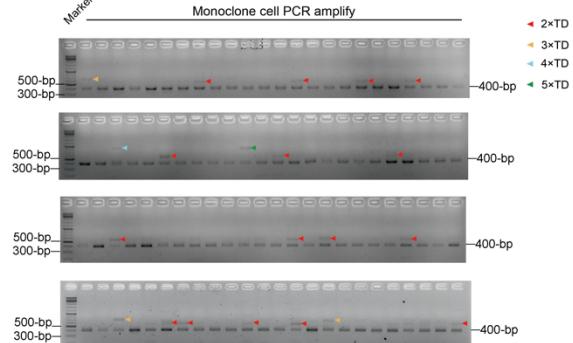


Supplemental Fig. S21 Undesired edits of *HEK3* TD 499-bp were determined by HTS.

Sequence corresponding to accurate TD allele was shown on top of the analysis. The sequences corresponding to each end of the TD were showed in red and yellow respectively. The red arrowhead pointed to the boundary of the duplication. Insertion was indicated by red box. Note that most inserted sequences were same as their 5' upstream sequences. The sequences with a ratio over 0.02% were shown.



B HEK3 TD 136-bp



C HEK3 TD 136-bp 2×TD

TD reference sequence: ATGGATGAGAGAAGCCTGGA GACAGGGATCCAGGGAAACG... TGGGGCCAGACTGAGCACCG GACAGGGATCCAGGGAAACG... TGGGGCCAGACTGAGCACCTGATGGAGAGGAAGGAG

HEK3 TD 136-bp 3×TD

TD reference sequence: ATGGATGAGAGAAGCCTGGA GACAGGGATCCAGGGAAACG... TGGGGCCAGACTGAGCACCG GACAGGGATCCAGGGAAACG... TGGGGCCAGACTGAGCACCTGATGGAGAGGAAGGAG

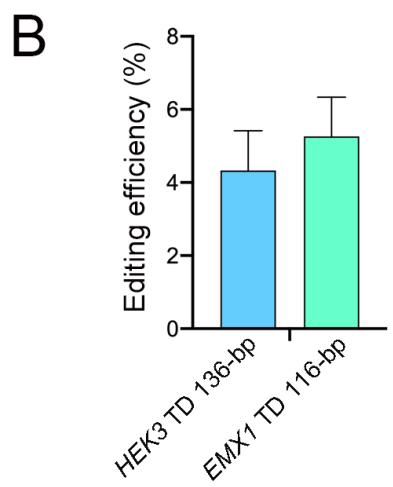
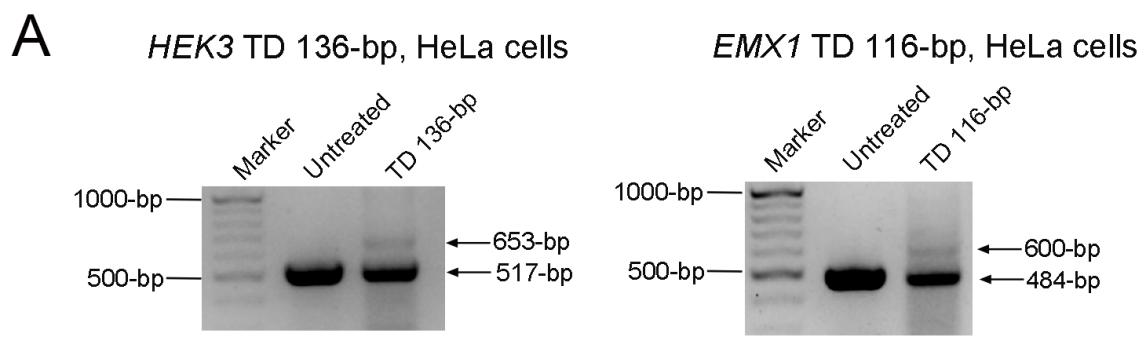
HEK3 TD 136-bp 4×TD

TD reference sequence: ATGGATGAGAGAAGCCTGGA GACAGGGATCCAGGGAAACG... TGGGGCCAGACTGAGCACCG GACAGGGATCCAGGGAAACG... TGGGGCCAGACTGAGCACCTGATGGAGAGGAAGGAG

HEK3 TD 136-bp 5×TD

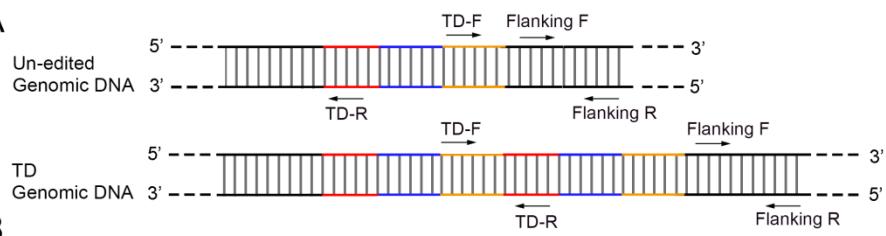
TD reference sequence: ATGGATGAGAGAAGCCTGGA GACAGGGATCCAGGGAAACG... TGGGGCCAGACTGAGCACCG GACAGGGATCCAGGGAAACG... TGGGGCCAGACTGAGCACCTGATGGAGAGGAAGGAG

Supplemental Fig. S22 Monoclonal analysis of the HEK3 TD 136-bp. (A) Diagram showing the procedure of monoclonal analysis. HEK293T cells were transfected with TD-PE plasmids, including PE2 and two pegRNA, and 48 hours post the transfection cells were diluted monoclonal isolation. (B) Agarose gel electrophoresis showed the amplicons derived from each monoclonal genome. The colonies containing different copies of TDs were highlighted with triangles of different colors. (C) Sanger sequencing results of representative TDs derived from monoclonal genomes. Reference sequences of each type of TDs were shown on top of the sequence chromatogram, and the duplicated sequences corresponding to each end of the TD were marked by yellow and red boxes respectively.

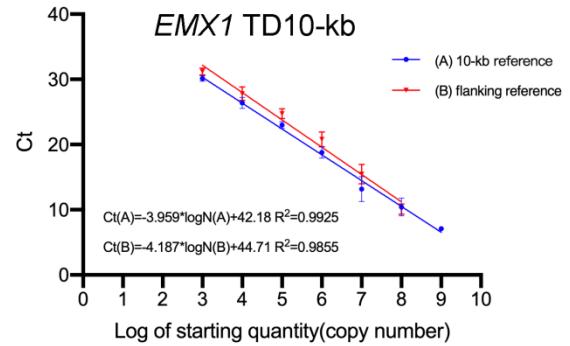
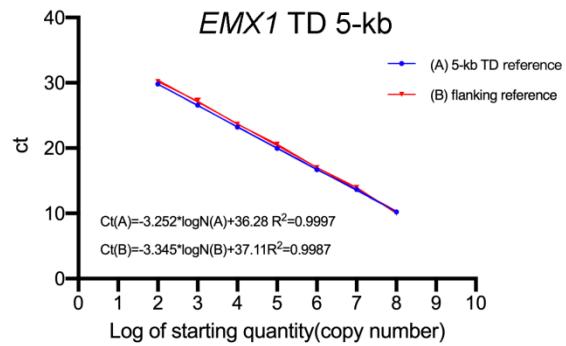


Supplemental Fig. S23 Verification of TD-PE strategy in HeLa cells. (A) Agarose gel analysis of the presence of targeted tandem duplication at indicated loci. (B) Quantification of the efficiency of targeted tandem duplication in (A) using Image J. Values and error bars reflect mean \pm s.d. of n=3 independent biological replicates.

A



B



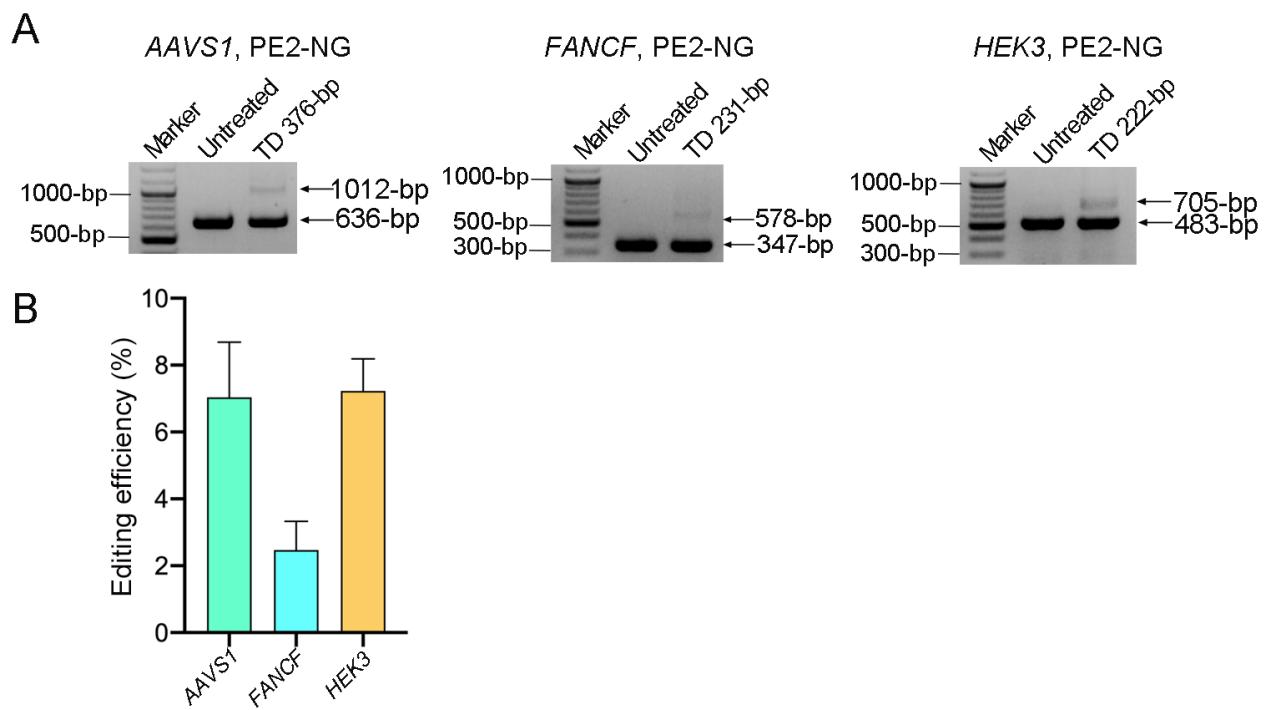
Supplemental Fig. S24 Standard curves for absolute quantification by real-time PCR of *EMX1* TD 5-kb and 10-kb. (A) Schematic diagram showing the design of qPCR primers. The flanking fragment of the target site or the one containing tandem duplication obtained by PCR were ligated to the blunt vector to construct reference plasmids. qPCR primers were designed inside the amplified fragment. The primers used to construct the reference plasmids and qPCR primers are listed in Supplemental Table S4. (B) The standard curves for the reference plasmids were determined by CT values against log-transformed concentrations of serial tenfold dilutions (2×10^2 , 10^3 , 10^4 , 10^5 , 10^6 , 10^7 , 10^8 and 10^9 copies per $1\mu\text{l}$).

EMX1 TD 5-kb

TD reference sequence:+ACCTGGAGGAGGAAGGGCTCTAGTCGCAAGAGA..... reads / total reads
2 x TD (Accurate) :+ACCTGGAGGAGGAAGGGCTCTAGTCGCAAGAGA.....	100690/100760 (99.93%)
2 x TD with insertion :+ACCTGGAGGAGGAAGGGCTCTAGTCGCAAGAGA.....	70/100760 (0.069%)
<input checked="" type="checkbox"/> Deletion <input type="checkbox"/> Insertion		
EMX1 TD-10-kb	 reads / total reads
TD reference sequence:+ACGGCAGAGCTGGAGGAGGAAGGGCTCTAGTCGCAAGAGA.....
	+ACCATGGGGAGCTGAGTCCCTATGTCGCAAGAGA.....	
2 x TD (Accurate) :+ACGGCAGAGCTGGAGGAGGAAGGGCTCTAGTCGCAAGAGA.....	417717418256 (99.87%)
2 x TD with insertion :+ACGGCAGAGCTGGAGGAGGAAGGGCTCTAGTCGCAAGAGA.....	146418256 (0.03%)
	+ACCATGGGGAGCTGAGTCCCTATGTCGCAAGAGA.....	
	+ACCATGGGGAGCTGAGTCCCTATGTCGCAAGAGA.....	83-kb
	+ACGGCAGAGCTGGAGGAGGAAGGGCTCTAGTCGCAAGAGA.....	144418256 (0.034%)
	+ACCATGGGGAGCTGAGTCCCTATGTCGCAAGAGA.....	73-kb
2 x TD with deletion :+ACGGCAGAGCTGGAGGAGGAAGGGCTCTAGTCGCAAGAGA.....	106418256 (0.025%)
	+ACCATGGGGAGCTGAGTCCCTATGTCGCAAGAGA.....	
	+ACGGCAGAGCTGGAGGAGGAAGGGCTCTAGTCGCAAGAGA.....	143/18256 (0.034%)

Supplemental Fig. S25 Undesired edits of *EMX1* TD 5-kb/10-kb were determined by HTS.

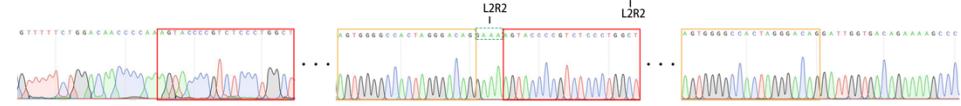
Sequence corresponding to accurate TD allele was shown on top of the analysis. The sequences corresponding to each end of the TD were showed in red and yellow respectively. The red arrowhead pointed to the boundary of the duplication. Deletion and insertion were indicated by black dash and red box respectively. Note that most inserted sequences were the same as their 5' upstream sequences. The sequences with a ratio over 0.02% were shown.



Supplemental Fig. S26 Targeted TDs produced by PE2-NG. (A) Targeted TDs produced by PE2-NG. Agarose gel analysis of the presence of targeted tandem duplication at indicated loci. B, Quantification of the efficiency of targeted tandem duplication in (B) using Image J. Values and error bars reflect mean \pm s.d. of n=3 independent biological replicates.

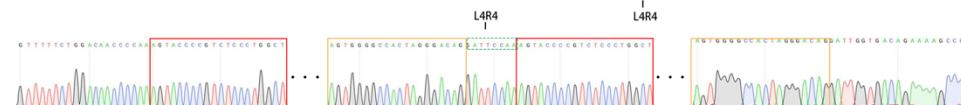
AAVS1 TD 376-bp L2R2

TD reference sequence: GTTTTCTGGACAACCCAAAGTACCCCGTCTCCCTGGCT...AGTGGGCCACTAGGGACAGGAAAAGTACCCCGTCTCCCTGGCT...AGTGGGCCACTAGGGACAGGATTGGTACAGAAAAGCCC



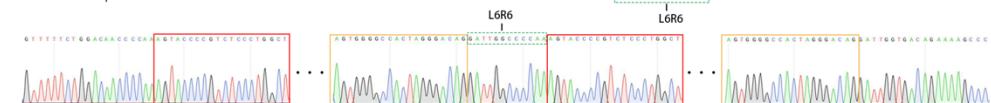
AAVS1 TD 376-bp L4R4

TD reference sequence: GTTTTCTGGACAACCCAAAGTACCCCGTCTCCCTGGCT...AGTGGGCCACTAGGGACAGGATTCCTAAAGTACCCCGTCTCCCTGGCT...AGTGGGCCACTAGGGACAGGATTGGTACAGAAAAGCCC



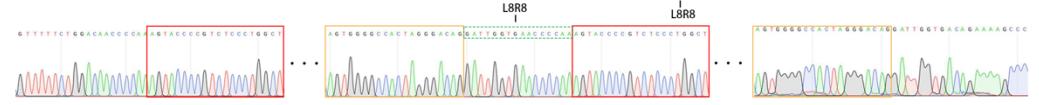
AAVS1 TD 376-bp L6R6

TD reference sequence: GTTTTCTGGACAACCCAAAGTACCCCGTCTCCCTGGCT...AGTGGGCCACTAGGGACAGGATTGGCCCAAAGTACCCCGTCTCCCTGGCT...AGTGGGCCACTAGGGACAGGATTGGTACAGAAAAGCCC



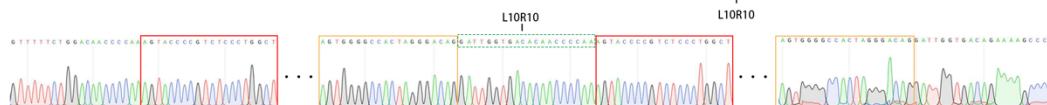
AAVS1 TD 376-bp L8R8

TD reference sequence: GTTTTCTGGACAACCCAAAGTACCCCGTCTCCCTGGCT...AGTGGGCCACTAGGGACAGGATTGGTGAAACCCAAAGTACCCCGTCTCCCTGGCT...AGTGGGCCACTAGGGACAGGATTGGTACAGAAAAGCCC



AAVS1 TD 376-bp L10R10

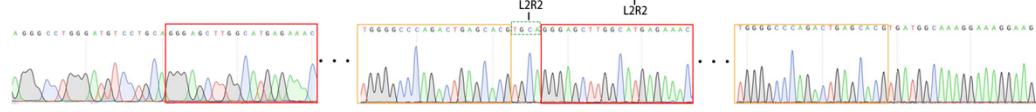
TD reference sequence: GTTTTCTGGACAACCCAAAGTACCCCGTCTCCCTGGCT...AGTGGGCCACTAGGGACAGGATTGGTGACACAACCCAAAGTACCCCGTCTCCCTGGCT...AGTGGGCCACTAGGGACAGGATTGGTACAGAAAAGCCC



Supplemental Fig.S27 Representative Sanger sequencing results of the additive fragment containing TDs at the *AAVS1* site (related to Fig. 4B). The PCR amplicons containing targeted TDs were gel-purified and subjected directly to Sanger sequencing. TD reference sequences were shown on top of each Sequencing chromatogram. The sequences corresponding to each end of the TDs were marked by yellow and red boxes respectively. Sequences in green dashed boxes represent additive fragments.

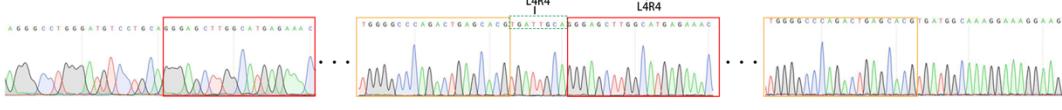
HEK3 TD 222-bp L2R2

TD reference sequence: AGGGCCTGGGATGTCTGCA **GGGAGCTTGGCATGAGAAC**...TGGGGCCAGACTGAGCACCG**TGCAGGGAGCTTGGCATGAGAAC**...TGGGGCCAGACTGAGCACCG**TGATGGCAGAGGAAGGAAG**



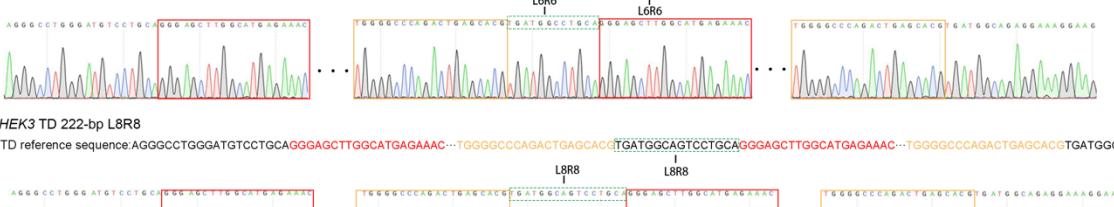
HEK3 TD 222-bp L4R4

TD reference sequence: AGGGCCTGGGATGTCTGCA **GGGAGCTTGGCATGAGAAC**...TGGGGCCAGACTGAGCACCG**TGATGGCAGAGGAAGGAAG**



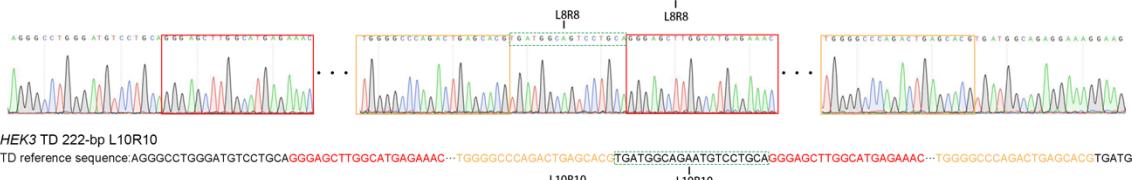
HEK3 TD 222-bp L6R6

TD reference sequence: AGGGCCTGGGATGTCTGCA **GGGAGCTTGGCATGAGAAC**...TGGGGCCAGACTGAGCACCG**TGATGGCCTGCA** **GGGAGCTTGGCATGAGAAC**...TGGGGCCAGACTGAGCACCG**TGATGGCAGAGGAAGGAAG**



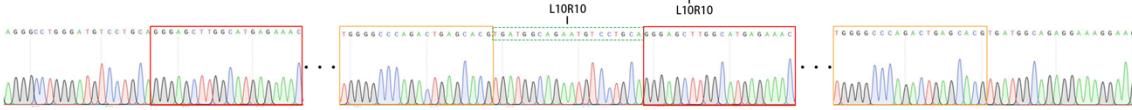
HEK3 TD 222-bp L8R8

TD reference sequence: AGGGCCTGGGATGTCTGCA **GGGAGCTTGGCATGAGAAC**...TGGGGCCAGACTGAGCACCG**TGATGGCAGTCTGCA** **GGGAGCTTGGCATGAGAAC**...TGGGGCCAGACTGAGCACCG**TGATGGCAGAGGAAGGAAG**



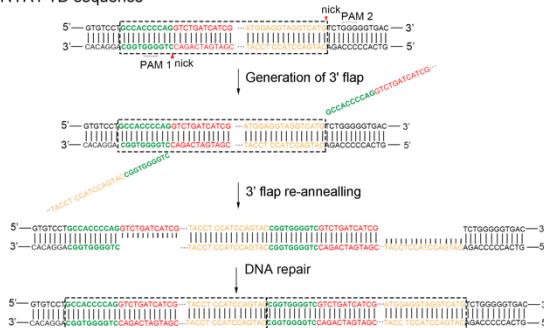
HEK3 TD 222-bp L10R10

TD reference sequence: AGGGCCTGGGATGTCTGCA **GGGAGCTTGGCATGAGAAC**...TGGGGCCAGACTGAGCACCG**TGATGGCAGAATGTCTGCA** **GGGAGCTTGGCATGAGAAC**...TGGGGCCAGACTGAGCACCG**TGATGGCAGAGGAAGGAAG**

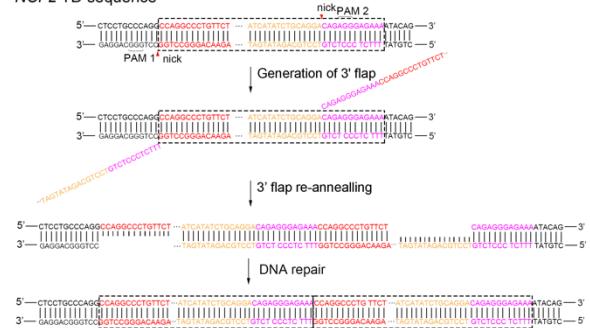


Supplemental Fig.S28 Representative Sanger sequencing results of the additive fragment containing TDs at the *HEK3* site (related to Fig. 4B). The PCR amplicons containing targeted TDs were gel-purified and subjected directly to Sanger sequencing. TD reference sequences were shown on top of each Sequencing chromatogram. The sequences corresponding to each end of the TDs were marked by yellow and red boxes respectively. Sequences in green dashed boxes represent additive fragments.

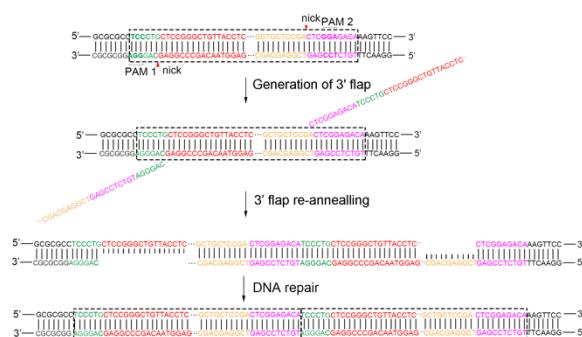
RYR1 TD sequence



NCF2 TD sequence



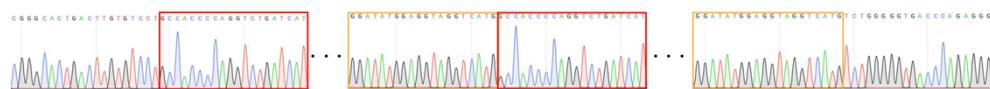
SATB2 TD sequence



Supplemental Fig.S29 PegRNAs design at three disease loci and the putative editing process of TD-PE strategy. PAM1 and PAM2 are the upstream and downstream PAMs respectively, and the red arrows indicate the positions of the nicks. Black dashed box indicates the sequences to be duplicated. Green and pink sequences denote the upstream and downstream addictive fragments respectively.

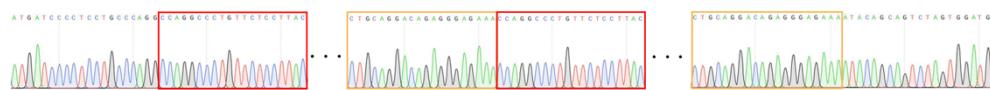
RYR1 TD 85-bp

TD reference sequence: CGGGCACTGACTTGTGTCCTGCCACCCAGGTCTGATCAT...GGATATGGAGGTAGGTCTGCCACCCAGGTCTGATCAT...GGATATGGAGGTAGGTCTGCCACCCAGGTCTGATCAT...GGATATGGAGGTAGGTCTGCCACCCAGGTCTGATCAT...GGATATGGAGGTAGGTCTGCCACCCAGGTCTGACCCAGAGGG



NCF2 TD 149-bp

TD reference sequence: ATGATCCCTCCCTGCCAGGCCAGGCCCTGTTCTCCCTTAC...CTGCAGGACAGAGGGAGAAACAGGCCCTGTTCTCCCTTAC...CTGCAGGACAGAGGGAGAAACAGGCCCTGTTCTCCCTTAC...ATACAGCAGTCTAGTGGATG



ENG TD 180-bp

TD reference sequence: CCCCACCCGACCCGCCATGGGACACTCACCGTTGGAAAC...ACCTGTTGGAGAACATCCGGGACACTCACCGTTGGAAAC...ACCTGTTGGAGAACATCCGGAAAGAGGCCAGGTGAGAAT



SA7B2 TD 209-bp

TD reference sequence: TGGGCCGCGGGCTGCGCGCTCCCTGCTCCGGCTGTTAC...GCTGCTCCGACTCGGAGACATCCCTGCTCCGGCTGTTAC...GCTGCTCCGACTCGGAGACAAAGTTCCCACCGCAGGTG



Supplemental Fig.S30 Representative Sanger sequencing results of 4 disease-related TDs at indicated loci (related to Fig. 5B). The PCR amplicons containing targeted tandem duplication were gel-purified and subjected directly to Sanger sequencing. TD reference sequences were shown on top of each Sequencing result. The sequences corresponding to each end of the TDs were marked by red and yellow boxes respectively.

Supplemental Table 1. List of the targets tested in this study.

sgRNA	Target sequence	Oligo-F	Oligo-R	Reference
<i>FANCF</i>	GGAATCCCTTCTG	CACCGGAATCCCTT	AAACGGTGCTGCA	(Anzalone et al. 2019)
	CAGCACC TGG	CTGCAGCACC	GAAGGGATTCC	
<i>FANCF</i> -50	GCCCTACTTCCGC	CACCGCCCTACTTC	AAACAGGTGAAAG	(Anzalone et al. 2019)
	TTTCACCT TGG	CGCTTCACCT	CGGAAGTAGGG	
<i>EMX1</i>	AGTCCGAGCAGA	CACCGAGTCCGAG	AAACTTCTTCTTCT	(Anzalone et al. 2019)
	AGAAGAA GGG	CAGAAGAAGAA	GCTCGGACTC	
<i>EMX1</i> -57	CCGTTGTACTTT	CACCGCCGTTGTA	AAACGAGGACAA	(Anzalone et al. 2019)
	GTCCTC CGG	CTTTGTCCTC	AGTACAAACGGC	
<i>FANCF</i> +115	GCAGAGAGTCGC	CACCGCAGAGAGT	AAACTGGAGACGG	This study
	CGTCTCCA AGG	CGCCGTCTCCA	CGACTCTCTGC	
<i>FANCF</i> -115	TATCCGTGTTCCCT	CACCGTATCCGTGT	AAACAGAGTCAAG	This study
	TGACTCT GGG	TCCTTGACTCT	GAACACGGATAC	
<i>FANCF</i> -171	TATCCGTGTTCCCT	CACCGTATCCGTGT	AAACAGAGTCAAG	This study
	TGACTCT GGG	TCCTTGACTCT	GAACACGGATAC	
<i>FANCF</i> -231	AAAGAGGTTCTT	CACCGAAAGAGGT	AAACCCACTAAGA	This study
	CTTAGTGG AGG	TCTTCTTAGTGG	AGAACCTCTTC	
<i>EMX1</i> -116	GGGGCACAGATG	CACCGGGGCACAG	AAACGAGTTCTC	(Anzalone et al. 2019)
	AGAAAATC AGG	ATGAGAAACTC	ATCTGTGCC	
<i>EMX1</i> -201	ACTGAGGCTACA	CACCGACTGAGGCT	AAACTAACCTAT	This study
	TAGGGTTA GGG	ACATAGGGTTA	GTAGCCTCAGTC	
<i>EMX1</i> -307	CAGTGGGCCAGA	CACCGCAGTGGC	AAACCCCTCTCT	This study
	GAGAAGGG TGG	CAGAGAGAAGGG	CTGGCCCAC	
<i>AAVS1</i>	GGGGCCACTAGG	CACCGGGGCCACT	AAACATCCTGTCC	This study
	GACAGGAT TGG	AGGGACAGGAT	CTAGTGGCCC	
<i>AAVS1</i> -376	CAGGGAGACGGG	CACCGCAGGGAGA	AAACCAAAGTACC	This study
	GTACTTG GGG	CGGGGTACTTTG	CCGTCTCCCTGC	
<i>AAVS1</i> -532	CAGCGAGTGAAG	CACCGCAGCGAGT	AAACCATGCCGT	This study
	ACGGCATG GGG	GAAGACGGCATG	TTCACTCGCTGC	
<i>AAVS1</i> -846	AAGGGGAAACAG	CACCGAAGGGAA	AAACCTGGCCCAC	This study
	TGGGCCAG AGG	ACAGTGGGCCAG	TGTTTCCCTTC	
<i>HEK3</i>	GGCCCAGACTGA	CACCGGCCAGACT	AAACTCACGTGCT	(Anzalone et al. 2019)
	GCACGTGA TGG	GAGCACGTGA	CAGTCTGGGCC	
<i>HEK3</i> -136	TCCCTGGGATCCC	CACCGTCCCTGGGA	AAACGGAGACAG	This study
	TGTCTCC AGG	TCCCTGTCTCC	GGATCCCAGGGAC	
<i>HEK3</i> -222	TCTCATGCCAAGC	CACCGTCTCATGCC	AAACGCAGGGAGC	This study
	TCCCTGC AGG	AAGCTCCCTGC	TTGGCATGAGAC	

<i>HEK3+1</i>	TAAGACAATGGA	CACCGTAAGACAA	AAACCCAGTGTAT	This study
	TACACTGG AGG	TGGATACACTGG	CCATTGTCTTAC	
<i>HEK3-397</i>	TCTGTTGAGCTCG	CACCGTCTGTTGAG	AAACTCAGGGTCG	This study
	ACCCCTGA AGG	CTCGACCCTGA	AGCTAACAGAC	
<i>HEK3-499</i>	GTCAACCAGTAT	CACCGTCAACCAGT	AAACGCACCGGGA	This study
	CCCGGTGC AGG	ATCCCGGTGC	TACTGGTTGAC	
<i>ENG+1</i>	GTTTCTGCAAGAC	CACCGTTCTGCAA	AAACCCAACAGGT	This study
	CTGTTGG AGA	GACCTGTTGG	CTTGCAGAAC	
<i>ENG-180</i>	CTTCCTGGAGTTC	CACCGCTTCCTGGA	AAACCCGTTGGGA	This study
	CCAACGG TGA	GTTCCTAACGG	ACTCCAGGAAG C	
<i>SATB2+1</i>	GCTCCATGCTGCT	CACCGCTCCATGCT	AAACGAGTCGGAG	This study
	CCGACTC GGA	GCTCCGACTC	CAGCATGGAGC	
<i>SATB2-209</i>	GAGGTAACAGCC	CACCGAGGTAACA	AAACCTGCTCCGG	This study
	CGGAGCAG GGA	GCCCAGGAGCAG	GCTGTTACCTC	
<i>NCF2+1</i>	AAATCATATCTGC	CACCGAAATCATAT	AAACCTGTCCTGC	This study
	AGGACAG AGG	CTGCAGGACAG	AGATATGATTTC	
<i>NCF2-149</i>	AGGAGAACAGGG	CACCGAGGAGAAC	AAACAGGCCAGGC	This study
	CCTGGCCT GGG	AGGGCCTGGCCT	CCTGTTCTCCTC	
<i>RYRI+1</i>	TATGGAGGTAGG	CACCGTATGGAGGT	AAACAGACATGAC	This study
	TCATGTCT GGG	AGGTCATGTCT	CTACCTCCATAC	
<i>RYRI-85</i>	AGCGTCGATGAT	CACCGAGCGTCGAT	AAACCAGGTCTGA	This study
	CAGACCTG GGG	GATCAGACCTG	TCATCGACGCTC	

Supplemental Table 2. Sequences of pegRNAs used in mammalian cell experiments.

pegRNA	spacer	3' extension (5' to 3')	PBS length (nt)	HA length (nt)
<i>FANCF</i> +TD50 pegRNA	GGAATCCCTCTGCAGCACC	CGCGAAGGCCCTACTTCCGCTTCAGCT GCAGAAGGGA	13	25
<i>FANCF</i> -TD50 pegRNA	GCCCTACTTCCGCTTCACCT	CACCTCATGGAATCCCTCTGCAGCTGA AAGCGGAAGT	13	25
<i>EMX1</i> +TD57 pegRNA	GAGTCCGAGCAGAAGAAGA A	CTCCAGCTTCTGCCGTTGTACTTGTCT TCTTCTGCTCGG	13	28
<i>EMX1</i> -TD57 pegRNA	CCGTTTGACTTGTCCCTC	GAGGAAGGCCCTGAGTCCGAGCAGAAG AAGACAAAGTACAAA	13	29
<i>FANCF</i> +TD115 pegRNA	GCAGAGAGTCGCCGTCTCCA	AGCGTCTTATCCGTGTTCCCTGACAGAC GGCGACTC	12	25
<i>FANCF</i> -TD115 pegRNA	TATCCGTGTTCCCTTGACTCT	ATCAGTACGCAGAGAGTCGCCGTCTGTC AAGGAACAC	12	25
<i>FANCF</i> +TD171 pegRNA	GGAATCCCTCTGCAGCACC	AGCGTCTTATCCGTGTTCCCTGACGCTG CAGAAGGGA	13	25
<i>FANCF</i> -TD171 pegRNA	TATCCGTGTTCCCTTGACTCT	ACCTCATGGAATCCCTCTGCAGCGTCA AGGAACACG	13	24
<i>FANCF</i> +TD231 pegRNA	GGAATCCCTCTGCAGCACC	GCCACACAAAGAGGTTCTCTAGGCTG CAGAAGGGA	13	24
<i>FANCF</i> -TD231 pegRNA	AAAGAGGTTCTCTAGTGG	ACCTCATGGAATCCCTCTGCAGC CTAAGAAGAACCT	13	24
<i>EMX1</i> +TD116 pegRNA	GAGTCCGAGCAGAAGAAGA A	GGGAGGGAGGGCACAGATGAGAAATT CTTCTGCTCGG	13	25
<i>EMX1</i> -TD116 pegRNA	GGGGCACAGATGAGAAACTC	AAGGGCCTGAGTCCGAGCAGAAGAATT CTCATCTGTG	13	25
<i>EMX1</i> +TD201 pegRNA	GAGTCCGAGCAGAAGAAGA A	ATGGGAAGACTGAGGCTACATAGGGTT TTCTGCTCGG	13	25
<i>EMX1</i> -TD201 pegRNA	ACTGAGGCTACATAGGGTTA	AAGGGCCTGAGTCCGAGCAGAAGAAC CTATGTAGCCT	13	25
<i>EMX1</i> +TD307 pegRNA	GAGTCCGAGCAGAAGAAGA A	GATGGCAGGGCAGGAAGAGGGACACAGT GGGCCAGAGAGAATTCTCTGCTCGG	13	40
<i>EMX1</i> -TD307 pegRNA	CAGTGGGCCAGAGAGAAGG G	AGAAGCTGGAGGGAGGAAGGGCCTGAGT CCGAGCAGAAGAATTCTCTGGCCC	13	40
<i>AAVS1</i> +TD376 pegRNA	GGGGCCACTAGGGACAGGAT	GAGGATGGAGAGGTGGCTAAAGCCAGG GAGACGGGGTACTCTGTCCCTAGTGG	13	40
<i>AAVS1</i> -TD376 pegRNA	CAGGGAGACGGGGTACTTTG	ATCTGTCCCCTCCACCCCCACAGTGGGGC CACTAGGGACAGAGTACCCGTCTC	13	40
<i>AAVS1</i> +TD532 pegRNA	GGGGCCACTAGGGACAGGAT	AGAAGGGAGAAGGAAAAGGGAACCCAGC GAGTGAAGACGGCCTGTCCCTAGTGG	13	40
<i>AAVS1</i> -TD532 pegRNA	CAGCGAGTGAAGACGGCATG	ATCTGTCCCCTCCACCCCCACAGTGGGGC CACTAGGGACAG GCCGTCTCACTC	13	40

<i>AAVSI</i> +TD846	GGGGCCACTAGGGACAGGAT	AGAGAAAGCAGGACCTGCCTGGGAAGG	13	40
pegRNA		GGAAACAGTGGGCCTGTCCTAGTGG		
<i>AAVSI</i> -TD846	AAGGGGAAACAGTGGGCCA	ATCTGCCCCCTCCACCCCCACAGTGGGGC	13	40
pegRNA	G	CACTAGGGACAGGCCACTGTTCC		
<i>HEK3</i> +TD136	GGCCCAGACTGAGCACGTGA	TGGGCCTTCCCTGGATCCCTGTCCGTG	13	25
pegRNA		CTCAGTCTG		
<i>HEK3</i> -TD136	GTCCCTGGGATCCCTGTCTC	ATCCTTGGGCCAGACTGAGCACGGAC	13	23
pegRNA	C	AGGGATCCCA		
<i>HEK3</i> +TD222	GGCCCAGACTGAGCACGTGA	AAGGTTTCTCATGCCAAGCTCCCCGTGC	13	23
pegRNA		TCAGTCTG		
<i>HEK3</i> -TD222	GTCTCATGCCAAGCTCCCTG	ATCCTTGGGCCAGACTGAGCACGGG	13	25
pegRNA	C	AGCTTGGCAT		
<i>HEK3</i> +TD397	TAAGACAATGGATACACTGG	TGGGTGCCCTGAGATCTTCCTCTGTTG	13	39
pegRNA		AGCTCGACCGTGTATCCATTGT		
<i>HEK3</i> -TD397	TCTGTTGAGCTGACCCCTGA	TAATATCTCCCCATGTCCCTACATAAGA	13	40
pegRNA		CAATGGATACACGGGTCGAGCTAA		
<i>HEK3</i> +TD499	GTCAACCAGTATCCGGTGC	TAATATCTCCCCATGTCCCTACATAAGA	13	40
pegRNA		CAATGGATACACCCGGGATACTGGT		
<i>HEK3</i> -TD499	TAAGACAATGGATACACTGG	TGGCTCCAGCCAGCCAAACTGTCAA	13	40
pegRNA		CCAGTATCCGGGTGTATCCATTGT		
<i>EMX1</i> +TD5K	GAGTCCGAGCAGAAGAAGA	AGCAGGAAAGGACCCCTCAAATCCTCCT	13	40
pegRNA	A	TTCCATGCCTCTTCTTGCTCGG		
<i>EMX1</i> -TD5K	CTCCTTCCATGCCTCTGAG	AGAAGCTGGAGGAGGAAGGGCCTGAGT	13	40
pegRNA		CCGAGCAGAAGAAAGAGGCATGGAAA		
<i>EMX1</i> +TD10K	GAGTCCGAGCAGAAGAAGA	AGCAAATGTTGCCAGCATAGGGACTCA	13	40
pegRNA	A	GCTCCACATGCTTCTTGCTCGG		
<i>EMX1</i> -TD10K	ACTCAGCTCCCACATGCAGG	AGAAGCTGGAGGAGGAAGGGCCTGAGT	13	40
pegRNA		CCGAGCAGAAGAAGCATGTGGGAGCT		
<i>AAVSI</i> +TD L2R2	GGGGCCACTAGGGACAGGAT	GAGGATGGAGAGGTGGCTAAAGCCAGG	13	40
pegRNA		GAGACGGGGTACTTTCCTGTCCCTAGT		
		GG		
<i>AAVSI</i> -TD L2R2	CAGGGAGACGGGGTACTTTG	ATCTGCCCCCTCCACCCCCACAGTGGGGC	13	40
pegRNA		CACTAGGGACAGGAAAAGTACCCCGTCT		
		C		
<i>AAVSI</i> +TD L4R4	GGGGCCACTAGGGACAGGAT	GAGGATGGAGAGGTGGCTAAAGCCAGG	13	40
pegRNA		GAGACGGGGTACTTGGAAATCCTGTCCC		
		TAGTGG		
<i>AAVSI</i> -TD L4R4	CAGGGAGACGGGGTACTTTG	ATCTGCCCCCTCCACCCCCACAGTGGGGC	13	40
pegRNA		CACTAGGGACAGGATTCCAAAGTACCCC		
		GTCTC		
<i>AAVSI</i> +TD L6R6	GGGGCCACTAGGGACAGGAT	GAGGATGGAGAGGTGGCTAAAGCCAGG	13	40
pegRNA		GAGACGGGGTACTTGGGGCCAATCCTG		
		TCCCTAGTGG		

<i>AAVS1</i> -TD L6R6	CAGGGAGACGGGGTACTTG	ATCTGTCCCCCTCCACCCCCACAGTGGGGC	13	40
pegRNA		CACTAGGGACAGGATTGGCCCCAAAGTA		
		CCCCGTCTC		
<i>AAVS1</i> +TD L8R8	GGGGCCACTAGGGACAGGAT	GAGGATGGAGAGGTGGCTAAAGCCAGG	13	40
pegRNA		GAGACGGGGTACTTGGGTTACCAAT		
		CCTGTCCCTAGTGG		
<i>AAVS1</i> -TD L8R8	CAGGGAGACGGGGTACTTG	ATCTGTCCCCCTCCACCCCCACAGTGGGGC	13	40
pegRNA		CACTAGGGACAGGATTGGTAACCCCAA		
		AGTACCCCGTCTC		
<i>AAVS1</i> +TD L10R10	GGGGCCACTAGGGACAGGAT	GAGGATGGAGAGGTGGCTAAAGCCAGG	13	40
pegRNA		GAGACGGGGTACTTGGGTTGTGTCAC		
		CAATCCTGTCCCTAGTGG		
<i>AAVS1</i> -TD L10R10	CAGGGAGACGGGGTACTTG	ATCTGTCCCCCTCCACCCCCACAGTGGGGC	13	40
pegRNA		CACTAGGGACAGGATTGGTACACAACC		
		CCAAAGTACCCCGTCTC		
<i>HEK3</i> +TD L2R2	GGCCCAGACTGAGCACGTGA	AAGGTTTCTCATGCCAAGCTCCCTGCAC	13	23
pegRNA		GTGCTCAGTCTG		
<i>HEK3</i> -TD L2R2	GTCTCATGCCAAGCTCCCTG	ATCCTGGGGCCCAGACTGAGCACGTGC	13	25
pegRNA	C	AGGGAGCTTGGCAT		
<i>HEK3</i> +TD L4R4	GGCCCAGACTGAGCACGTGA	AAGGTTTCTCATGCCAAGCTCCCTGCAA	13	23
pegRNA		TCACGTGCTCAGTCTG		
<i>HEK3</i> -TD L4R4	GTCTCATGCCAAGCTCCCTG	ATCCTGGGGCCCAGACTGAGCACGTGA	13	25
pegRNA	C	TTGCAGGGAGCTTGGCAT		
<i>HEK3</i> +TD L6R6	GGCCCAGACTGAGCACGTGA	AAGGTTTCTCATGCCAAGCTCCCTGCAG	13	23
pegRNA		GCCATCACGTGCTCAGTCTG		
<i>HEK3</i> -TD L6R6	GTCTCATGCCAAGCTCCCTG	ATCCTGGGGCCCAGACTGAGCACGTGA	13	25
pegRNA	C	TGGCCTGCAGGGAGCTTGGCAT		
<i>HEK3</i> +TD L8R8	GGCCCAGACTGAGCACGTGA	AAGGTTTCTCATGCCAAGCTCCCTGCAG	13	23
pegRNA		GAGCCATCACGTGCTCAGTCTG		
<i>HEK3</i> -TD L8R8	GTCTCATGCCAAGCTCCCTG	ATCCTGGGGCCCAGACTGAGCACGTGA	13	25
pegRNA	C	TGGCAGTCCTGCAGGGAGCTTGGCAT		
<i>HEK3</i> +TD L10R10	GGCCCAGACTGAGCACGTGA	AAGGTTTCTCATGCCAAGCTCCCTGCAG	13	23
pegRNA		GACATTCTGCCATCACGTGCTCAGTCTG		
<i>HEK3</i> -TD L10R10	GTCTCATGCCAAGCTCCCTG	ATCCTGGGGCCCAGACTGAGCACGTGA	13	25
pegRNA	C	TGGCAGAATGTCCCTGCAGGGAGCTTGGC		
		AT		
<i>ENG</i> +180	GTTCCTGCAAGACCTGTTGG	CATGTCTCTTCCCTGGAGTTCCCAACGGT	13	25
pegRNA		GAGTGTCCCGGATGTTCTCCAACAGGT		
		CTTGCAG		
<i>ENG</i> -180	CTTCCTGGAGTTCCCAACGG	AATGGACTGTTCTGCAAGACCTGTTGG	13	25
pegRNA		AGAAACATCCGGGACACTCACCGTTGG		
		AACTCCAG		
<i>SATB2</i> +209	GCTCCATGCTGCTCCGACTC	GGCCGTGGAGGTAACAGCCGGAGCA	13	25
pegRNA		GGGATGTCTCCGAGTCGGAGCAGCATG		

<i>SATB2</i> -209	GAGGTAACAGCCGGAGCA	GCTCCGCCGCTCCATGCTGCTCCGACTC	13	25
pegRNA	G	GGAGACATCCCTGCTCCGGCTGTTA		
<i>NCF2</i> +149	AGGAGAACAGGGCCTGGCCT	GTGAGGTAAGGAGAACAGGGCCTGGTTT	13	25
pegRNA		CTCCCTCTGTCCTGCAGATATG		
<i>NCF2</i> -149	AAATCATATCTGCAGGACAG	TGATAGCCAAATCATATCTGCAGGACAG	13	25
pegRNA		AGGGAGAAACCAGGCCCTGTT		
<i>RYR1</i> +85	TATGGAGGTAGGTATGTCT	TCACCAAAAGCGTCGATGATCAGACCTG	13	25
pegRNA		GGGTGCCATGACCTACCTC		
<i>RYR1</i> -85	AGCGTCGATGATCAGACCTG	AAGGAGGATATGGAGGTAGGTATGGC	13	25
pegRNA		CACCCAGGTCTGATCATCGA		

Supplemental Table 3. Summary of primers for amplification of each target sites.

Target site	Forward primer	Reverse primer	Figures
<i>FANCF</i>	AAGACGCTGGAGATTGACA	CAAAGCGCCGATGGATGTG	Fig. 1B
<i>EMX1</i>	TGGGGGCCTCCTGAGTTT	CTCTGCCCTCGTGGGTTGT	Fig. 1B
<i>FANCF</i>	CATCATCTCGCACGTGGTC	TGCTGACGTAGGTAGTGCTT	Fig. 2B
<i>EMX1</i>	GCTTCCAGAGCCTGCACTCC	TCGATGTCCTCCCCATTGGC	Fig. 2B
<i>AAVS1</i>	GAACTCTGCCCTCTAACGCT	AGAGGTTCTGGCAAGGAGAG	Fig. 2B
<i>HEK3</i>	GTGAGCTGCTGTCCCAGAGG	ATGCAGGTGCTGAAAGCCAC	Fig. 2B
<i>HEK3</i>	CACGTGATGGCAGAGGAAAG	GGCATTGTTATTCCCTCCA	Fig. 2B
<i>HEK3</i>	TTGATGAATCAGTGCTGGAG	GAGCTGCACATACTAGCC	Supplemental
TD136-bp			Fig. S22B
<i>EMX1</i> TD 5- kb	TGGGGGCCTCCTGAGTTT	AGCTCCTCACAGAACCACTC	Fig. 2D
<i>EMX1</i> TD 10- kb	TGGGGGCCTCCTGAGTTT	CAACACACACACTGGAGACT	Fig. 2D
<i>AAVS1</i>	CCTGTGCCATCTCTCGTTTC	AGAGGTTCTGGCAAGGAGAG	Fig. 3B
<i>ENG</i>	CTAACGAGGACTCAGCCACTG	AGTC CCTAGGAGACGTTGGA	Fig. 3C
<i>SATB2</i>	TAATCCTACACCGCGACAGC	TGGCCTGAACGTCTATCACG	Fig. 3C
<i>NCF2</i>	CATGGTCCTTGCGCTTCAC	GGGAAGGGAGAAGCAGACTC	Fig. 3C
<i>RYR1</i>	GCAAGCCCTGGAGGTAGGTA	CACCCCTTCCTAACTGGC	Fig. 3C

Supplemental Table S4. Primers used for qPCR to detect the copy number of target DNA fragments.

Target site	Forward primer	Reverse primer
<i>EMX1</i> TD 5-kb/10-kb flanking (plasmid)	AGGGCTCCCATCACATCAA	ACCGCGTTGCTCTACCAGCC
<i>EMX1</i> TD 5-kb (plasmid)	CTGTGACCCTTGTTGAGA	CACCGTGAGCTAGGAACAAA
<i>EMX1</i> TD 10-kb (plasmid)	CTGCCATCCCCTCTGTGAA	CTTCCCCTTCTCCACCTCT
<i>EMX1</i> TD 5-kb/10-kb flanking	GGCCAATGGGGAGGACATC	TTGTCCCTCTGTCAATGGCG
<i>EMX1</i> TD 5-kb	GGCCTGAGTCCGAGCAGAA	TCCCCCTCTTATCCATCCCTC
<i>EMX1</i> TD 10-kb	CGAGCAGAAGAAGCATGTGGG	GTTTCTGCATGGCGTGTG

Supplemental Table S5. Sequences of primers used for HTS.

sample name	Primer Name	Sequence
<i>FANCF</i> TD 50-bp	<i>FANCF</i> -HTS-1-forward	acacagtAGGCGTATCATT CGCGGAT
	<i>FANCF</i> -HTS-reverse	AAAAGCGATCCAGGTGCTGC
<i>FANCF</i> TD 50-bp	<i>FANCF</i> -HTS-2-forward	acagtcaAGGCGTATCATT CGCGGAT
	<i>FANCF</i> -HTS-reverse	AAAAGCGATCCAGGTGCTGC
<i>FANCF</i> TD 50-bp	<i>FANCF</i> -HTS-3-forward	actctgaAGGCGTATCATT CGCGGAT
	<i>FANCF</i> -HTS-reverse	AAAAGCGATCCAGGTGCTGC
<i>FANCF</i> -CTR	<i>FANCF</i> -HTS-4-forward	gtagtctAGGCGTATCATT CGCGGAT
	<i>FANCF</i> -HTS-reverse	AGGTAGCGCGCCC ACTGCAA
<i>FANCF</i> -CTR	<i>FANCF</i> -HTS-5-forward	gtcacatAGGCGTATCATT CGCGGAT
	<i>FANCF</i> -HTS-reverse	AGGTAGCGCGCCC ACTGCAA
<i>FANCF</i> -CTR	<i>FANCF</i> -HTS-6-forward	gtctagtAGGCGTATCATT CGCGGAT
	<i>FANCF</i> -HTS-reverse	AGGTAGCGCGCCC ACTGCAA
<i>EMX1</i> TD 57-bp	<i>EMX1</i> -HTS-1-forward	catcagaTGGGGGCCTCCTGAGTTT
	<i>EMX1</i> -HTS-reverse	TGATGTGATGGGAGCCCTTC
<i>EMX1</i> TD 57-bp	<i>EMX1</i> -HTS-2-forward	catgtctTGGGGGCCTCCTGAGTTT
	<i>EMX1</i> -HTS-reverse	TGATGTGATGGGAGCCCTTC
<i>EMX1</i> TD 57-bp	<i>EMX1</i> -HTS-3-forward	cacaagtTGGGGGCCTCCTGAGTTT
	<i>EMX1</i> -HTS-reverse	TGATGTGATGGGAGCCCTTC
<i>EMX1</i> -CTR	<i>EMX1</i> -HTS-4-forward	acacgtTGGGGGCCTCCTGAGTTT
	<i>EMX1</i> -HTS-reverse	TGATGTGATGGGAGCCCTTC
<i>EMX1</i> -CTR	<i>EMX1</i> -HTS-5-forward	acagatcTGGGGGCCTCCTGAGTTT
	<i>EMX1</i> -HTS-reverse	TGATGTGATGGGAGCCCTTC
<i>EMX1</i> -CTR	<i>EMX1</i> -HTS-6-forward	acagcatTGGGGGCCTCCTGAGTTT
	<i>EMX1</i> -HTS-reverse	TGATGTGATGGGAGCCCTTC
<i>AAVS1</i> TD 376-bp	<i>AAVS1</i> TD-376 -HTS -1-forward	cttgagtATTCCCAGGGCCGGTTAATG
	<i>AAVS1</i> TD-376 -reverse	GTGACCCGAATCCACAGGAG
<i>AAVS1</i> TD 376-bp	<i>AAVS1</i> TD-376 -HTS -2-forward	ctcaugaATTCCCAGGGCCGGTTAATG
	<i>AAVS1</i> TD-376 -reverse	GTGACCCGAATCCACAGGAG
<i>AAVS1</i> TD 376-bp	<i>AAVS1</i> TD-376 -HTS -3-forward	ctcagacATTCCCAGGGCCGGTTAATG
	<i>AAVS1</i> TD-376 -reverse	GTGACCCGAATCCACAGGAG
<i>AAVS1</i> TD 532-bp	<i>AAVS1</i> TD-532 -HTS -1-forward	ctctcagATTCCCAGGGCCGGTTAATG
	<i>AAVS1</i> TD-532 -reverse	GAGACATCCGTCGGAGAAGG
<i>AAVS1</i> TD 532-bp	<i>AAVS1</i> TD-532 -HTS -2-forward	ctctgtaATTCCCAGGGCCGGTTAATG
	<i>AAVS1</i> TD-532 -reverse	GAGACATCCGTCGGAGAAGG
<i>AAVS1</i> TD 532-bp	<i>AAVS1</i> TD-532 -HTS -3-forward	ctgatgtATTCCCAGGGCCGGTTAATG
	<i>AAVS1</i> TD-532 -reverse	GAGACATCCGTCGGAGAAGG
<i>AAVS1</i> TD 846-bp	<i>AAVS1</i> TD-846-HTS -1-forward	ctgtagaATTCCCAGGGCCGGTTAATG
	<i>AAVS1</i> TD-846 -reverse	CAAGCTGCAGACAGAAAGCG
<i>AAVS1</i> TD 846-bp	<i>AAVS1</i> TD-846 -HTS -2-forward	gaacactATTCCCAGGGCCGGTTAATG
	<i>AAVS1</i> TD-846 -reverse	CAAGCTGCAGACAGAAAGCG
<i>AAVS1</i> TD 846-bp	<i>AAVS1</i> TD-846 -HTS -3-forward	gatctcaATTCCCAGGGCCGGTTAATG
	<i>AAVS1</i> TD-846-reverse	CAAGCTGCAGACAGAAAGCG

<i>EMX1</i> TD 116-bp	<i>EMX1</i> TD-116-HTS -1-forward	cacactgTCCAGAACCGGAGGACAAAG
	<i>EMX1</i> TD-116-reverse	ACCACACCTTCACCTGGG
<i>EMX1</i> TD 116-bp	<i>EMX1</i> TD-116-HTS -2-forward	cacttgaTCCAGAACCGGAGGACAAAG
	<i>EMX1</i> TD-116-reverse	ACCACACCTTCACCTGGG
<i>EMX1</i> TD 116-bp	<i>EMX1</i> TD-116-HTS -3-forward	cactgatTCCAGAACCGGAGGACAAAG
	<i>EMX1</i> TD-116-reverse	ACCACACCTTCACCTGGG
<i>EMX1</i> TD 201-bp	<i>EMX1</i> TD-201-HTS -1-forward	cagatcaTGGGGGCCTCCTGAGTTT
	<i>EMX1</i> TD-201-reverse	CCTCAACACTCAGGCTGAGC
<i>EMX1</i> TD 201-bp	<i>EMX1</i> TD-201-HTS -2-forward	cagacatTGGGGGCCTCCTGAGTTT
	<i>EMX1</i> TD-201-reverse	CCTCAACACTCAGGCTGAGC
<i>EMX1</i> TD 201-bp	<i>EMX1</i> TD-201-HTS -3-forward	cagagtcTGGGGGCCTCCTGAGTTT
	<i>EMX1</i> TD-201-reverse	CCTCAACACTCAGGCTGAGC
<i>EMX1</i> TD 307-bp	<i>EMX1</i> TD-307-HTS -1-forward	cagtactGTTTCTCATCTGTGCCCTCC
	<i>EMX1</i> TD-307-reverse	GCTCCCATGGGTCTAACATTTC
<i>EMX1</i> TD 307-bp	<i>EMX1</i> TD-307-HTS -2-forward	cagtctaGTTTCTCATCTGTGCCCTCC
	<i>EMX1</i> TD-307-reverse	GCTCCCATGGGTCTAACATTTC
<i>EMX1</i> TD 307-bp	<i>EMX1</i> TD-307-HTS -3-forward	ctagacaGTTTCTCATCTGTGCCCTCC
	<i>EMX1</i> TD-307-reverse	GCTCCCATGGGTCTAACATTTC
<i>FANCF</i> TD 115-bp	<i>FANCF</i> TD-115-HTS -1-forward	cagtactATAGCATTGCAGAGAGGC GT
	<i>FANCF</i> TD-115-reverse	GGTCGAAATGCATGTCAATC
<i>FANCF</i> TD 115-bp	<i>FANCF</i> TD-115-HTS -1-forward	cagtactATAGCATTGCAGAGAGGC GT
	<i>FANCF</i> TD-115-reverse	GGTCGAAATGCATGTCAATC
<i>FANCF</i> TD 115-bp	<i>FANCF</i> TD-115-HTS -1-forward	cagtactATAGCATTGCAGAGAGGC GT
	<i>FANCF</i> TD-115-reverse	GGTCGAAATGCATGTCAATC
<i>FANCF</i> TD 171-bp	<i>FANCF</i> TD-171-HTS -1-forward	tgtcagtATAGCATTGCAGAGAGGC GT
	<i>FANCF</i> TD-171-reverse	GGTCGAAATGCATGTCAATC
<i>FANCF</i> TD 171-bp	<i>FANCF</i> TD-171-HTS -1-forward	tgtgtcaATAGCATTGCAGAGAGGC GT
	<i>FANCF</i> TD-171-reverse	GGTCGAAATGCATGTCAATC
<i>FANCF</i> TD 171-bp	<i>FANCF</i> TD-171-HTS -1-forward	caactgtATAGCATTGCAGAGAGGC GT
	<i>FANCF</i> TD-171-reverse	GGTCGAAATGCATGTCAATC
<i>FANCF</i> TD 231-bp	<i>FANCF</i> TD-231-HTS -1-forward	catcagaCGCTGGGAGATTGACATGCA
	<i>FANCF</i> TD-231-reverse	CCGTGTTCTTGACTCTGGG
<i>FANCF</i> TD 231-bp	<i>FANCF</i> TD-231-HTS -1-forward	catgtctCGCTGGGAGATTGACATGCA
	<i>FANCF</i> TD-231-reverse	CCGTGTTCTTGACTCTGGG
<i>FANCF</i> TD 231-bp	<i>FANCF</i> TD-231-HTS -1-forward	cacaagtCGCTGGGAGATTGACATGCA
	<i>FANCF</i> TD-231-reverse	CCGTGTTCTTGACTCTGGG
<i>HEK3</i> TD 136-bp	<i>HEK3</i> TD-136-HTS -1-forward	gacatctGCATTGTAGGCTTGATGCT
	<i>HEK3</i> TD-136-reverse	ATGCTTACTTGCAGCAGAAA
<i>HEK3</i> TD 136-bp	<i>HEK3</i> TD-136-HTS -2-forward	gacagtaGCATTGTAGGCTTGATGCT
	<i>HEK3</i> TD-136-reverse	ATGCTTACTTGCAGCAGAAA
<i>HEK3</i> TD 136-bp	<i>HEK3</i> TD-136-HTS -3-forward	gactacaGCATTGTAGGCTTGATGCT
	<i>HEK3</i> TD-136-reverse	ATGCTTACTTGCAGCAGAAA
<i>HEK3</i> TD 222-bp	<i>HEK3</i> TD-222-HTS -1-forward	gactcacGCATTGTAGGCTTGATGCT
	<i>HEK3</i> TD-222-reverse	ATGCTTACTTGCAGCAGAAA
<i>HEK3</i> TD 222-bp	<i>HEK3</i> TD-222-HTS -2-forward	gagtctgtGCATTGTAGGCTTGATGCT

	<i>HEK3</i> TD-222-reverse	ATGCTTACTTGCAGCAGAAA
<i>HEK3</i> TD 222-bp	<i>HEK3</i> TD-222-HTS -3-forward	gtacagaGCATTGTAGGCTTGATGCT
	<i>HEK3</i> TD-222-reverse	ATGCTTACTTGCAGCAGAAA
<i>HEK3</i> TD 397-bp	<i>HEK3</i> TD-397-HTS -1-forward	gtagtctTGAGCACTTGTACACGCAG
	<i>HEK3</i> TD-397-reverse	CTGATTTCATGCAGGTGCT
<i>HEK3</i> TD 397-bp	<i>HEK3</i> TD-397-HTS -2-forward	gtcacatTGAGCACTTGTACACGCAG
	<i>HEK3</i> TD-397-reverse	CTGATTTCATGCAGGTGCT
<i>HEK3</i> TD 397-bp	<i>HEK3</i> TD-397-HTS -3-forward	gtcttagTGAGCACTTGTACACGCAG
	<i>HEK3</i> TD-397-reverse	CTGATTTCATGCAGGTGCT
<i>HEK3</i> TD 499-bp	<i>HEK3</i> TD-499-HTS -1-forward	gtcgtgaGTTACACGCAGGGCACTATG
	<i>HEK3</i> TD-499-reverse	TGTTGAGCTCGACCCTGAAG
<i>HEK3</i> TD 499-bp	<i>HEK3</i> TD-499-HTS -2-forward	gtgaacaGTTACACGCAGGGCACTATG
	<i>HEK3</i> TD-499-reverse	TGTTGAGCTCGACCCTGAAG
<i>HEK3</i> TD 499-bp	<i>HEK3</i> TD-499-HTS -3-forward	gtgaetcGTTACACGCAGGGCACTATG
	<i>HEK3</i> TD-499-reverse	TGTTGAGCTCGACCCTGAAG
<i>EMX1</i> TD 5-kb	<i>EMX1</i> TD 5kb-HTS-1-forward	atctgctTCCAGAACCGGAGGACAAAG
	<i>EMX1</i> TD 5kb-reverse	TCCCCCTCTTATCCATCCCTC
<i>EMX1</i> TD 5-kb	<i>EMX1</i> TD 5kb-HTS-2-forward	atgtgacTCCAGAACCGGAGGACAAAG
	<i>EMX1</i> TD 5kb-reverse	TCCCCCTCTTATCCATCCCTC
<i>EMX1</i> TD 5-kb	<i>EMX1</i> TD 5kb-HTS-3-forward	atgcactTCCAGAACCGGAGGACAAAG
	<i>EMX1</i> TD 5kb-reverse	TCCCCCTCTTATCCATCCCTC
<i>EMX1</i> TD 10-kb	<i>EMX1</i> TD 10kb-HTS-1-forward	acaactgGTTCCAGAACCGGAGGACAAA
	<i>EMX1</i> TD 10kb-reverse	CCAGCCTCCCTACTGTCTTTTT
<i>EMX1</i> TD 10-kb	<i>EMX1</i> TD 10kb-HTS-2-forward	acatgaaGTTCCAGAACCGGAGGACAAA
	<i>EMX1</i> TD 10kb-reverse	CCAGCCTCCCTACTGTCTTTTT
<i>EMX1</i> TD 10-kb	<i>EMX1</i> TD 10kb-HTS-3-forward	acatgagGTTCCAGAACCGGAGGACAAA
	<i>EMX1</i> TD 10kb-reverse	CCAGCCTCCCTACTGTCTTTTT

Supplemental Reference

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