

```

A
#!/usr/local/perl -w
@filenames=("H3K27ac_rep1","H3K27ac_rep2","H3K4me1_rep1","H3K4me1_rep2");
@chromo=("chr1","chr2","chr3","chr4","chr5","chr6","chr7","chr8","chr9","chr10","chr11","chr12","chr13","chr14","chr15","chr16","chr17","chr18","chr19","chrX","chrY");
foreach $sam (@filenames)
{
open(OUT.">$sam\_ChIP\_whole\_gene\_Jerry.txt")||die("Can't write OUT:$!\n");
my $total_reads=$overlap=$others=0;
foreach $chromosome (@chromo)
{
open(INA,"Mus_musculus.GRCm38.75.gtf.gene")||die("Can't open INA:$!\n");
open(INC,"$sam\_sam.$chromosome")||die("Can't open INC:$!\n");
while(<INA>)
{
chomp;
if(/^(w|w?)\t(w+)\tgene\t(d+)\t(d+)\t([+|-])\t(ENSMUSG\d+)\t([w\.\-|\(\)]+)$/ )
{
$chromosome_gene="chr$1";
if($chromosome_gene eq $chromosome)
{
$total_gene_id++; $TSS_id="$chromosome_gene:$3-$4:$6_$7:$2:$5"; $count{$TSS_id}=0;
for($i=$3;$i<=$4;$i++)
{
if(exists $gene{$i}){$gene{$i}.=";;";$TSS_id";} ## overlaped regions, more TSS IDs
else{$gene{$i}=$TSS_id;}}
} else{print "error1\t$_\n";}
}
}
while(<INC>)
{
chomp;
if(/^(w\.\-|\(\)]+)\t(d+)\t(chrw|w?)\t(d+)\t(d+)\t(d+)\tM\t/)
{
if($2 == 0){$str="plus"; $left=$4; $right=$4+250-1;} ## extend to 250 bp for each fragment
elsif($2 == 16){$str="minus"; $right=$4+$5-1; $left=$right-250+1;}
else{print "error Strand-FLAG: $_\n";}
$read_id=$1; $chr=$3; if($chr ne $chromosome){print "error wrong sam-chr: $_\n";}
$label=0; ##
for ($j=$left; $j<=$right; $j++)
{
if(exists $gene{$j}) ## read overlap with gene
{
$label=1;
if(exists $name{$read_id}) ## this read already overlap with gene
{
if($name{$read_id} =~ /$gene{$j}/){} ## already this gene ID
else{$name{$read_id}.=";;";$gene{$j}";} ## add this gene ID to this reads
}
else{$name{$read_id}=$gene{$j};} ## First intron overlap with this read
}
}
if($label==0){$non_overlap++;}
elsif($label==1) ## fragment mapped to gene
{
$overlap++; @arraylist=split(/;/, $name{$read_id}); foreach (@arraylist){$tmphash{$_}=0;} @newlist=keys %tmphash; foreach (@newlist){$count{$_}++;}
}
else{$others++;}
delete $name{$read_id}; foreach (keys %tmphash){delete $tmphash{$_};}
} else{print "error2\t$_\n";}
}
}
foreach (keys %count){print OUT "$_ \t$count{$_}\n"; $result++; delete $count{$_};} foreach (keys %gene){delete $gene{$_};}
close INA; close INC;close INA; close OUT;

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B
library("edgeR")
data <- read.table("Total_merge_Kdm5c_exonic_gene_Jerry.txt",sep="\t",header=T,row.names=1)
data_y <- DGEList(counts=data, group=c("WT","WT","WT","KO","KO","KO"))
keep <- rowSums(cpm(data_y)) >= 2
y <- data_y[keep,]
y$samples$lib.size <- colSums(y$counts)
y <- calcNormFactors(y)
y <- estimateCommonDisp(y,verbose=TRUE)
y <- estimateTagwiseDisp(y)
test_result <- exactTest(y, pair=c("WT","KO"))
diff_total <- data.frame(topTags(test_result, n=length(y$AveLogCPM)))
length_gene <- read.table("Gene_length.txt",header=T,row.names=1,sep="\t")
total <- cbind(diff_total,Length=length_gene[row.names(diff_total),])
write.table(total,file="results_edgeR_Kdm5c_WT_KO_exonic_gene.xls",quote=FALSE,sep="\t")

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Supplemental Figure S8. Scripts used for gene based counting and comparison. (A) Perl script used to count sequencing signals in gene bodies for ChIP-seq, ATAC-seq, GRO-seq, and RNA-seq data sets. Counting ChIP-seq signals of H3K27ac and H3K4me1 was used as an example. (B) R script used to compare gene expression changes. Comparison of RNAseq counts between WT and *Kdm5c* mutant was used as an example.