

# ***MYC* overexpression leads to increased chromatin interactions at superenhancers and MYC binding sites**

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## Supplemental Tables

Supplemental Table S1 | SIQHiC sequencing statistics for Low *MYC* and High *MYC* cells.

	Low <i>MYC</i> R1	Low <i>MYC</i> R2	High <i>MYC</i> R1	High <i>MYC</i> R2
Total Reads	466,371,873	473,073,105	489,552,289	466,324,553
Total Mapped Reads (Human hg19 ONLY)	449,798,985 96.4%	457,408,641 96.7%	483,099,602 98.7%	456,793,366 98.0%
Total Mapped Reads (Human hg19 + Mouse mm10)	463,092,913 99.3%	469,974,590 99.3%	486,308,738 99.3%	462,613,290 99.2%
Discarded Reads	1,097,259 0.2%	1,085,736 0.2%	826,538 0.2%	927,729 0.2%
Human-Human Paired Reads	448,297,483 96.1%	455,970,215 96.4%	482,156,004 98.5%	455,706,205 97.7%
Mouse-Mouse Paired Reads	13,698,171 2.9%	12,918,639 2.7%	3,326,196 0.7%	5,979,356 1.3%
Human-Human Paired Reads	448,297,483	455,970,215	482,156,004	455,706,205
Human Unique Reads	310,510,965 69.3%	322,597,046 70.7%	302,156,580 62.7%	328,864,586 72.2%
Intra-Fragment Reads	721,931	383,939	1,659,119	455,654
Below MAPQ30 Threshold	43,677,206	47,649,671	39,799,686	47,840,823
Hi-C Contacts	266,111,828 85.7%	274,563,436 85.1%	260,697,775 86.3%	280,568,109 85.3%
Inter-chromosomal	22,188,336	29,460,191	11,484,151	25,330,251
Intra-chromosomal	243,923,492 91.7%	245,103,245 89.3%	249,213,624 95.6%	255,237,858 91.0%
Short (<20kb)	131,525,754	106,418,212	183,310,972	113,572,967
Long (>20kb)	112,397,575	138,684,836	65,902,548	141,664,640
Mouse-Mouse Paired Reads	13,698,171	12,918,639	3,326,196	5,979,356
Mouse Unique Reads	9,591,745 70.0%	9,195,144 71.2%	2,137,464 64.3%	4,361,700 72.9%
Intra-Fragment Reads	34,711	16,340	20,531	9,946
Below MAPQ30 Threshold	1,584,670	1,644,047	342,276	757,343
Hi-C Contacts	7,972,364 83.1%	7,534,757 81.9%	1,774,657 83.0%	3,594,411 82.4%
Inter-chromosomal	841,107	819,222	143,395	380,435
Intra-chromosomal	7,131,257 89.4%	6,715,535 89.1%	1,631,262 91.9%	3,213,976 89.4%
Short (<20kb)	3,898,627	3,186,260	1,153,069	1,574,659
Long (>20kb)	3,232,626	3,529,269	478,192	1,639,311

**Supplemental Table S2 | Calculation of Low MYC and High MYC SIQHiC Ratio.**

	<b>Total reads</b>	<b>Human Contacts</b>	<b>Mouse Contacts</b>	<b>Human Contacts / Mouse Contacts (HMR)</b>	<b>HMR Fold Change vs. Low MYC</b>	<b>SIQHiC Ratio</b>
<b>Low MYC Replicate 1</b>	466,371,873	266,111,828	7,972,364	33.379	1.000	0.227
<b>Low MYC Replicate 2</b>	473,073,105	274,563,436	7,534,757	36.440	1.000	0.467
<b>High MYC Replicate 1</b>	489,552,289	260,697,775	1,774,657	146.900	4.401	1.000
<b>High MYC Replicate 2</b>	466,324,553	280,568,109	3,594,411	78.057	2.142	1.000
				100.812	2.891	1.000

**Supplemental Table S3 | SIQHiC sequencing statistics for siControl and siCTCF cells.**

	siControl_R1	siControl_R2	siCTCF_R1	siCTCF_R2
Total Reads	486,430,445	471,799,212	494,076,632	464,953,616
Total Mapped Reads (Human hg19 ONLY)	478,571,646 98.4%	465,466,367 98.7%	478,635,584 96.9%	452,965,855 97.4%
Total Mapped Reads (Human hg19 + Mouse mm10)	484,412,681 99.6%	469,633,466 99.5%	492,190,679 99.6%	463,577,777 99.7%
Discarded Reads	8,946,230 1.8%	8,676,392 1.8%	9,467,542 1.9%	8,385,251 1.8%
Human-Human Paired Reads	456,579,962 94.3%	446,730,331 95.1%	444,058,460 90.2%	421,007,523 90.8%
Mouse-Mouse Paired Reads	18,886,489 3.9%	14,226,743 3.0%	38,664,677 7.9%	34,185,003 7.4%
Human-Human Paired Reads	456,579,962	446,730,331	444,058,460	421,007,523
Human Unique Reads	350,524,868 76.8%	349,173,808 78.2%	339,280,375 76.4%	324,857,931 77.2%
Intra-Fragment Reads	1,432,009	1,406,054	791,344	867,189
Below MAPQ30 Threshold	48,226,797	47,048,482	50,259,186	47,455,721
Hi-C Contacts	300,866,062 85.8%	300,719,272 86.1%	288,229,845 85.0%	276,535,021 85.1%
Inter-chromosomal	20,419,370	20,936,012	24,006,698	22,347,537
Intra-chromosomal	280,446,692 93.2%	279,783,260 93.0%	264,223,147 91.7%	254,187,484 91.9%
Short (<20kb)	174,860,580	172,296,248	132,033,769	127,643,655
Long (>20kb)	105,584,341	107,485,122	132,188,499	126,543,115
Mouse-Mouse Paired Reads	18,886,489	14,226,743	38,664,677	34,185,003
Mouse Unique Reads	14,686,344 77.8%	11,244,958 79.0%	29,608,422 76.6%	26,428,013 77.3%
Intra-Fragment Reads	77,091	59,623	84,404	89,673
Below MAPQ30 Threshold	2,291,250	1,723,683	5,061,344	4,473,166
Hi-C Contacts	12,318,003 83.9%	9,461,652 84.1%	24,462,674 82.6%	21,865,174 82.7%
Inter-chromosomal	792,851	629,814	1,868,727	1,664,169
Intra-chromosomal	11,525,152 93.6%	8,831,838 93.3%	22,593,947 92.4%	20,201,005 92.4%
Short (<20kb)	7,247,274	5,494,816	11,458,406	10,285,651
Long (>20kb)	4,277,798	3,336,949	11,135,460	10,285,651

**Supplemental Table S4 | Calculation of siControl and siCTCF SIQHIC Ratio.**

	Total reads	Human Contacts	Mouse Contacts	Human Contacts / Mouse Contacts	HMR Fold Change vs. siCTCF	SIQHIC Ratio
siControl Replicate 1	486,430,444	300,866,062	12,318,003	24.425	2.073	1.000
siControl Replicate 2	471,799,212	300,719,272	9,461,652	31.783	2.513	1.000
siCTCF Replicate 1	494,076,632	288,229,845	24,462,674	11.782	1.000	0.482
siCTCF Replicate 2	464,953,616	276,535,021	21,865,174	12.647	1.000	0.398

**Supplemental Table S5 | RT-qPCR and 4C primer sequences.**

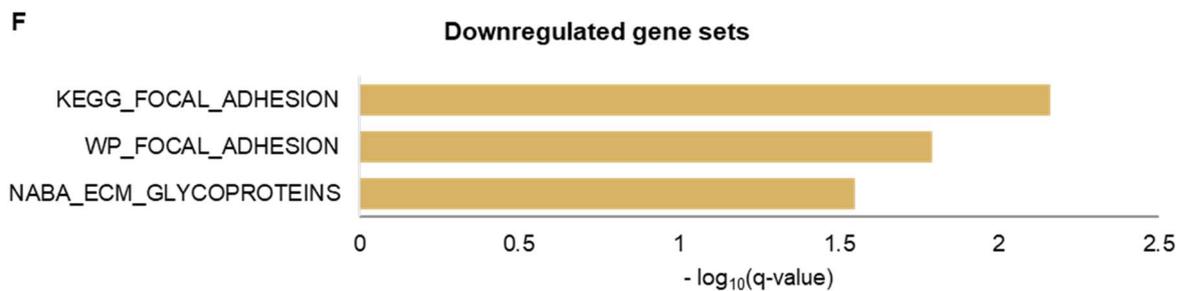
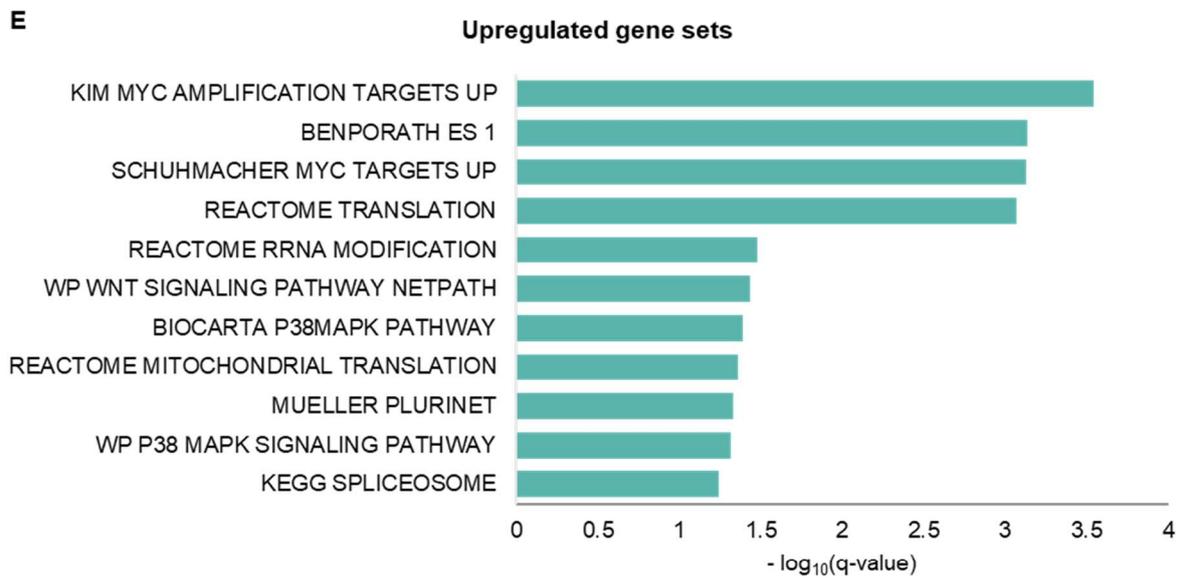
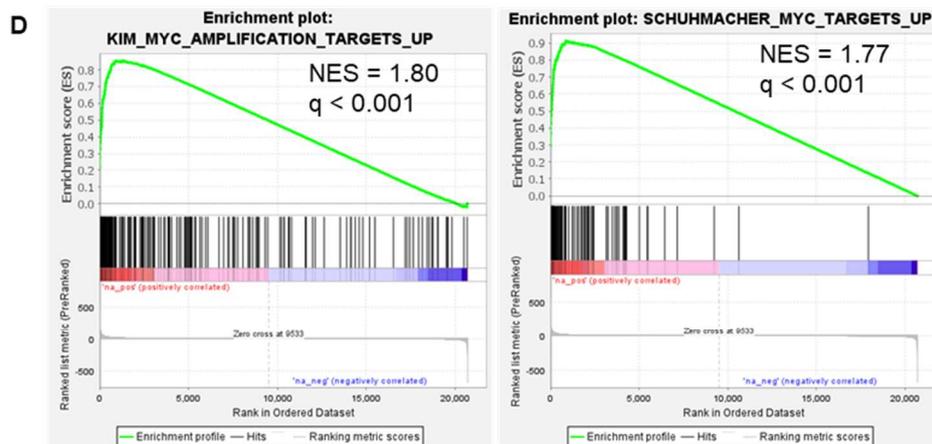
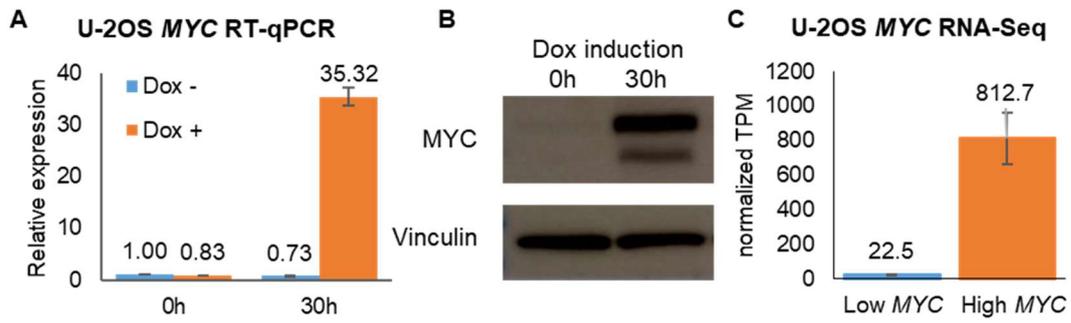
RT-qPCR Primers	Sequence
<i>TBP</i> _F	TAATCCCAAGCGATTTGCTG
<i>TBP</i> _R	CAGTTGTCCGTGGCTCTCTT
<i>MYC</i> _F	CACCAGCAGCGACTCTGA
<i>MYC</i> _R	GATCCAGACTCTGACCTTTTGC
<i>CTCF</i> _F	CAAACACACCCACGAGAA
<i>CTCF</i> _R	CTCCAGTATGAGAGCGAATG

4C Viewpoint	4C Primer	Sequence
Chr2_PROC_SE	Outer F	TTAGCCACGAAGTGCCTGC
	Outer R	AGGAGAACTGTTTTTCATTGCCATG
	Nested F	AATGATACGGCGACCACCGAGATCTACACXXXXXXXXTCG TCGGCAGCGTCAGATGTGTATAAGAGACAGGGTGGAGTT
		TCCTCCCTGT
	Nested R	CAAGCAGAAGACGGCATAACGAGATYYYYYYYGTCTCGTG GGCTCGGAGATGTGTATAAGAGACAGTGATTGGATGCAG CACTTGG

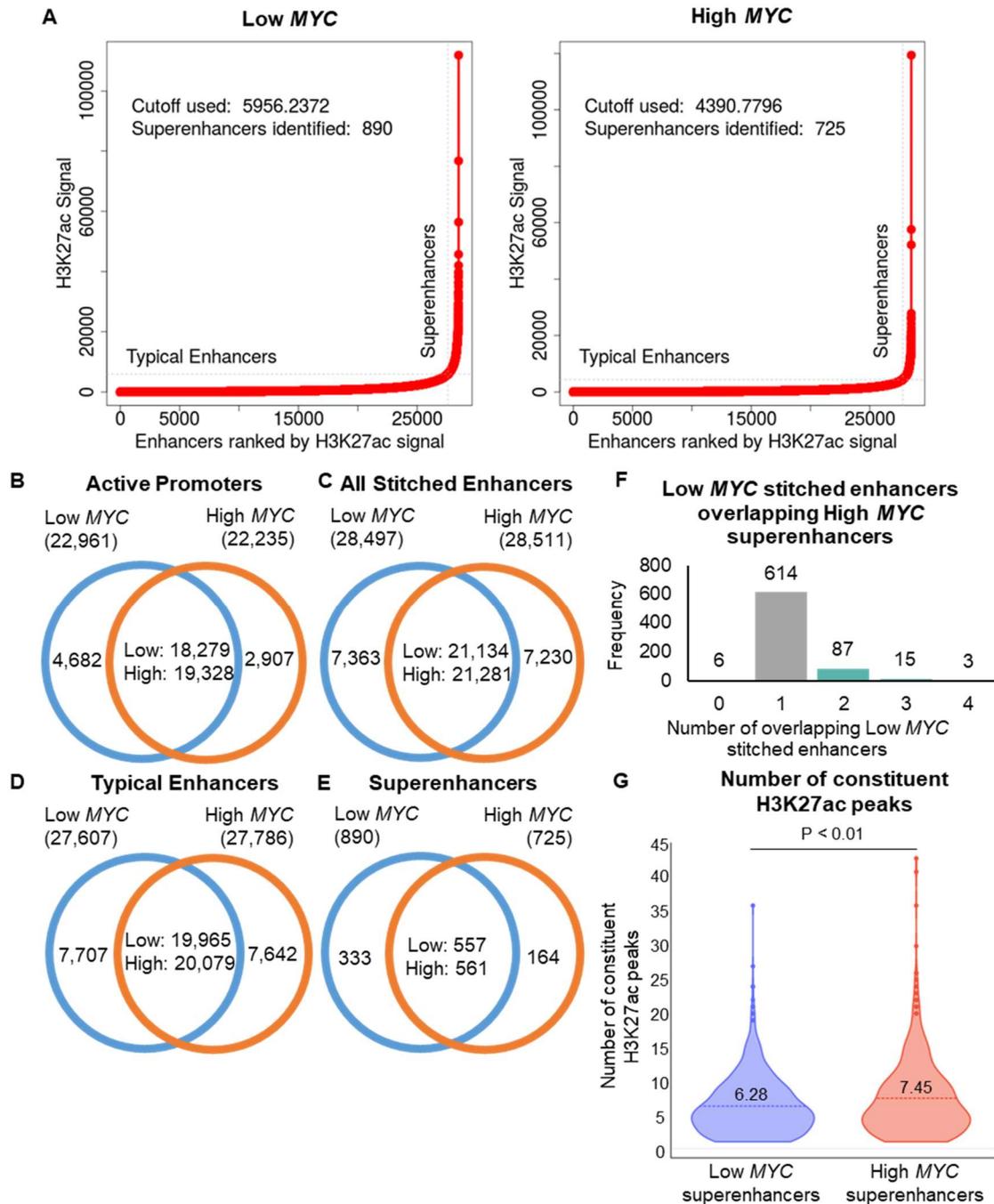
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# Supplemental Figures

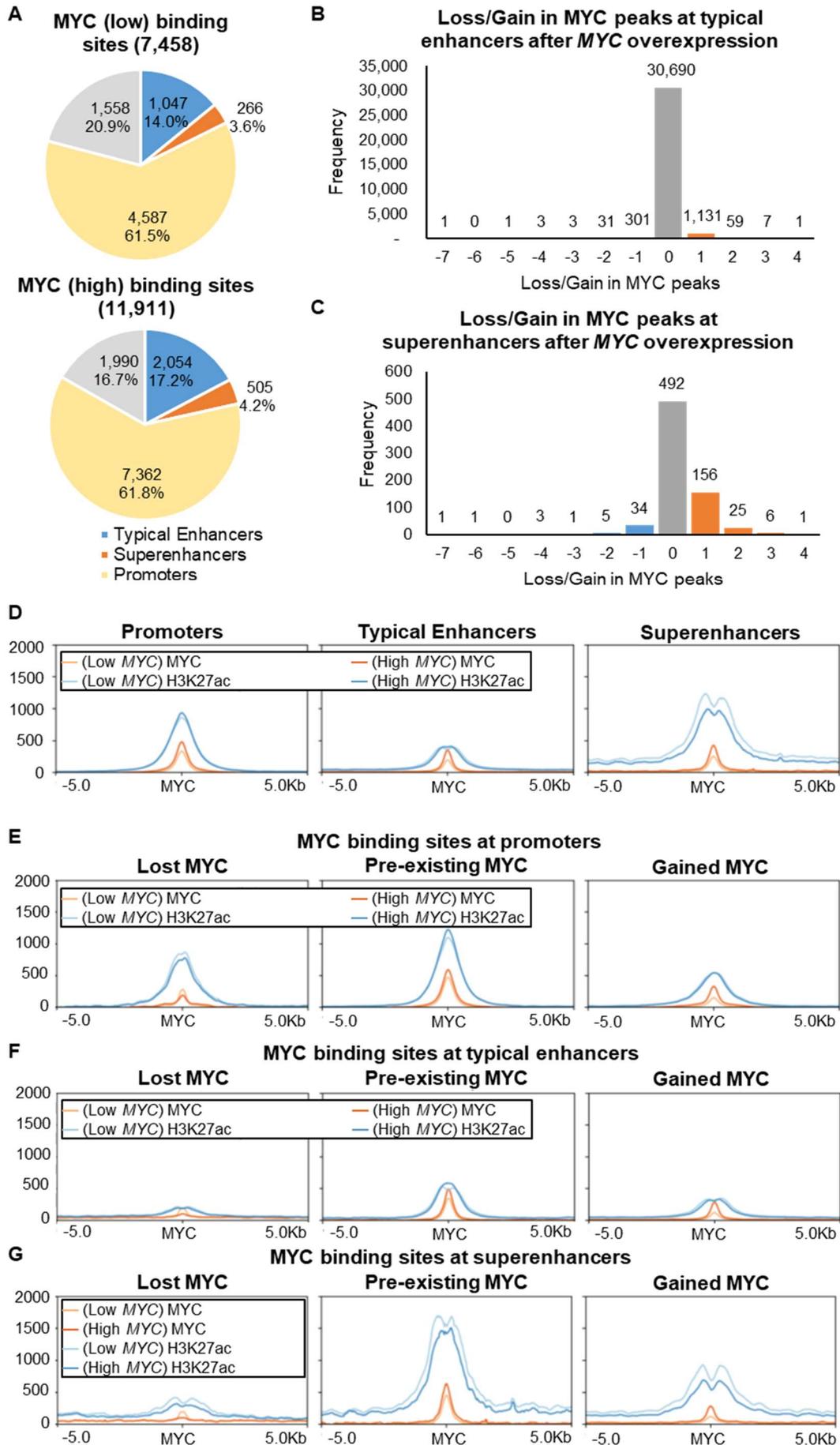


**Supplemental Figure S1 | *MYC* expression and regulated genes.** (A) *MYC* gene expression in U2OS cells before and after doxycycline (Dox) induction, normalized against *TBP* gene. Error bars show standard error across 3 biological replicates. (B) Western blot showing *MYC* protein expression with and without doxycycline (Dox) induction. Vinculin is shown as a loading control. (C) RNA-Seq normalized transcripts per million (TPM) expression of *MYC* in U2OS Low *MYC* and High *MYC* cells. (D) Gene set enrichment analysis (GSEA) plots of 2 previously published *MYC* regulated gene sets. (E) Top upregulated gene sets after *MYC* overexpression. (F) Top downregulated gene sets after *MYC* overexpression.

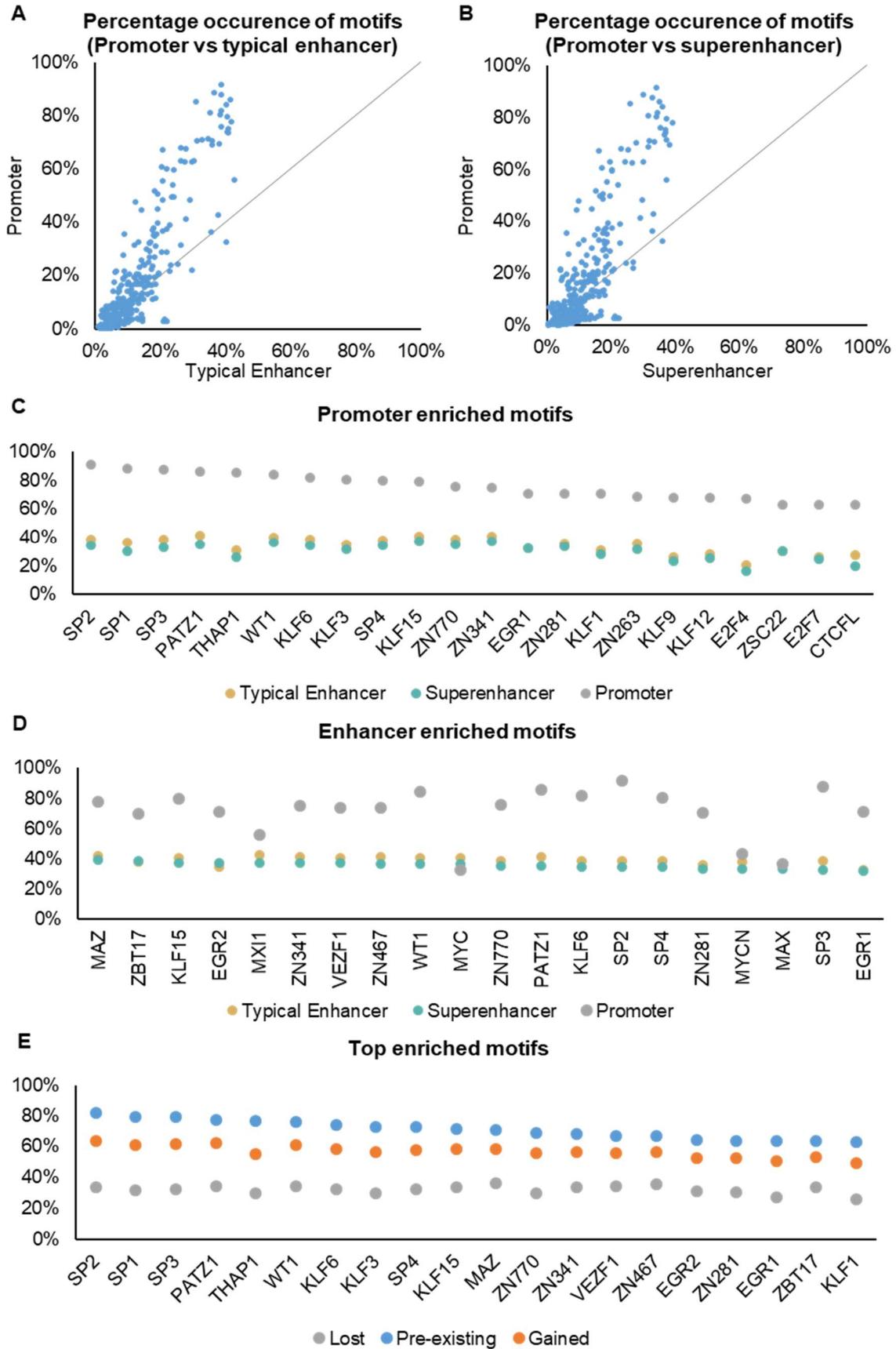


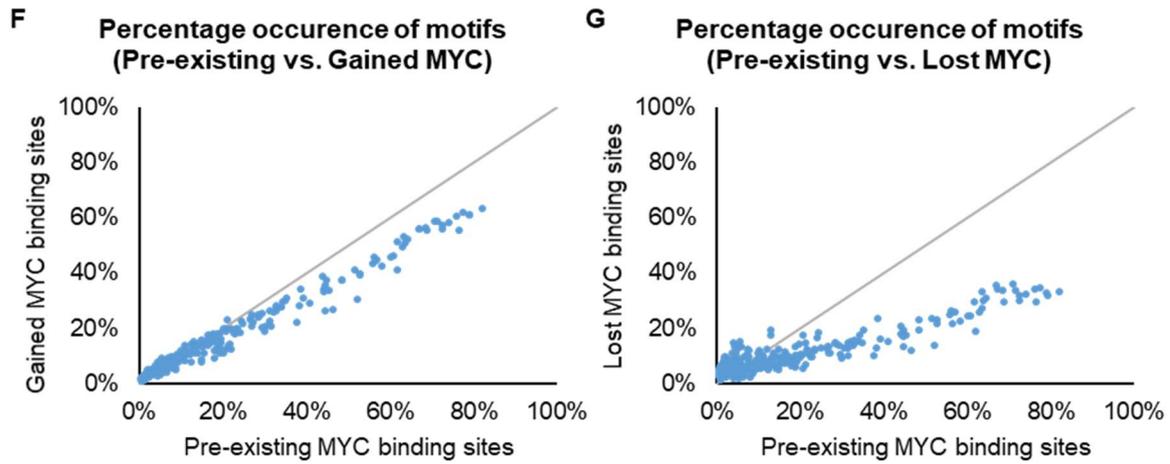
**Supplemental Figure S2 | Enhancer and superenhancers.** (A) Identification of Low *MYC* (left) and High *MYC* (right) superenhancers by ranking stitched enhancers based on H3K27ac (see Methods). (B-E) Overlap of (B) active promoter peaks, (C) all stitched enhancers, (D) typical enhancers and (E) superenhancers between Low *MYC* and High *MYC* cells. (F) Number of Low *MYC* stitched enhancers overlapping High *MYC* superenhancers. (G) Violin plot showing number of constituent H3K27ac

peaks merged with Low MYC and High MYC superenhancers. P values were calculated using two-tailed t-test assuming unequal variances.

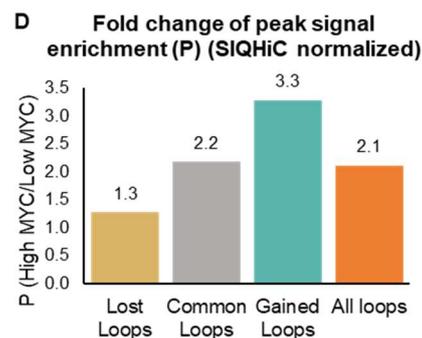
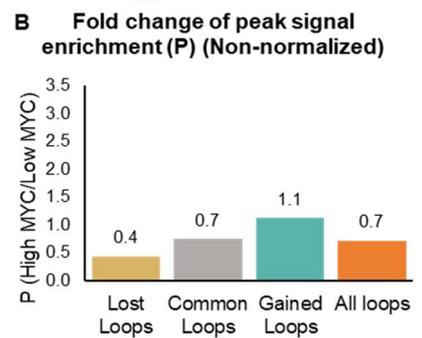
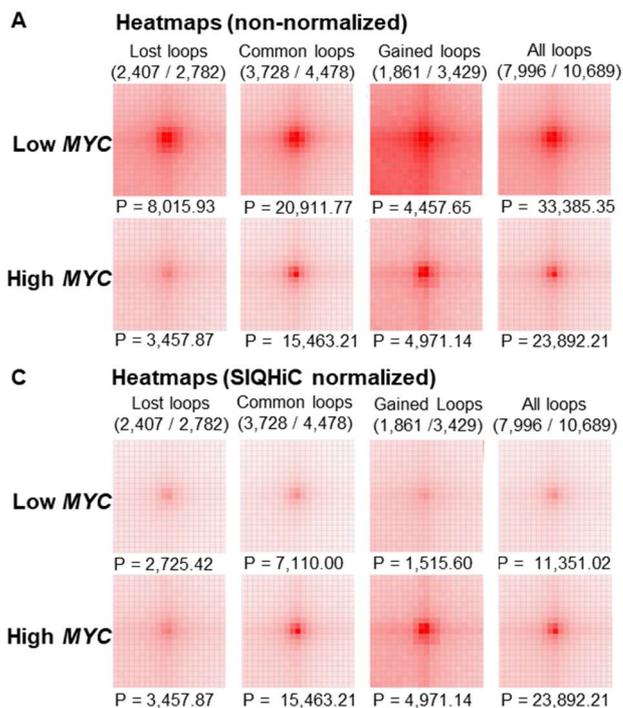
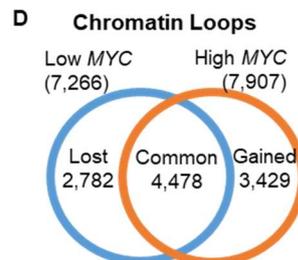
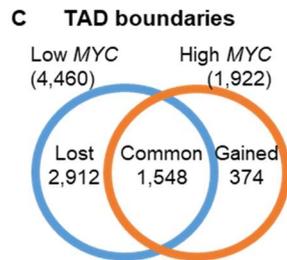
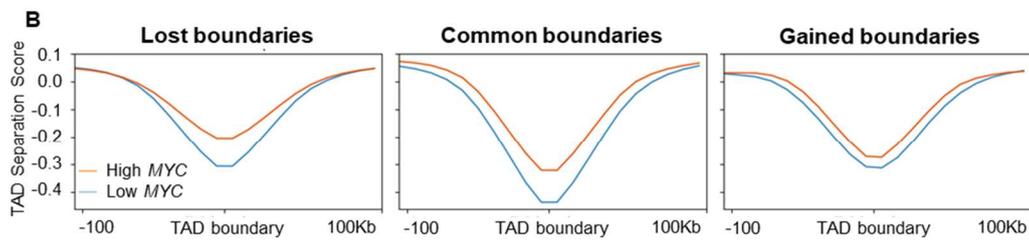
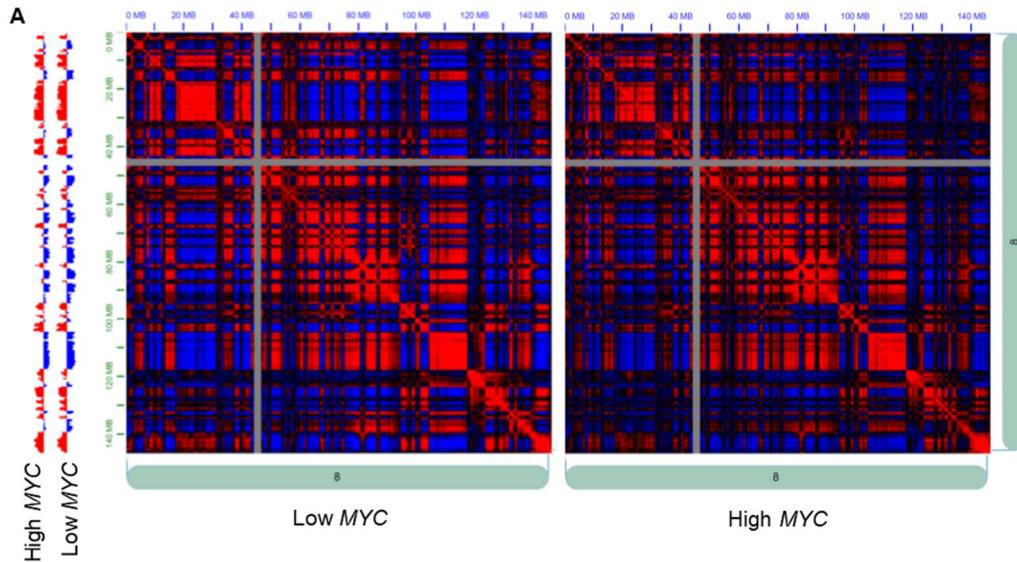


**Supplemental Figure S3 | MYC binding sites at promoters and enhancers.** (A) Distribution of MYC binding sites at cis-regulatory elements in Low *MYC* (left) and High *MYC* (right) cells. (B-C) Gain and loss of MYC peaks at (B) typical enhancers and (C) superenhancers after *MYC* overexpression. (D) H3K27ac and MYC ChIP-seq signal at MYC binding sites on promoters, typical enhancers and superenhancers. (E-G) H3K27ac and MYC ChIP-seq signal at lost, pre-existing and gained MYC binding sites on (E) typical enhancers, (F) superenhancers and (G) promoters.



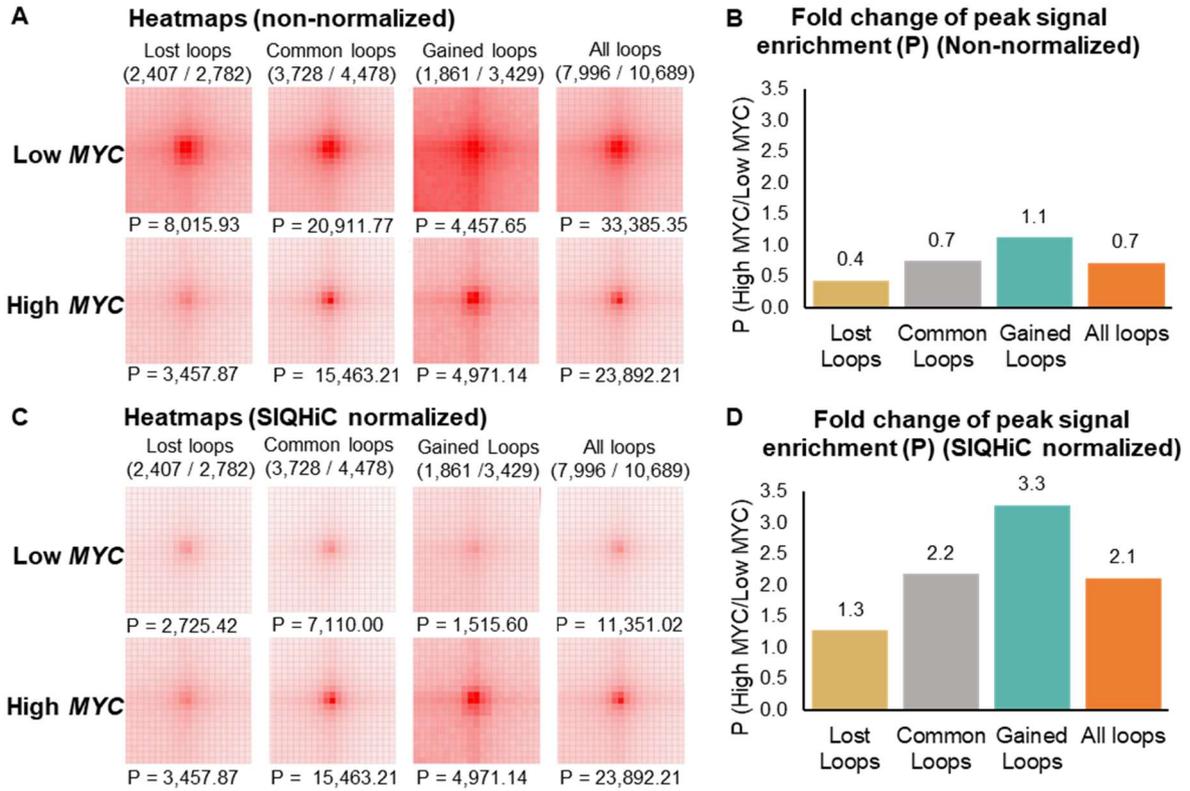


**Supplemental Figure S4 | Motif occurrence at MYC binding sites.** (A-B) Percentage occurrence of HOCOMOCO transcription factor motifs at promoter MYC binding sites versus (A) typical enhancer and (B) superenhancer MYC binding sites. Each point represents 1 motif. The grey line shows equal percentage occurrence. (C-D) Top enriched HOCOMOCO transcription factor motifs occurring at (C) promoter and (D) enhancer MYC binding sites. (E) Top enriched motifs at lost, pre-existing and gained MYC binding sites. (F-G) Percentage occurrence of HOCOMOCO transcription factor motifs at (F) gained and (G) lost MYC binding sites versus pre-existing MYC binding sites. Each point represents 1 motif. The grey line shows equal percentage occurrence.

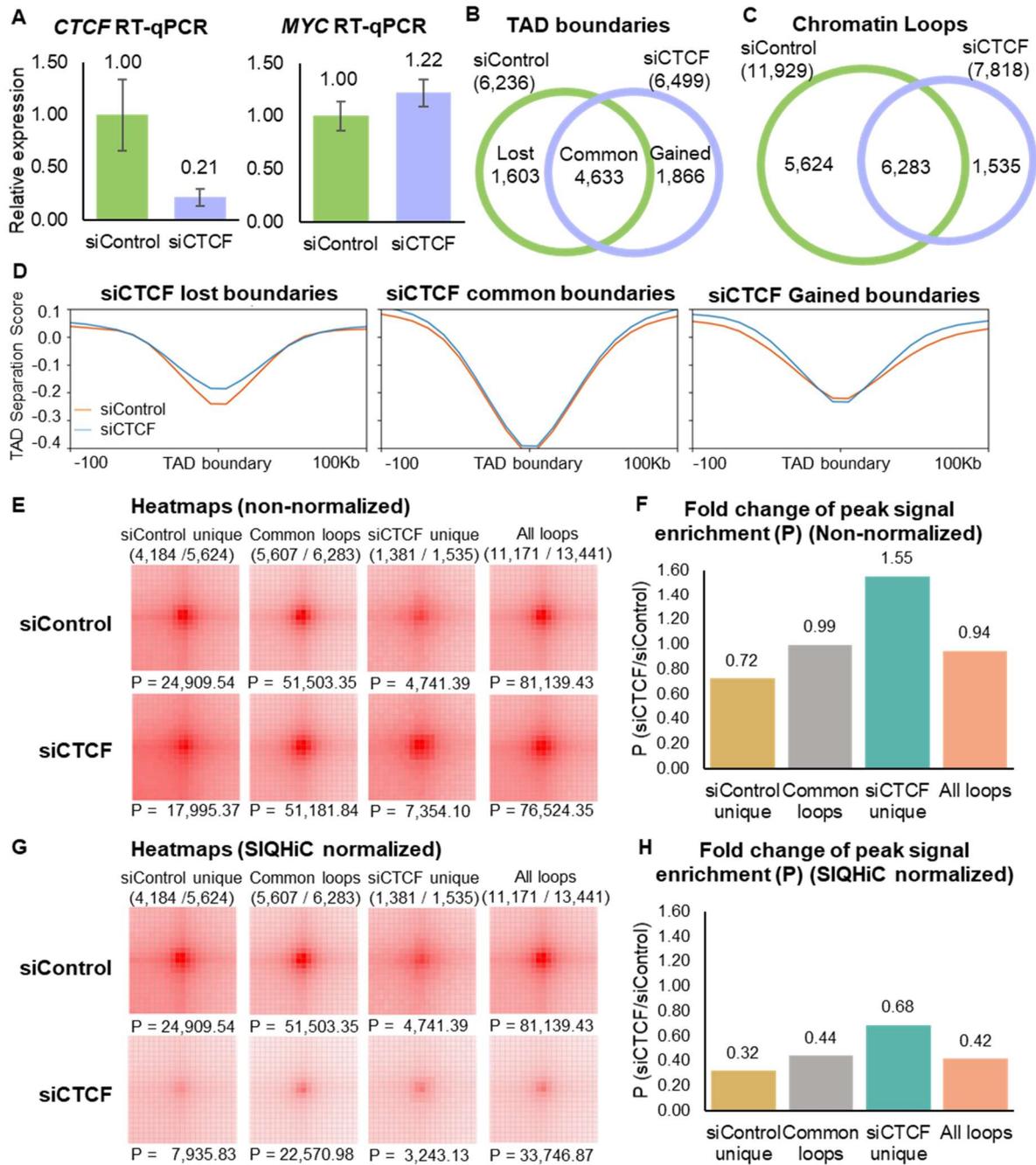


**Supplemental Figure S5 | A/B compartments, topological associated domains and loops**

**identified using SIQHiC.** (A) A/B compartments are not altered after *MYC* overexpression, as shown in a representative view of chromosome 8. Pearson's correlation heatmaps were drawn using first principal component (PC1) at 100kb resolution, shown as tracks above the heatmaps. (B) Topologically associated domain (TAD) separation scores of lost, common and gained TAD boundaries in Low *MYC* and High *MYC* cells. (C) Overlap between TAD boundaries identified in Low *MYC* and High *MYC* cells. (D) Overlap between chromatin loops identified in Low *MYC* and High *MYC* cells.

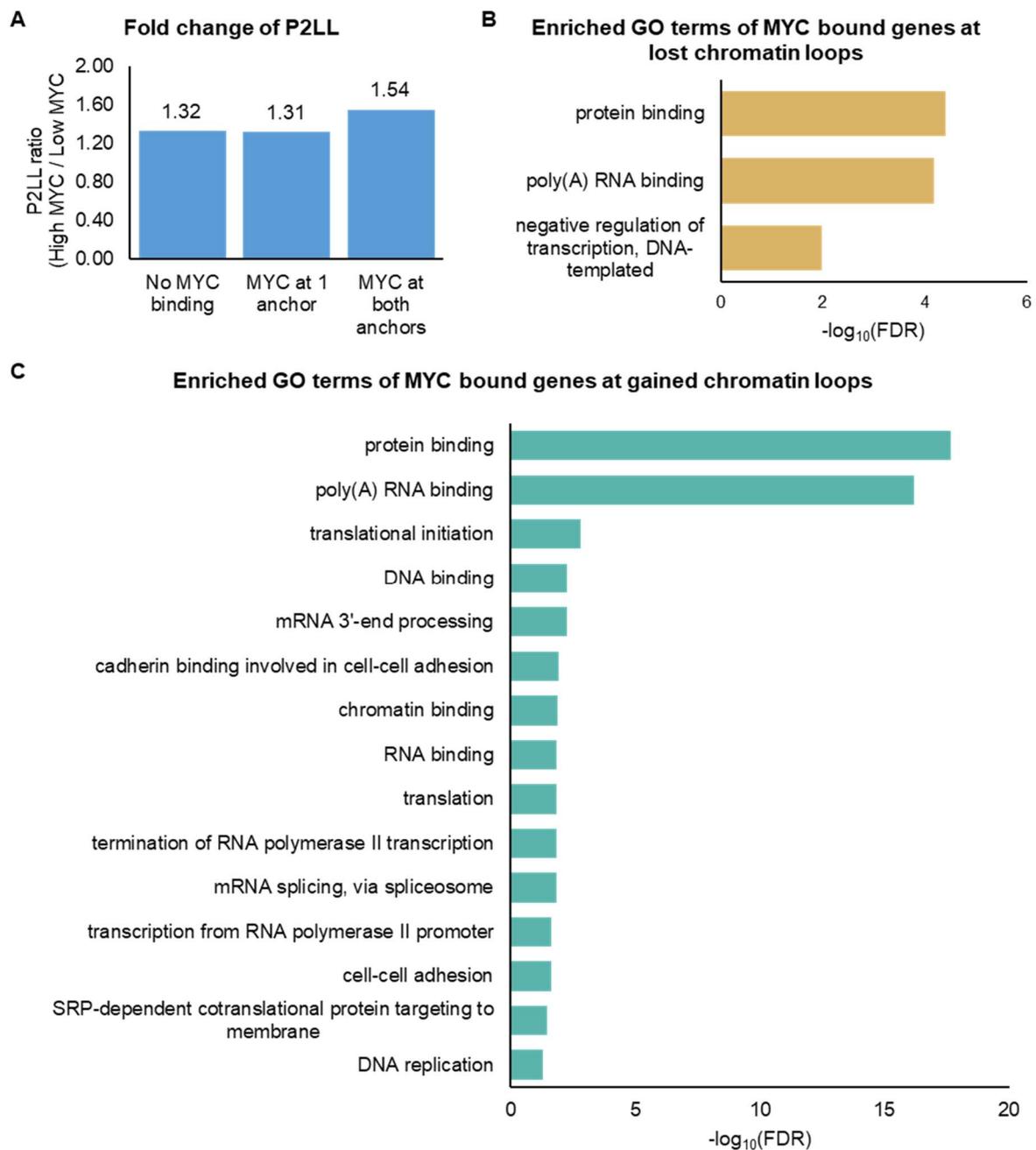


**Supplemental Figure S6 | Aggregate peak analysis (APA).** (A) Aggregate Peak Analysis (APA) plots at 5kb resolution showing the aggregate non-normalized signal of lost, common and gained chromatin loop sets in Low *MYC* and High *MYC* cells. Loop sets were filtered to remove short loops near the diagonal (shown above each APA plot). P: Peak signal at the centre pixel. (B) Fold change of non-normalized peak signal enrichment at lost, common, gained and all chromatin loop sets between High *MYC* and Low *MYC* cells. (C) Same APA analysis as in (A), but with SIQHiC normalization applied. (D) Fold change of SIQHiC normalized peak signal enrichment at lost, common and gained chromatin loop sets between High *MYC* and Low *MYC* cells.

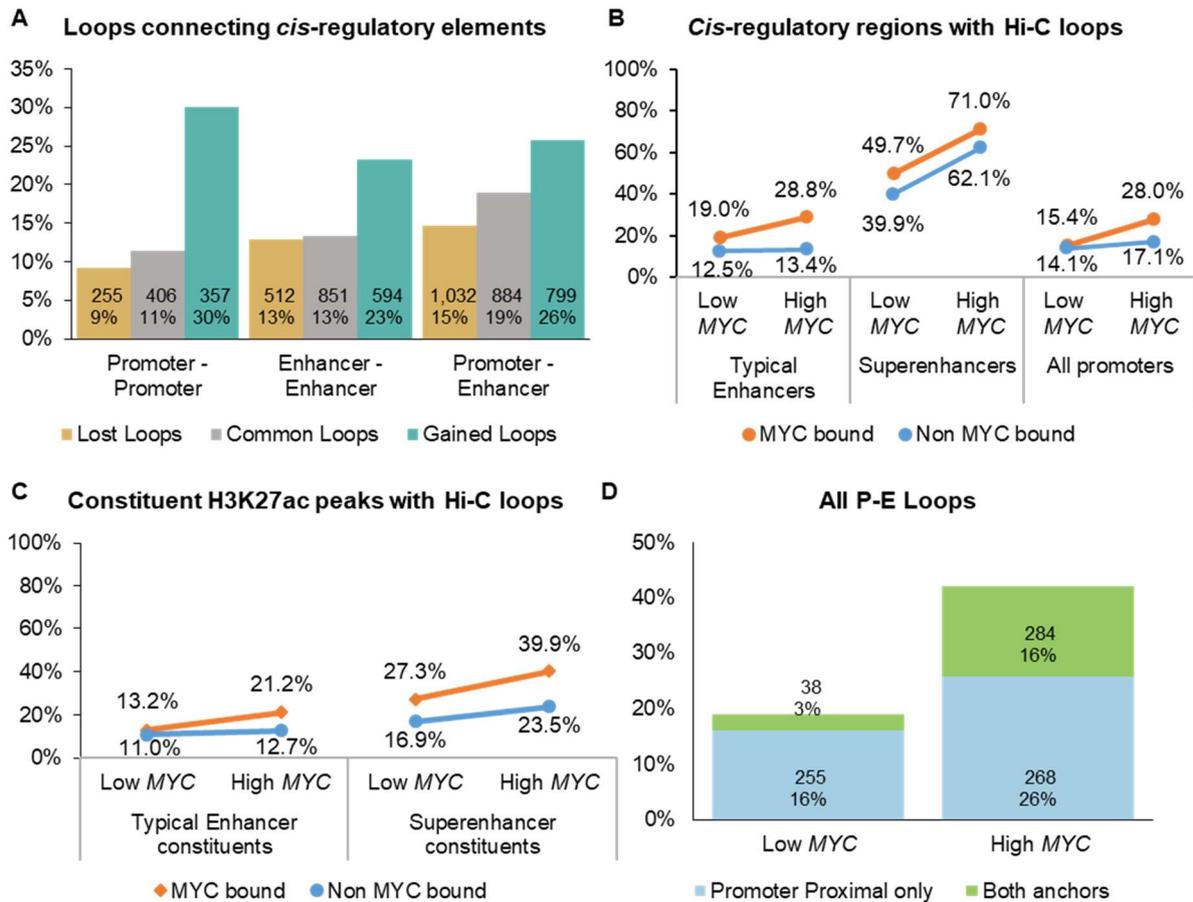


**Supplemental Figure S7 | *CTCF* siRNA knockdown reduced insulation at selected TAD boundaries and reduced chromatin contact frequencies at all chromatin loops. (A) *CTCF* (left) and *MYC* (right) gene expression in *MYC* overexpressed U2OS cells transfected with control (siControl) and *CTCF* siRNA (siCTCF), normalized against the *TBP* gene. Error bars show standard error across 2 biological replicates. (B) Overlap between TAD boundaries identified in siControl and siCTCF cells. (C) Overlap between chromatin loops identified in siControl and siCTCF cells. (D)**

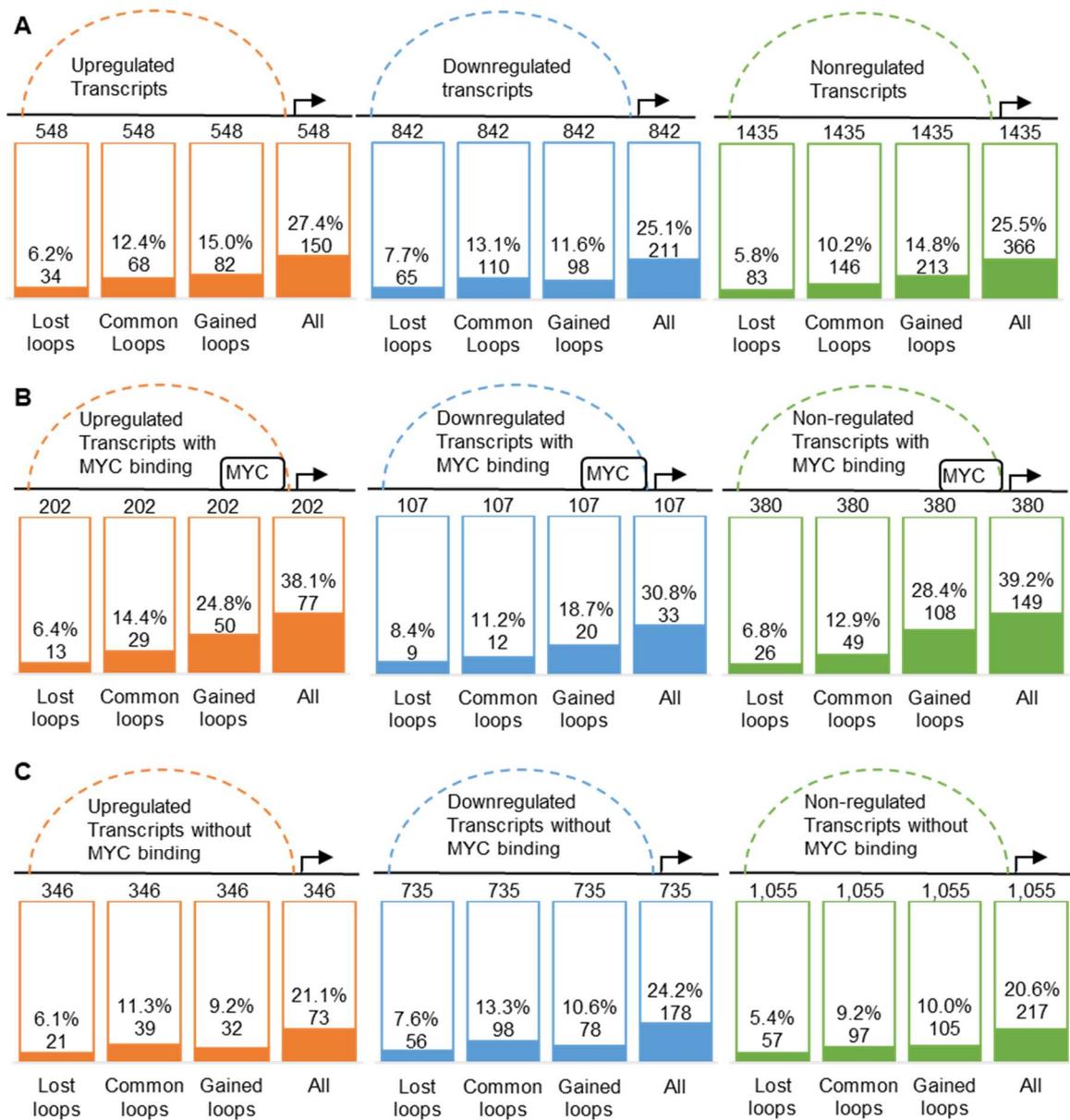
Topologically associated domain (TAD) separation scores of TAD boundaries that were lost, common and gained between siControl and siCTCF cells. **(E)** Aggregate Peak Analysis (APA) plots at 5kb resolution showing the aggregate non-normalized signal of lost, common and gained chromatin loop sets in siControl and siCTCF cells. Loop sets were filtered to remove short loops near the diagonal (shown above each APA plot). P: Peak signal at the centre pixel. **(F)** Fold change of non-normalized peak signal enrichment at lost, common, gained and all chromatin loop sets between siControl and siCTCF cells. **(G)** Same APA analysis as in **(E)**, but with SIQHiC normalization applied. **(H)** Fold change of SIQHiC normalized peak signal enrichment at lost, common and gained chromatin loop sets between siControl and siCTCF cells.



**Supplemental Figure S8 | MYC binding at chromatin loops.** (A) Fold change of P2LL signal between High *MYC* and Low *MYC* cells at chromatin loops with no MYC binding sites, loops with MYC binding sites at 1 loop anchor and loops with MYC at both loop anchors. P: peak enrichment signal. LL: Signal at lower left corner of the APA plot, representing local background signal. P2LL: Ratio of P to LL. (B-C) Gene ontology (GO) terms enriched in MYC bound genes at (B) lost and (C) gained chromatin loops.

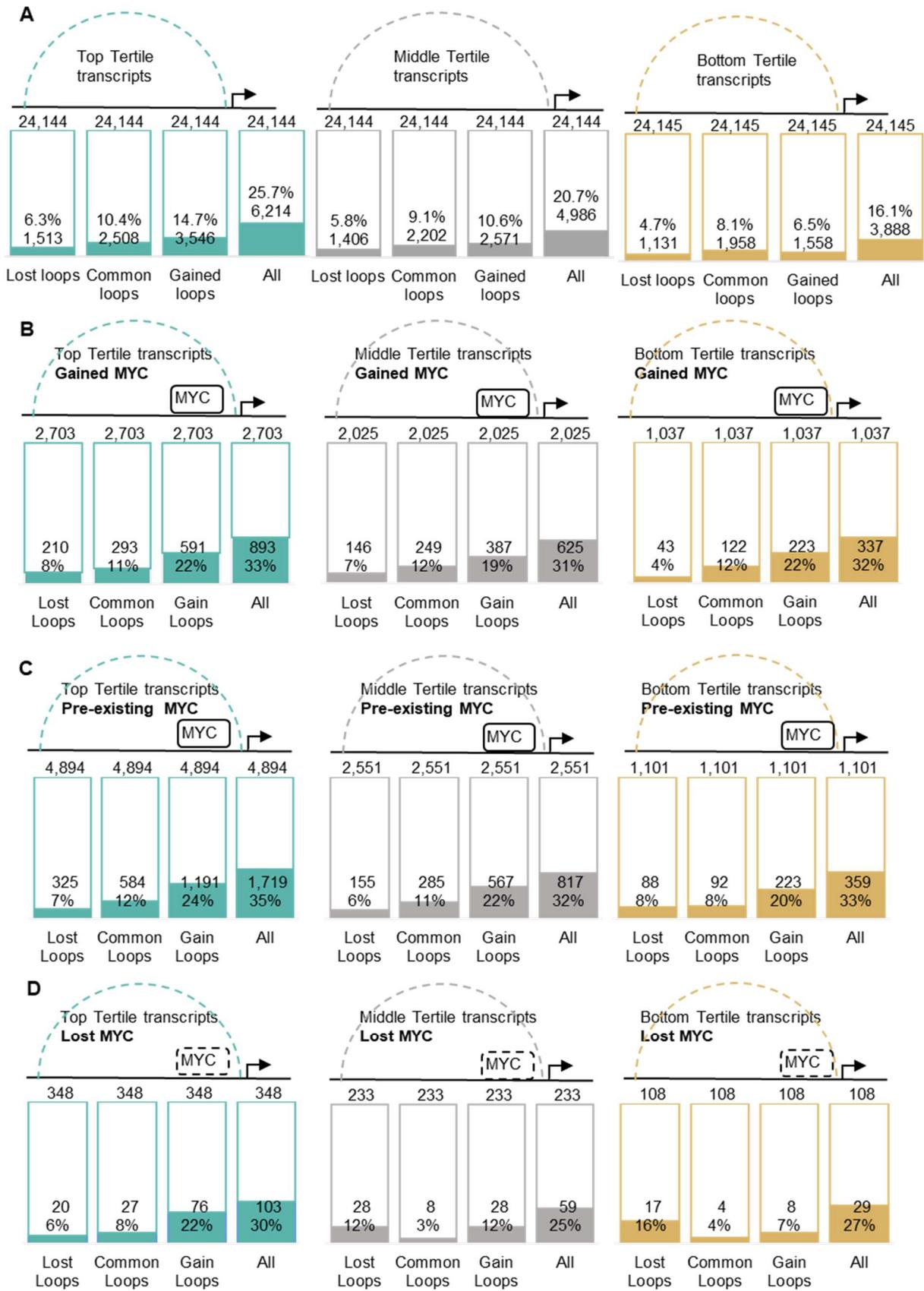


**Supplemental Figure S9 | MYC binding at promoter-enhancer chromatin loops.** (A) Percentage of promoter-promoter, enhancer-enhancer and promoter-enhancer loops within the lost, common and gained chromatin loop sets. (B) Percentage of MYC unbound (blue) and bound (orange) typical enhancers, superenhancers and promoters with Hi-C chromatin loops in Low *MYC* and High *MYC* cells. (C) Percentage of MYC unbound (blue) and bound (orange) typical enhancer and superenhancers H3K27ac constituent peaks. (D) Percentage of all promoter-enhancer chromatin loops bound by MYC at the promoter proximal loop anchor or at both anchors.



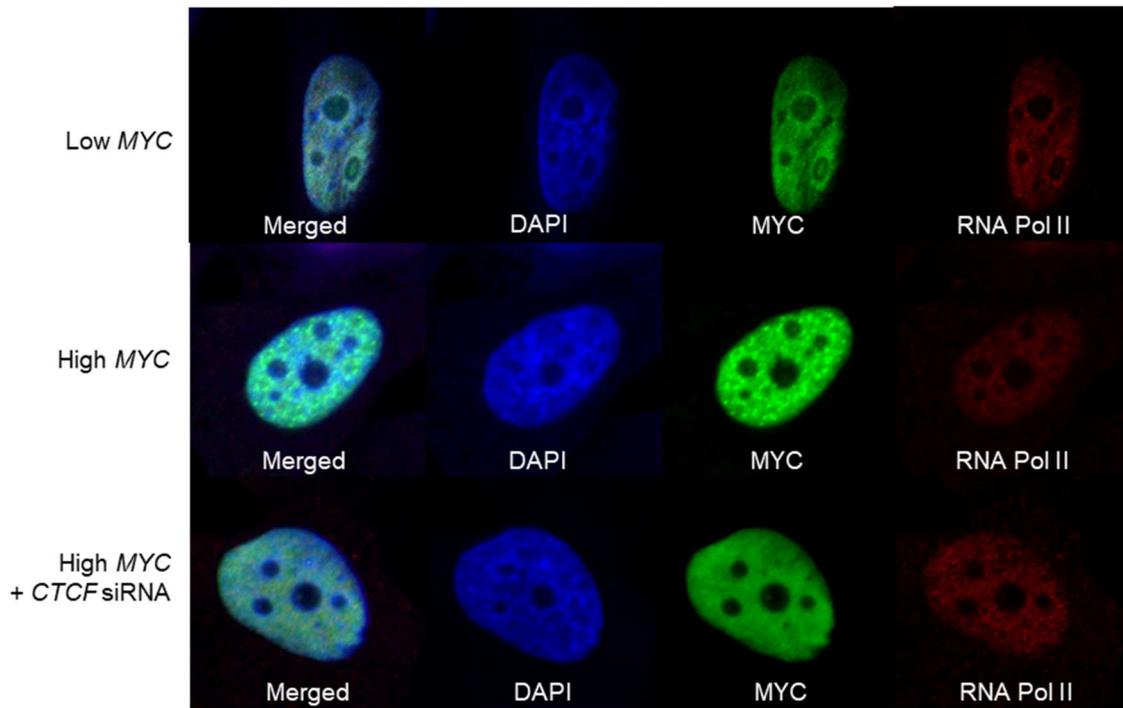
**Supplemental Figure S10 | Correlation between MYC regulated transcripts and chromatin**

**loops.** (A-C) Percentage of (A) all, (B) MYC bound and (C) non MYC bound transcripts with Hi-C chromatin loops. Transcripts were stratified into upregulated (orange), downregulated (blue) and non-regulated (green) transcripts.



**Supplemental Figure S11 | Correlation between transcript expression and chromatin loops.**

(A-D) Percentage of (A) all transcripts, (B) transcripts with gained MYC binding sites, (C) transcripts with pre-existing MYC binding sites and (D) transcripts with lost MYC binding sites with Hi-C chromatin loops. Transcripts were stratified into top (teal), middle (grey) and bottom (khaki) tertiles based on transcript expression after *MYC* overexpression.



**Supplemental Figure S12 | Immunofluorescence staining of MYC and RNA polymerase II.** From left to right, immunofluorescence images show merged signal, DAPI staining of DNA in blue, MYC staining in green and RNA polymerase II staining in red. RNA Pol II: RNA polymerase II.