

	10	20	30	40	50	60	70	80	90
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DhhSerCAG	GATACGATGGCCGAGT	GGTT	AAGGCGAAGGATGCAGGTTCCCTTTGGGC	---	TCTGCCCG	CGCAGGTT	CGAACCCCTGCTCGTGTCC	82	
CaSerCAG	GATACGATGGCCGAGT	GGTT	AAGGCGAAGGATGCAGGTTCCCTTTGGGC	---	ATTGCCCG	CGCAGGTT	CGAACCCCTGCTCGTGTCC	82	
CadSerCAG	GATACGATGGCCGAGT	GGTT	AAGGCGAAGGATGCAGGTTCCCTTTGGGC	---	ATTGCCCG	CGCAGGTT	CGAACCCCTGCTCGTGTCC	82	
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CllSerCAG	GATGTTGGTGGCCGAGT	GGTT	AAGGCGTGAAGGATGCAGGTTCCCTTTGGGC	---	TCTGCTCT	CGCAGGTT	CGAACCCCTGCTCGTGTCC	82	
MefSerCAG	GATGTTGGTGGCCGAGT	GGTT	AAGGCGTGAAGGATGCAGGTTCCCTTTGGGC	---	TCTGCTCT	CGCAGGTT	CGAACCCCTGCTCGTGTCC	82	
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CnmSerCAGa	GATACGATGGCCGAGT	GGTT	AAGGCGAAGGATGCAGGTTCCCTTTGGGC	---	ATTGCCCG	CGCAGGTT	CGAACCCCTGCTCGTGTCC	82	
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Ct_aSerUGAa	GGCCGATGTCCGAGT	GGTT	AAGGAGAAAGACTTGAATCTTTTGGGC	---	TCTGCCCG	CGCAGGTT	CGAGTCTCGTGTCC	82	
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YlSerCGAa	GGCAACTTGTCCGAGT	GGTT	AAGGAGAAAGACTTGAATCTTTTGGGC	---	TTTTCCCG	CGCAGGTT	CGAACCCCTGCTCGTGTCC	82	
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WaLeuCAGa	GGGAGTGTGGCCGAGT	GGTT	AAGGCGCCCGTACAGGTTGCAAGTCTCTT	---	TTAAGAGG	CGCGAGTT	CGAACCCCTGCTACCTTCCA	83	
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KmmLeuCAG	GGGTGTGTGGCCGAGT	GGTT	AAGGCGCTGCTTACAGGTTGCAAGTCTGTTAAAGTTAACAGG	---	CGCGAGTT	CGAACCCCTGCGCCGATCCA	87		
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SpLeuCAG	GGCGAAGTGGCCGAGT	GGTCT	ATGGCCGTAGCTTACAGGTTGCTAGTCTACG	---	TATGTGAG	CGTGGGTT	CGAACCCCTGCTCGTCA	84	
CaLeuAAGa	GACTGGATGGCCGAGT	GGTC	AAGGCGCCAGGTTAAGGTTCTGGTCTACG	---	AAAGTGGG	CGCGAGTT	CGAACCCCTGCTCGTCA	83	
PtaLeuUAGa	GAGGTTTGGCCGAGT	GGTT	AAGGCGCATGTTAAGGCTGTTATCA	---	TTGCGATG	CGCGGTT	CGAACCCCTGCTCGTCA	82	
PtaLeuAAGa	GGCAACTTGGCCGAGT	GGTCT	AAGGCGCCAGGTTAAGGTTCTGGTCTC	---	TTCCGAGG	CGTGGGTT	CGAACCCCTGCTCGTCA	82	
PiuLeuAAGa	GGCAGCTTGGCCGAGT	GGTCT	AAGGCGTACAGGTTAAGGTTCTGGTCTC	---	TTCCGAGG	CGTGGGTT	CGAACCCCTGCTCGTCA	82	
KaaLeuAAGa	GGTACTCTGGCCGAGT	GGTCT	AAGGCGTACAGGTTAAGGTTCTGGTCTC	---	TACGGAGG	CGCGGGTT	CGAACCCCTGCTCGTCA	82	
YlLeuAAGa	GGCAACTTGGCCGAGT	GGTCT	AAGGCGCCAGGTTAAGGTTCTGGTCTC	---	TTCCGAGG	CGAGAGTT	CGAACCCCTGCTCGTCA	82	
LoeLeuAAGa	GGATGGATGGCCGAGT	GGTTT	AAGGCGTACAGGTTAAGGTTCTGATCTC	---	TTCCGAGG	CGCGAGTT	CGAACCCCTGCTCGTCA	83	
MrgLeuAAGa	GGTACTCTGGCCGAGT	GGTCT	AAGGCGCCAGGTTAAGGTTCTGGTCTC	---	TTCCGAGG	CGCGAGTT	CGAACCCCTGCTCGTCA	83	
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CglLeuGAG	GGTACTCTGGCCGAGT	GGTCT	AAGGCGCCAGGTTAAGGTTCTGGTCTC	---	TTCCGAGG	CGCGGGTT	CGAACCCCTGCTCGTCA	82	
KlLeuGAG	GGCACTATGGCCGAGT	GGTTT	AAGGCGCCAGGTTACAGGTTCTGGTATC	---	TCCGGATG	CGCGAGTT	CGAACCCCTGCTCGTCA	82	
ErgLeuGAG	GGTACTCTGGCCGAGT	GGTT	AAGGCGCCAGGTTACAGGTTCTGGTCTC	---	TTCCGAGG	CGCGAGTT	CGAACCCCTGCTCGTCA	81	
YlLeuUAGa	GGGACTTGGCCGAGC	GGTCT	AAGGCGCCAGGTTAAGGCTGTTATC	---	GCAAGATG	CGCGAGTT	CGAACCCCTGCTCGTCA	82	
LatLeuUAGc	GGGAGTTTGGCCGAGT	GGTT	AAGGCGTACAGTTAAGGTTCTGATATC	---	TGCGGATG	CGCGGGTT	CGAACCCCTGCTCGTCA	81	
PiuLeuUAGa	GTTGGGTTTGGCCGAGT	GGTCT	AAGGCGTACAGTTAAGGCTGTTATC	---	TTCCGATG	CAAGAGTT	CGAACCCCTTACGCCACA	82	
KaaLeuUAGa	GGGAGTTTGGCCGAGT	GGTTT	AAGGCGTACAGTTAAGGTTCTGATATC	---	TTCCGATG	CGTGGGTT	CGAACCCCTGCTCGTCA	82	
PiuLeuUAGb	GGCAGTTTGGCCGAGT	GGTCT	AAGGCGTACAGTTAAGGCTGTTATC	---	TTCCGAGG	CGTGGGTT	CGAACCCCTGCTCGTCA	82	
ErgLeuUAGa	GAGAGTTTGGCCGAGT	GGTT	AAGGCGTACAGTTAAGGCTGTTATC	---	TACGGATG	CGCGGGTT	CGAACCCCTGCTCGTCA	81	

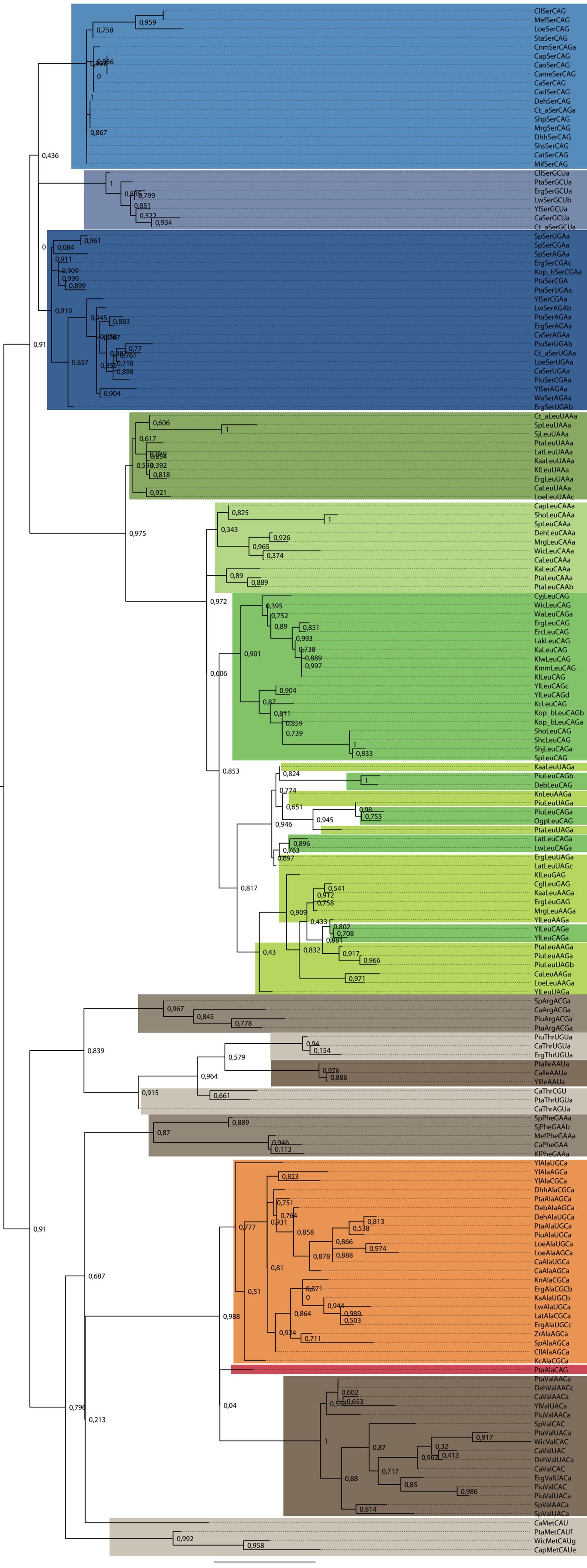


Fig. S2: Phylogenetic tree computed on the alignment of 172 tRNA sequences (Fig. S1) using the Maximum-Likelihood approach as implemented in FastTree v. 2.1.7 with 1000 replicates. Support values are shown as relative numbers. Branches are colored according to Fig. 1B. The scale bar denotes substitutions per site.

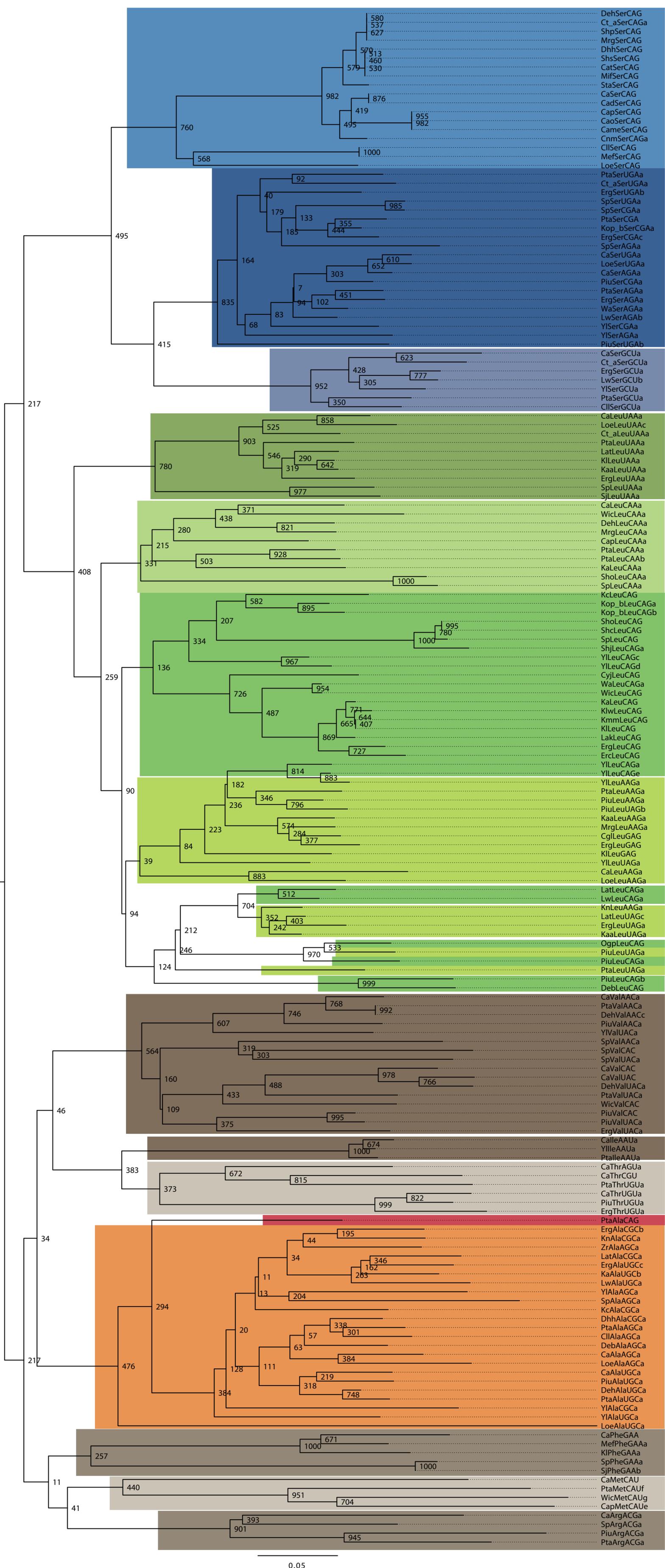


Fig. S3: Phylogenetic tree computed on the alignment of 172 tRNA sequences (Fig. S1) using the Neighbour-Joining approach as implemented in ClustalW v. 2.1 with 1000 replicates. Support values are shown as absolute numbers. Branches are colored according to Fig. 1B. The scale bar denotes substitutions per site.

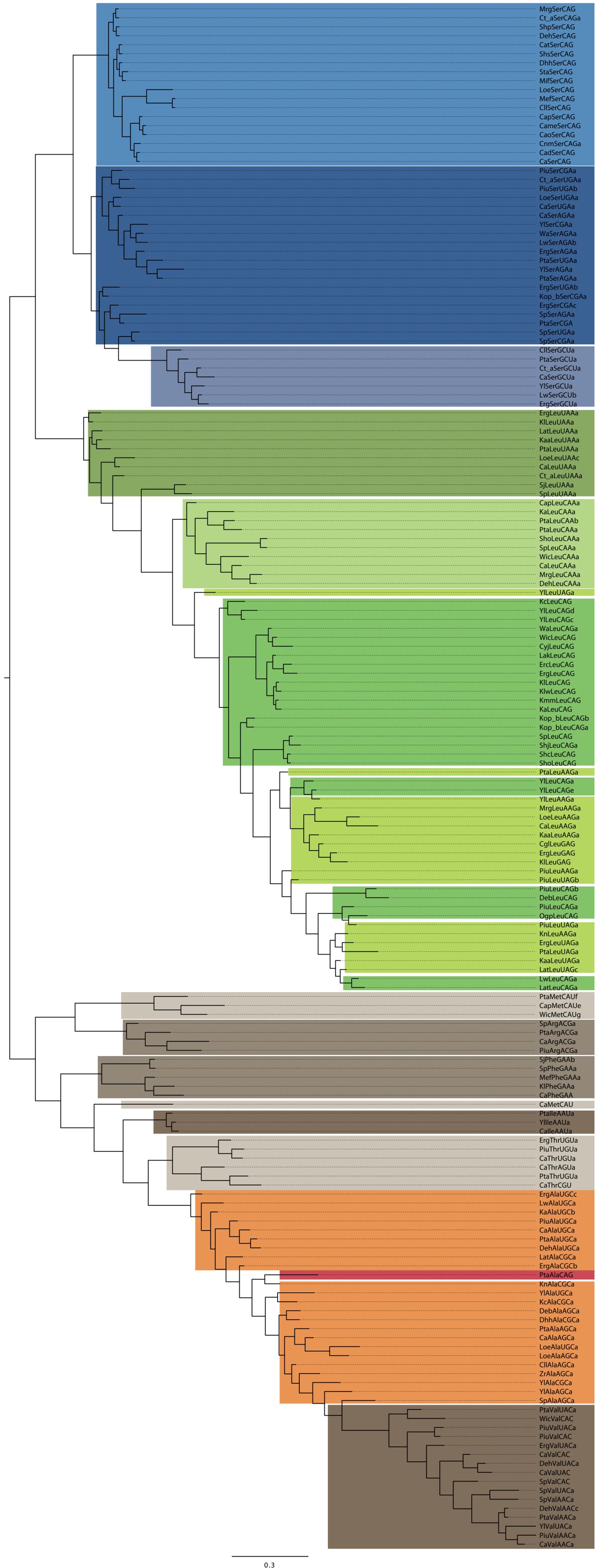


Fig. S4: Phylogenetic tree computed on the alignment of 172 tRNA sequences (Fig. S1) using the Bayesian approach as implemented in Phase v. 2 with 1,000,000 iterations. Branches are colored according to Fig. 1B. The scale bar denotes substitutions per site.

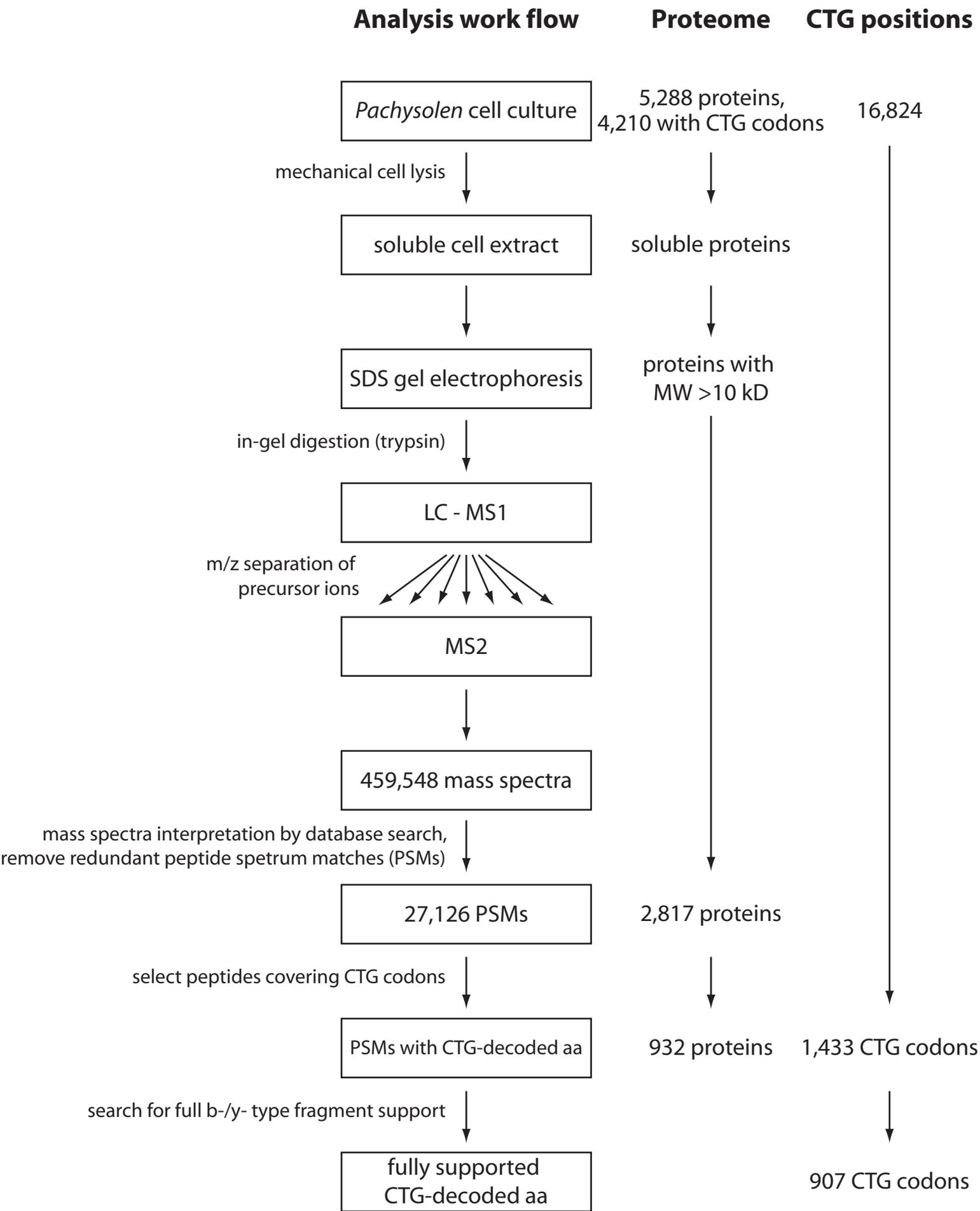


Fig. S5: Summary of the proteome analysis workflow. For the database search we generated a *Pachysolen* gene prediction dataset, in which all codons were iteratively translated into all amino acids (except isoleucine and leucine, which have identical molecular weights). Therefore, the database consists of 1,216 (64 x 19) times 5,288 proteins. The peptides with differently translated codons distinguish in their total MW, accordingly have different precursor ion masses and result in different spectra.

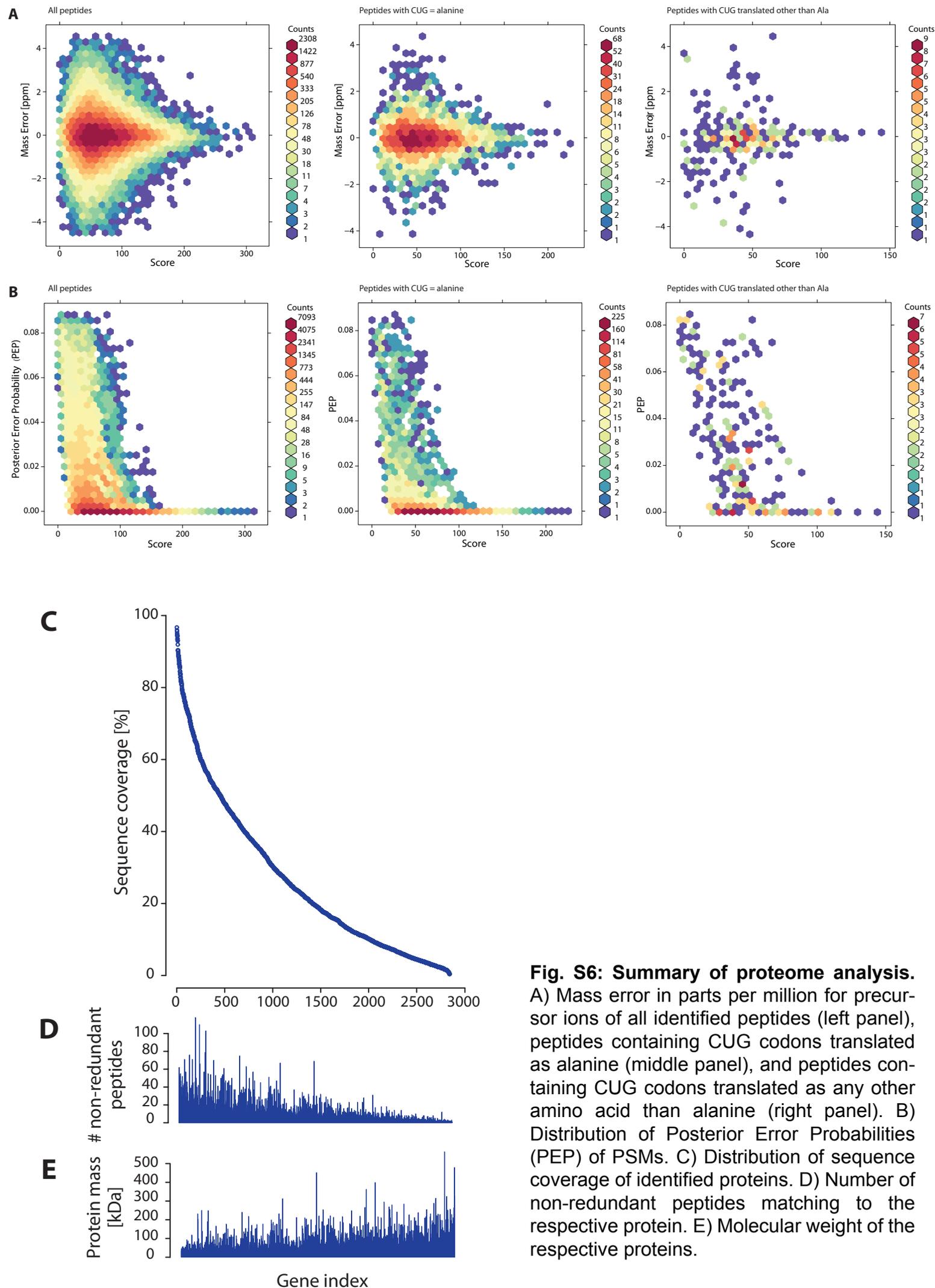


Fig. S6: Summary of proteome analysis. A) Mass error in parts per million for precursor ions of all identified peptides (left panel), peptides containing CUG codons translated as alanine (middle panel), and peptides containing CUG codons translated as any other amino acid than alanine (right panel). B) Distribution of Posterior Error Probabilities (PEP) of PSMs. C) Distribution of sequence coverage of identified proteins. D) Number of non-redundant peptides matching to the respective protein. E) Molecular weight of the respective proteins.

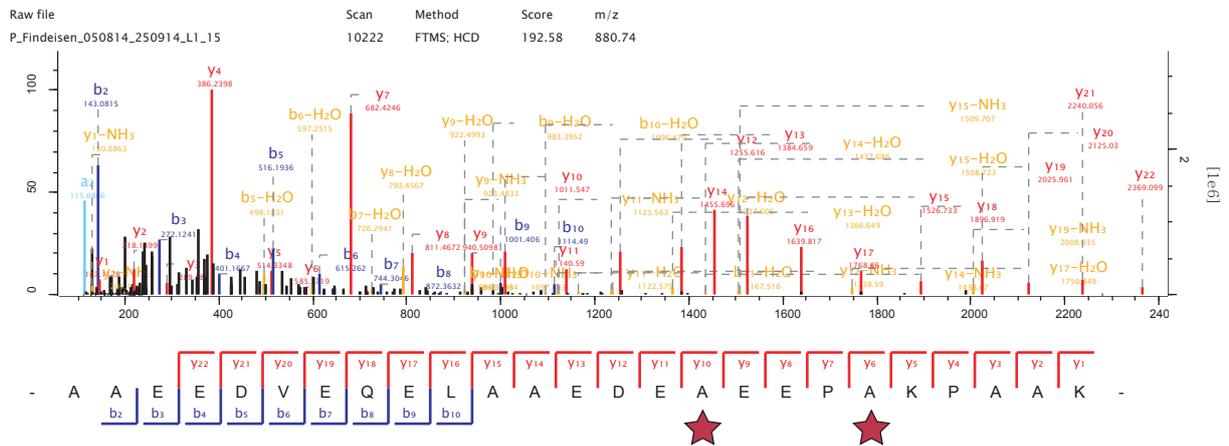
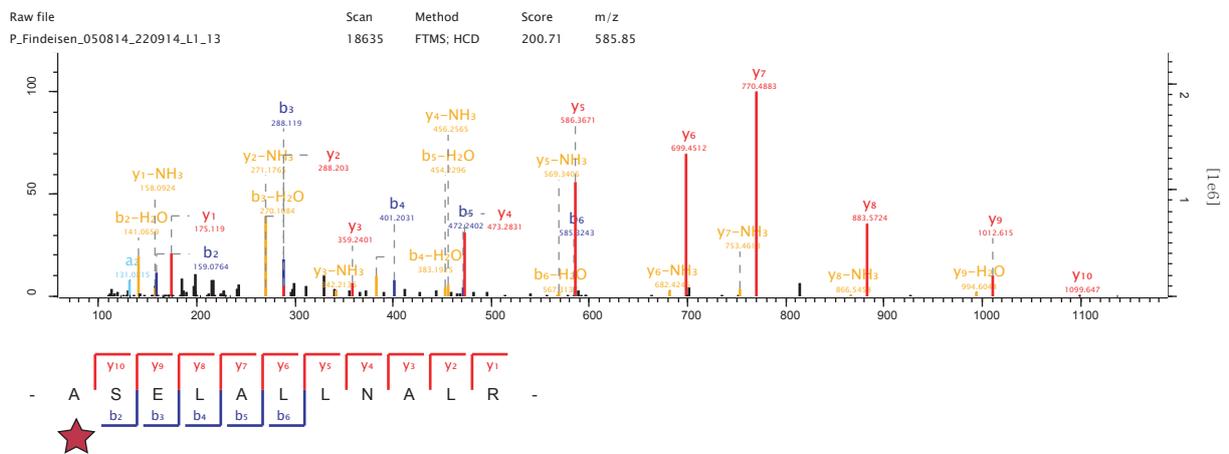
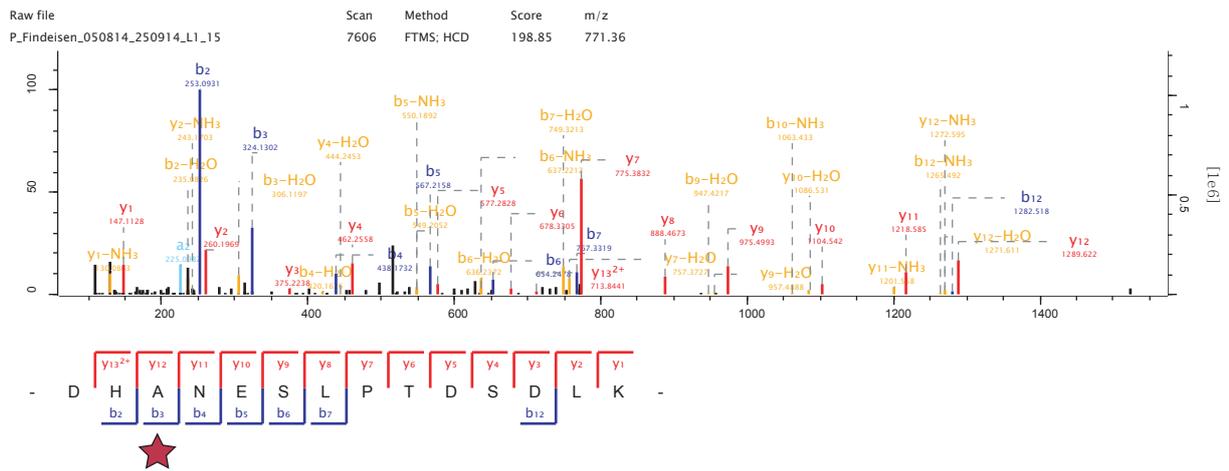


Fig. S7: Representative LC-MS/MS spectra featuring CUG codons translated as alanine (marked with stars).

	# peptide spectrum matches (PSMs) covering AUG positions	# AUG positions covered by PSMs	# AUG positions with full b-/y-type fragment ion support	# fully supported AUG positions with ambiguous translation
Trp	6	6	-	-
Tyr	11	9	3	2
Phe	4	3	-	-
Met	9234	2799	1860	5
Ile/Leu	38	13	7	1
Val	23	17	2	-
Ala	5	5	-	-
Ser	12	8	1	-
Thr	18	9	1	-
Cys	4	4	-	-
Asn	8	7	1	-
Gln	13	4	1	-
Asp	20	16	4	1
Glu	19	12	4	1
His	13	8	1	-
Lys	9	9	-	-
Arg	14	11	-	-
Gly	53	29	3	-
Pro	14	10	1	-

Fig. S8: Decoding of the unambiguously decoded AUG codon. Number and distribution of observed AUG translations in all peptide spectrum matches from MS/MS analysis.

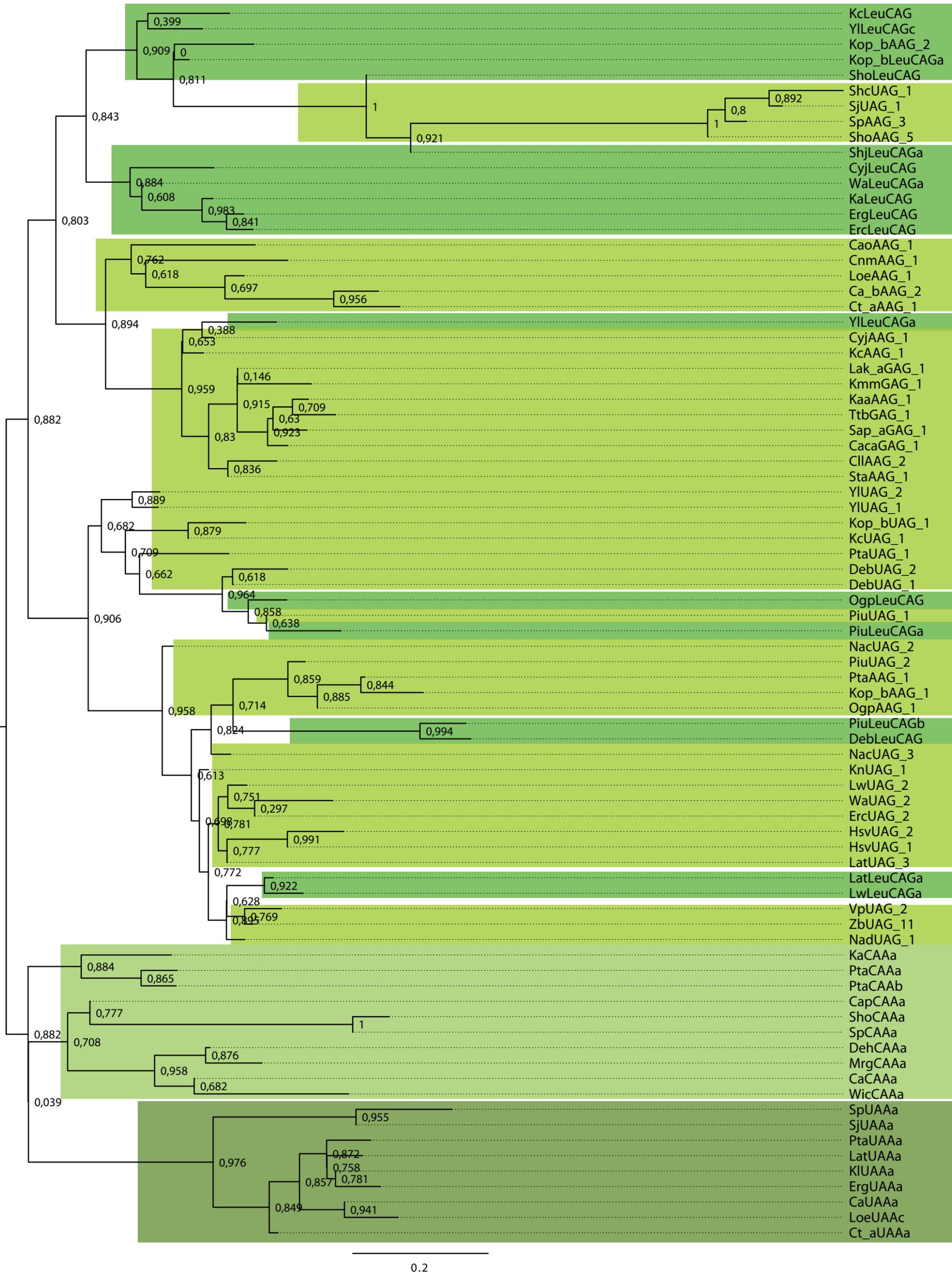


Fig. S9: Phylogenetic tree computed on the alignment of 80 representative Leu-tRNA sequences using the Maximum-Likelihood approach as implemented in FastTree v. 2.1.7 with 1000 replicates. Support values are shown as relative numbers. Branches are colored according to Fig. 1B. The scale bar denotes substitutions per site.

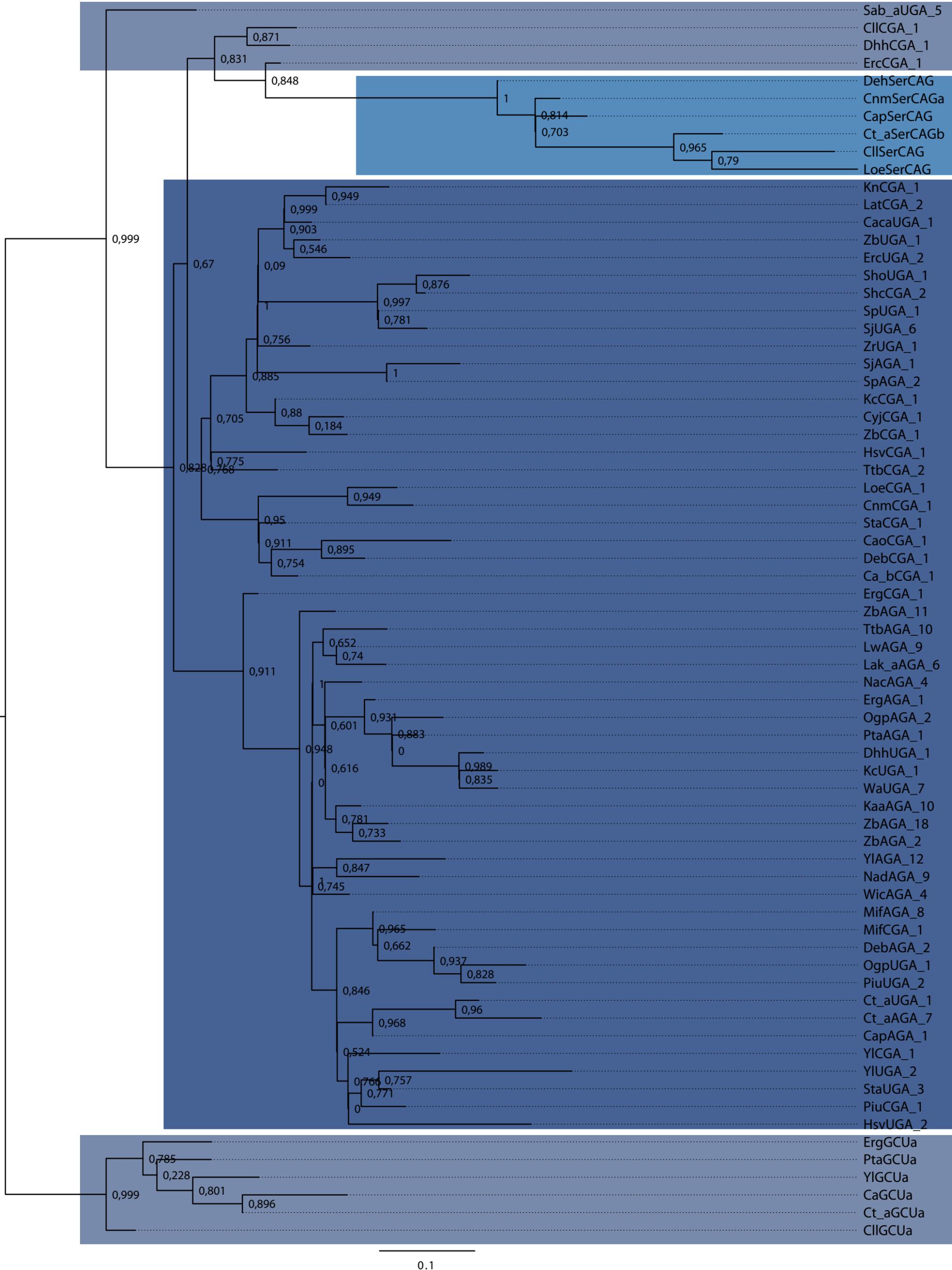


Fig. S10: Phylogenetic tree computed on the alignment of 70 representative Ser-tRNA sequences using the Maximum-Likelihood approach as implemented in FastTree v. 2.1.7 with 1000 replicates. Support values are shown as relative numbers. Branches are colored according to Fig. 1B. The scale bar denotes substitutions per site.

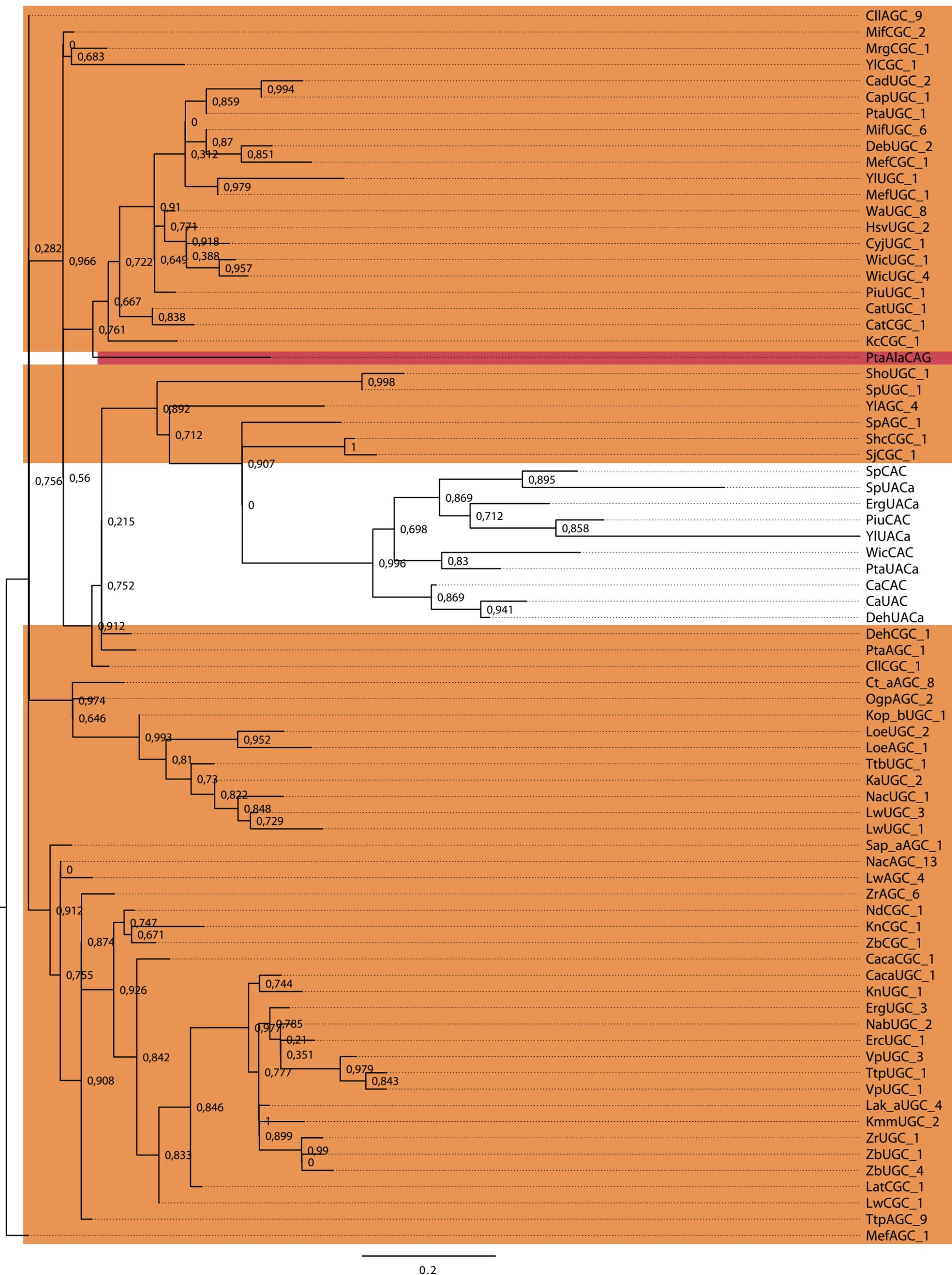
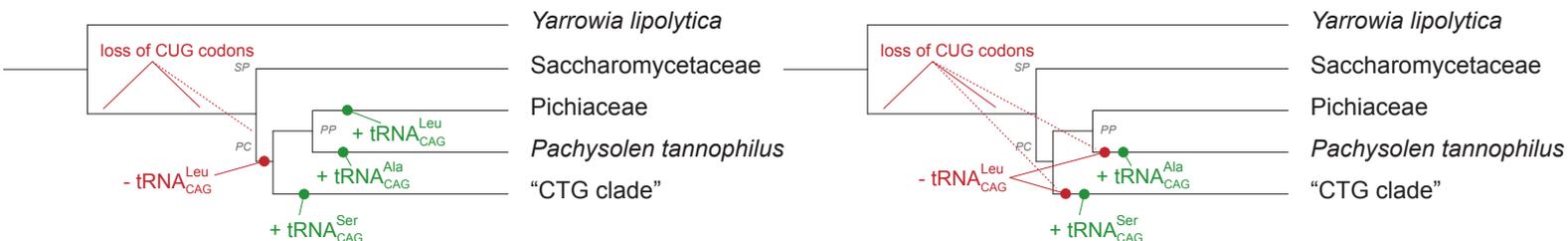
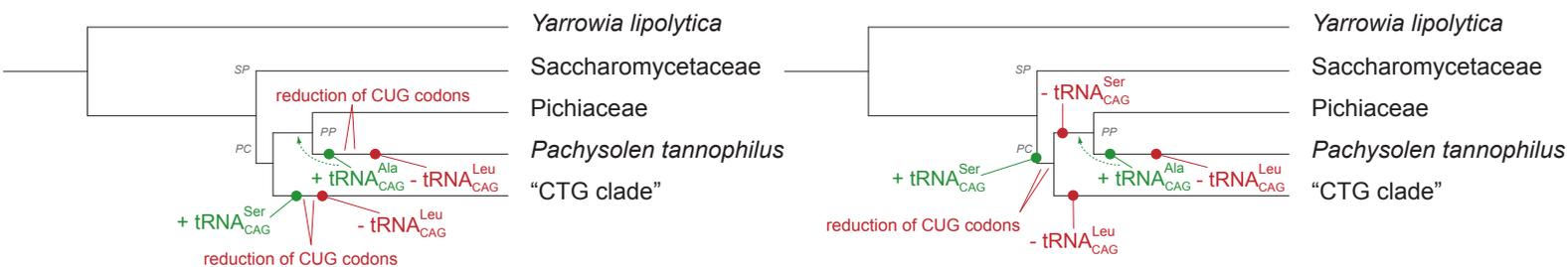


Fig. S11: Phylogenetic tree computed on the alignment of 76 representative Ala-tRNA sequences, with Val-tRNAs as outgroup, using the Maximum-Likelihood approach as implemented in FastTree v. 2.1.7 with 1000 replicates. Support values are shown as relative numbers. Branches are colored according to Fig. 1B. The scale bar denotes substitutions per site.

Codon capture



Ambiguous intermediate



tRNA loss driven codon reassignment

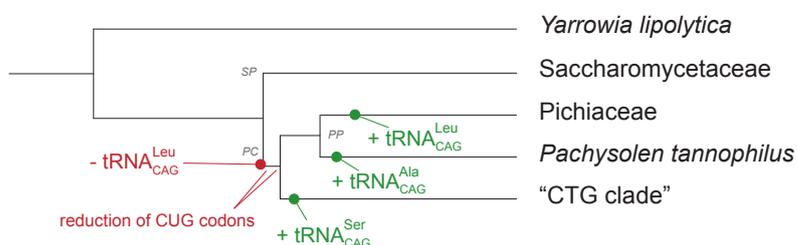


Fig. S12: The mechanism of CUG codon reassignment. The scheme contrasts the presence of $tRNA_{CAG}$ s and evolution of CUG codons according to the *tRNA loss driven codon reassignment* hypothesis with assumptions based on the *codon capture* and *ambiguous intermediate* theories. The schemes present alternative but less likely scenarios compared to the most probable sequence of events as shown for each hypothesis in Fig. 4. Mixed models seem extremely unlikely and are not shown.

Table S1: Summary of the leucine, serine, and alanine tRNA data.

Species	Abbreviation	GenBank accession numbers	Reference	TTA	TTG	CTT	CTC	CTA	CTG	TCT	TCC	TCA	TCG	AGT	AGC	GCT	GCC	GCA	GCG
<i>Candida albicans</i>	Ca_b	CH672346, CH672349-CH672350, CH672354, CM000309-CM000313, GG670278, GG670282	Butler G, Rasmussen MD, Lin MF et. al. (2009) Evolution of pathogenicity and sexual reproduction in eight <i>Candida</i> genomes. <i>Nature</i> 459, 657-662.	5	6	2	0	0	1	4	0	3	1	0	2	6	0	2	0
<i>Candida bracarensis</i>	Cnb	CAPU01000001-CAPU01000251	Gabalton T, Martin T, Marcet-Houben M et. al. (2013) Comparative genomics of emerging pathogens in the <i>Candida glabrata</i> clade. <i>BMC Genomics</i> 14, 623.	4	9	0	1	4	0	9	0	3	1	0	3	11	0	6	0
<i>Candida castelii</i>	Caca	CAPW01000001-CAPW01000101	Gabalton T, Martin T, Marcet-Houben M et. al. (2013) Comparative genomics of emerging pathogens in the <i>Candida glabrata</i> clade. <i>BMC Genomics</i> 14, 623.	2	5	0	1	2	0	5	0	2	1	0	2	5	0	3	1
<i>Candida dubliniensis</i>	Cad	FM992688-FM992695	Jackson AP, Gamble JA, Yeomans T et. al. (2009) Comparative genomics of the fungal pathogens <i>Candida dubliniensis</i> and <i>Candida albicans</i> . <i>Genome Res</i> 19, 2231-2244.	5	5	2	0	0	1	4	0	3	1	0	1	6	0	2	0
<i>Candida glabrata</i>	Cgl	CR380947-CR380959	Dujon B, Sherman D, Fischer G et. al. (2004) Genome evolution in yeasts. <i>Nature</i> 430, 35-44.	3	8	0	1	4	0	9	0	2	1	0	3	10	0	5	0
<i>Candida maltosa</i>	Cnm	AOGT01000001-AOGT01002947	http://www.ncbi.nlm.nih.gov/nuccore/AOGT00000000.1	1	0	1	0	0	1	0	0	2	1	0	0	0	0	1	0
<i>Candida metapsilosis</i>	Came	CBZN020000001-CBZN020000096	http://www.ncbi.nlm.nih.gov/nuccore/CBZN00000000.2	3	3	1	0	0	1	2	0	3	1	0	1	3	0	3	0
<i>Candida nivariensis</i>	Cdn	CAPV01000001-CAPV01000123	Gabalton T, Martin T, Marcet-Houben M et. al. (2013) Comparative genomics of emerging pathogens in the <i>Candida glabrata</i> clade. <i>BMC Genomics</i> 14, 623.	4	10	0	1	4	0	10	0	3	1	0	3	12	0	6	0
<i>Candida orthopsilosis</i>	Cao	HE681719-HE681726	Riccombeni A, Vidanes G, Proux-Wera E et. al. (2012) Sequence and analysis of the genome of the pathogenic yeast <i>Candida orthopsilosis</i> . <i>PLoS One</i> 7, e35750.	3	1	1	0	0	1	2	0	2	1	0	1	3	0	2	0
<i>Candida parapsilosis</i>	Cap	CABE01000001-CABE01000024	Butler G, Rasmussen MD, Lin MF et. al. (2009) Evolution of pathogenicity and sexual reproduction in eight <i>Candida</i> genomes. <i>Nature</i> 459, 657-662.	3	3	1	0	0	1	2	0	3	1	0	1	3	0	3	0
<i>Candida tenuis</i>	Cat	GL996507-GL996531	Wohlbach DJ, Kuo A, Sato TK et. al. (2011) Comparative genomics of xylose-fermenting fungi for enhanced biofuel production. <i>Proc Natl Acad Sci U S A</i> 108, 13212-13217.	2	5	1	0	0	1	4	0	2	1	0	3	4	0	2	1
<i>Candida tropicalis</i>	Ct_a	GG692395-GG692418	Butler G, Rasmussen MD, Lin MF et. al. (2009) Evolution of pathogenicity and sexual reproduction in eight <i>Candida</i> genomes. <i>Nature</i> 459, 657-662.	6	6	2	0	0	1	7	0	4	1	0	3	9	0	5	0
<i>Clavispora lusitaniae</i>	ClI	CH408076-CH408083, GG670277	Butler G, Rasmussen MD, Lin MF et. al. (2009) Evolution of pathogenicity and sexual reproduction in eight <i>Candida</i> genomes. <i>Nature</i> 459, 657-662.	2	11	3	0	0	1	7	0	1	2	0	3	9	0	3	1
<i>Cyberlindnera jadinii</i>	Cyj	DG000065-DG000077	Tomita Y, Ikeo K, Tamakawa H et. al. (2012) Genome and Transcriptome Analysis of the Food-Yeast <i>Candida utilis</i> . <i>PLoS One</i> 7, e37226.	1	7	3	0	1	1	6	0	2	1	0	2	6	0	2	0

Species	Abbreviation	GenBank accession numbers	Reference	TTA	TTG	CTT	CTC	CTA	CTG	TCT	TCC	TCA	TCG	AGT	AGC	GCT	GCC	GCA	GCG
Debaryomyces hansenii	Deh	CR382133-CR382139	Dujon B, Sherman D, Fischer G et. al. (2004) Genome evolution in yeasts. Nature 430, 35-44.	9	4	2	0	0	1	6	0	3	1	0	3	7	0	4	1
Debaryomyces hansenii var hansenii	Dhh	AHBE01000001-AHBE01000541	Kumar S, Randhawa A, Ganesan K et. al. (2012) Draft Genome Sequence of Salt-Tolerant Yeast Debaryomyces hansenii var. hansenii MTCC 234. Eukaryot Cell 11, 961-962.	1	0	2	0	0	1	0	0	2	1	0	2	0	0	0	1
Dekkera bruxellensis	Deb	KB454452-KB454483	Piskur J, Ling Z, Marcet-Houben M et. al. (2012) The genome of wine yeast Dekkera bruxellensis provides a tool to explore its food-related properties. Int J Food Microbiol 157, 202-209.	1	2	2	0	2	1	2	0	0	1	0	2	3	0	3	0
Eremothecium cymbalariae	Erc	CP002497-CP002504	Wendland J, Walther A (2011) Genome evolution in the eremothecium clade of the Saccharomyces complex revealed by comparative genomics. G3 (Bethesda) 1, 539-548.	2	8	0	1	2	1	5	0	2	1	0	3	5	0	3	1
Eremothecium gossypii	Erg	AE016814-AE016820	Dietrich FS, Voegeli S, Brachat S et. al. (2004) The Ashbya gossypii genome as a tool for mapping the ancient Saccharomyces cerevisiae genome. Science 304, 304-7.	2	7	0	1	6	1	7	0	2	3	0	3	7	0	4	2
Hanseniaspora vineae	Hsv	JFAV02000001-JFAV02000305	Giorello FM, Berna L, Greif G et. al. (2014) Genome Sequence of the Native Apiculate Wine Yeast Hanseniaspora vineae T02/19AF. Genome Announc 2, e00530-14.	5	9	0	0	3	0	7	0	3	1	0	3	7	0	5	0
Kazachastania naganishii	Kn	HE978314-HE978326	Gordon JL, Armisen D, Proux-Wera E et. al. (2011) Evolutionary erosion of yeast sex chromosomes by mating-type switching accidents. Proc Natl Acad Sci U S A 108, 20024-20029.	3	5	1	0	3	0	7	0	1	1	0	3	7	0	1	1
Kazachstania africana	Kaa	HE650821-HE650832	Gordon JL, Armisen D, Proux-Wera E et. al. (2011) Evolutionary erosion of yeast sex chromosomes by mating-type switching accidents. Proc Natl Acad Sci U S A 108, 20024-20029.	14	2	2	0	2	0	11	0	4	1	0	4	12	0	5	0
Kluyveromyces aestuarii	Ka	AEAS01000001-AEAS01000336	Baker CR, Tuch BB, Johnson AD (2011) Extensive DNA-binding specificity divergence of a conserved transcription regulator. Proc Natl Acad Sci U S A 108, 7493-7498.	3	7	0	1	2	1	6	0	3	1	0	2	7	0	5	0
Kluyveromyces lactis	Kl	CR382121-CR382126	Dujon B, Sherman D, Fischer G et. al. (2004) Genome evolution in yeasts. Nature 430, 35-44.	3	7	0	1	2	1	6	0	2	1	0	2	7	0	3	0
Kluyveromyces marxianus var marxianus	Kmm	JH924896-JH924903	Jeong H, Lee DH, Kim SH et. al. (2012) Genome Sequence of the Thermotolerant Yeast Kluyveromyces marxianus var. marxianus KCTC 17555. Eukaryot Cell 11, 1584-1585	3	8	0	1	2	1	7	0	2	1	0	2	7	0	4	0
Kluyveromyces wickerhamii	Klw	AEAV01000001-AEAV01000510	Baker CR, Tuch BB, Johnson AD (2011) Extensive DNA-binding specificity divergence of a conserved transcription regulator. Proc Natl Acad Sci U S A 108, 7493-7498.	4	6	0	1	2	1	6	0	3	1	0	1	8	0	4	0
Komagataella pastoris	Kop_b	FN392319-FN392325	De Schutter K, Lin YC, Tiels P et. al. (2009) Genome sequence of the recombinant protein production host Pichia pastoris. Nat Biotechnol 27, 561-566.	1	3	2	0	1	2	4	0	2	1	0	2	5	0	2	0
Kuraishia capsulata	Kc	HG793125-HG793131	Morales L, Noel B, Porcel B et. al. (2013) Complete DNA sequence of Kuraishia capsulata illustrates novel genomic features among budding yeasts (Saccharomycotina). Genome Biol Evol 5, 2524-2539.	1	3	2	0	1	1	2	0	2	1	0	1	3	0	2	1
Lachancea kluyveri	Lak_a	CM000687-CM000694	Cliften, P., Sudarsanam et. al. (2003) Finding functional features in Saccharomyces genomes by phylogenetic footprinting. Science 301, 71-76.	3	11	0	1	3	1	11	0	2	1	0	4	13	0	4	0

Species	Abbreviation	GenBank accession numbers	Reference	TTA	TTG	CTT	CTC	CTA	CTG	TCT	TCC	TCA	TCG	AGT	AGC	GCT	GCC	GCA	GCG
<i>Lachancea thermotolerans</i>	Lat	CU928165-CU928171, CU928180	Souciet JL, Dujon B, Gaillardin C et. al. (2009) Comparative genomics of protoploid Saccharomycetaceae. <i>Genome Res</i> 19, 1696-1709.	2	8	0	2	3	3	9	0	2	2	0	3	10	0	3	2
<i>Lachancea waltii</i>	Lw	AADM01000001-AADM01000713	Manolis Kellis, Bruce W. Birren, Eric S. Lander (2004) Proof and evolutionary analysis of ancient genome duplication in the yeast <i>Saccharomyces cerevisiae</i> . <i>Nature</i> 428, 617-624.	2	0	0	2	4	2	9	0	3	2	0	5	11	0	4	1
<i>Lodderomyces elongisporus</i>	Loe	CH981524-CH981534, DS236866-DS236881	Butler G, Rasmussen MD, Lin MF et. al. (2009) Evolution of pathogenicity and sexual reproduction in eight <i>Candida</i> genomes. <i>Nature</i> 459, 657-662.	3	3	2	0	0	1	2	0	3	1	0	2	4	0	2	0
<i>Metschnikowia fructicola</i>	Mef	ANFW01000001-ANFW01008430	Hershkovitz V, Sela N, Taha-Salaime L et. al. (2013) De-novo assembly and characterization of the transcriptome of <i>Metschnikowia fructicola</i> reveals differences in gene expression following interaction with <i>Penicillium digitatum</i> and grapefruit peel. <i>BMC Genomics</i> 14, 168.	3	18	4	0	0	4	15	0	4	6	0	4	15	0	3	3
<i>Meyerozyma guilliermondii</i>	Mrg	CH408155-CH408162, DS236883	Butler G, Rasmussen MD, Lin MF et. al. (2009) Evolution of pathogenicity and sexual reproduction in eight <i>Candida</i> genomes. <i>Nature</i> 459, 657-662.	2	5	2	0	0	1	4	0	3	2	0	3	5	0	3	1
<i>Millerozyma farinosa</i>	Mif	FO082046-FO082059	Louis VL, Despons L, Friedrich A et. al. (2012) <i>Pichia sorbitophila</i> , an Interspecies Yeast Hybrid, Reveals Early Steps of Genome Resolution After Polyploidization. <i>G3 (Bethesda)</i> 2, 299-311.	4	14	4	0	0	2	10	0	4	2	0	6	12	0	8	2
<i>Nakaseomyces bacillisporus</i>	Nab	CAPX01000001-CAPX01000186	Gabalton T, Martin T, Marcet-Houben M et. al. (2013) Comparative genomics of emerging pathogens in the <i>Candida glabrata</i> clade. <i>BMC Genomics</i> 14, 623.	7	4	0	1	1	0	6	0	3	1	0	3	7	0	5	0
<i>Nakaseomyces delphensis</i>	Nd	CAPT01000001-CAPT01000179	Gabalton T, Martin T, Marcet-Houben M et. al. (2013) Comparative genomics of emerging pathogens in the <i>Candida glabrata</i> clade. <i>BMC Genomics</i> 14, 623.	4	7	0	1	4	0	9	0	3	1	0	3	4	0	6	1
<i>Naumovozyma castellii</i>	Nac	AACF01000001-AACF01000570	Cliften, P., Sudarsanam et. al. (2003) Finding functional features in <i>Saccharomyces</i> genomes by phylogenetic footprinting. <i>Science</i> 301, 71-76.	7	9	2	0	4	0	13	0	4	1	0	4	13	0	5	0
<i>Naumovozyma dairenensis</i>	Nad	HE580267-HE580278	Gordon JL, Armisen D, Proux-Wera E et. al. (2011) Evolutionary erosion of yeast sex chromosomes by mating-type switching accidents. <i>Proc Natl Acad Sci U S A</i> 108, 20024-20029.	9	8	1	0	4	0	11	0	3	1	0	5	10	0	5	0
<i>Ogataea parapolyomorpha</i>	Ogp	AEOI01000001-AEOI01000013	Ravin NV, Eldarov MA, Kadnikov VV et. al. (2013) Genome sequence and analysis of methylotrophic yeast <i>Hansenula polyomorpha</i> DL1. <i>BMC Genomics</i> 14, 837.	1	3	1	0	1	1	2	0	1	1	0	2	3	0	2	0
<i>Pachysolen tannophilus</i>	Pta	CAHV01000001-CAHV01000267	Liu X, Kaas RS, Jensen PR et. al. (2012) Draft genome sequence of the yeast <i>Pachysolen tannophilus</i> CBS 4044/NRRL Y-2460. <i>Eukaryot Cell</i> 11, 827.	5	2	2	0	1	1	3	0	4	1	0	3	3	0	4	0
<i>Pichia kudriavzevii</i>	Piu	ALNQ01000001-ALNQ01000621	Chan GF, Gan HM, Ling HL et. al. (2012) Genome Sequence of <i>Pichia kudriavzevii</i> M12, a Potential Producer of Bioethanol and Phytase. <i>Eukaryot Cell</i> 11, 1300-1301.	2	3	3	0	2	2	5	0	2	1	0	2	0	0	11	0
<i>Saccharomyces arboricola</i>	Saa	CM001563-CM001578	Liti G, Nguyen Ba AN, Blythe M et. al. (2013) High quality de novo sequencing and assembly of the <i>Saccharomyces arboricolus</i> genome. <i>BMC Genomics</i> 14, 69.	8	9	0	1	3	0	10	0	4	1	0	4	11	0	5	0

Species	Abbreviation	GenBank accession numbers	Reference	TTA	TTG	CTT	CTC	CTA	CTG	TCT	TCC	TCA	TCG	AGT	AGC	GCT	GCC	GCA	GCG
Saccharomyces bayanus	Sab_a	AACG02000001-AACG02000586	Cliften, P., Sudarsanam et. al. (2003) Finding functional features in Saccharomyces genomes by phylogenetic footprinting. Science 301, 71-76.	8	11	0	1	3	0	15	0	5	1	0	4	12	0	5	0
Saccharomyces cerevisiae	Sc_c	NC_001133-NC_001148	Otero JM, Vongsangnak W, Asadollahi MA et. al. (2010) Whole genome sequencing of Saccharomyces cerevisiae: from genotype to phenotype for improved metabolic engineering applications. BMC Genomics 11, 723.	7	10	0	1	3	0	11	0	3	1	0	2	11	0	5	0
Saccharomyces kudriavzevii	Sak	JH796010-JH798063	Cliften, P., Sudarsanam et. al. (2003) Finding functional features in Saccharomyces genomes by phylogenetic footprinting. Science 301, 71-76.	7	10	0	1	3	0	9	0	4	1	0	4	11	0	5	0
Saccharomyces mikatae	Smi	AACH01000001-AACH01002808	Kellis, M., Patterson et. al. (2003) Sequencing and comparison of yeast species to identify genes and regulatory elements. Nature 423, 241-254.	6	8	0	1	3	0	10	0	2	1	0	3	9	0	5	0
Saccharomyces paradoxus	Sap_a	AABY01000001-AABY01000832	Kellis, M., Patterson et. al. (2003) Sequencing and comparison of yeast species to identify genes and regulatory elements. Nature 423, 241-254.	7	0	0	1	3	0	11	0	3	1	0	4	11	0	5	0
Scheffersomyces stipitidis	Shs	CM000437, CP000496-CP000502	Jeffries TW, Grigoriev IV, Grimwood J et. al. Right (2007) Genome sequence of the lignocellulose-bioconverting and xylose-fermenting yeast Pichia stipitidis. Nat Biotechnol 25, 319-326.	3	8	3	0	0	1	6	0	2	2	0	2	7	0	3	0
Spathaspora arborariae	Sta	KI547134-KI547163	Lobo FP, Goncalves DL, Alves SL Jr et. al. (2014) Draft Genome Sequence of the D-Xylose-Fermenting Yeast Spathaspora arborariae UFMG-HM19.1AT. Genome Announc 2, e01163-13.	8	3	2	0	0	1	6	0	4	1	0	2	6	0	4	0
Spathaspora passalidarum	Shp	GL996499-GL996506	Wohlbach DJ, Kuo A, Sato TK et. al. (2011) Comparative genomics of xylose-fermenting fungi for enhanced biofuel production. Proc Natl Acad Sci U S A 108, 13212-13217.	6	5	2	0	0	1	6	0	3	1	0	2	7	0	3	0
Tetrapisispora blattae	Ttb	HE806316-HE806325	Gordon JL, Armisen D, Proux-Wera E et. al. (2011) Evolutionary erosion of yeast sex chromosomes by mating-type switching accidents. Proc Natl Acad Sci U S A 108, 20024-20029.	12	9	0	1	3	0	16	0	4	2	0	6	19	0	5	0
Tetrapisispora phaffii	Ttp	HE612856-HE612871	Gordon JL, Armisen D, Proux-Wera E et. al. (2011) Evolutionary erosion of yeast sex chromosomes by mating-type switching accidents. Proc Natl Acad Sci U S A 108, 20024-20029.	9	2	0	1	2	0	8	0	2	1	0	3	9	0	3	0
Torulaspora delbrueckii	Tod	HE616742-HE616749	Gordon JL, Armisen D, Proux-Wera E et. al. (2011) Evolutionary erosion of yeast sex chromosomes by mating-type switching accidents. Proc Natl Acad Sci U S A 108, 20024-20029.	3	8	0	1	4	0	7	0	2	1	0	3	8	0	4	0
Vanderwaltozyma polyspora	Vp	DS480378-DS480658	Scannell DR, Frank AC, Conant GC et. al. (2007) Independent sorting-out of thousands of duplicated gene pairs in two yeast species descended from a whole-genome duplication. Proc Natl Acad Sci U S A 104, 8397-8402.	6	8	0	1	2	0	10	0	3	1	0	4	10	0	4	0
Wickerhamomyces anomalus	Wa	JH119215-JH119570	Schneider J, Rupp O, Trost E et. al. (2012) Genome sequence of Wickerhamomyces anomalus DSM 6766 reveals genetic basis of biotechnologically important antimicrobial activities. FEMS Yeast Res 12, 382-386.	12	6	2	0	2	2	3	0	7	1	0	4	10	0	8	0
Wickerhamomyces ciferrii	Wic	CAIF01000001-CAIF01000364	Schneider J, Andrea H, Blom J et. al. (2012) Draft genome sequence of Wickerhamomyces ciferrii NRRL Y-1031 F-60-10.	7	3	1	0	1	1	4	0	5	1	0	2	6	0	4	0

Species	Abbreviation	GenBank accession numbers	Reference	TTA	TTG	CTT	CTC	CTA	CTG	TCT	TCC	TCA	TCG	AGT	AGC	GCT	GCC	GCA	GCG
			Eukaryot Cell 11, 1582-1583.																
Yarrowia lipolytica	Yl	CR382127-CR382132	Dujon B, Sherman D, Fischer G et. al. (2004) Genome evolution in yeasts. Nature 430, 35-44.	1	3	21	0	2	13	21	0	2	4	0	6	30	0	4	2
Zygosaccharomyces bailii	Zb	CBTC01000001-CBTC010000154	Galeote V, Bigey F, Devillers H et. al. (2013) Genome Sequence of the Food Spoilage Yeast Zygosaccharomyces bailii CLIB 213T. Genome Announc 1.	5	19	0	3	12	0	21	0	4	5	0	9	28	0	9	2
Zygosaccharomyces rouxii	Zr	CU928173-CU928176, CU928178-CU928179, CU928181	Souciet JL, Dujon B, Gaillardin C et. al. (2009) Comparative genomics of protoploid Saccharomycetaceae. Genome Res 19, 1696-1709.	4	10	0	1	5	0	12	0	3	2	0	5	12	0	9	0
Schizosaccharomyces cryophilus	Shc	KE546988-KE547017	Rhind N, Chen Z, Yassour M et. al. (2011) Comparative functional genomics of the fission yeasts. Science 332, 930-936.	3	4	6	0	1	1	11	0	2	2	0	5	11	0	3	1
Schizosaccharomyces japonicus	Sj	KE651166-KE651197	Rhind N, Chen Z, Yassour M et. al. (2011) Comparative functional genomics of the fission yeasts. Science 332, 930-936.	26	3	0	0	2	8	1	0	32	8	0	8	0	0	12	3
Schizosaccharomyces octosporus	Sho	KE503206-KE503210	Rhind N, Chen Z, Yassour M et. al. (2011) Comparative functional genomics of the fission yeasts. Science 332, 930-936.	3	5	8	0	1	1	11	0	2	2	0	5	15	0	3	1
Schizosaccharomyces pombe	Sp	NC_003421, NC_003423-NC_003424	Rhind N, Chen Z, Yassour M et. al. (2011) Comparative functional genomics of the fission yeasts. Science 332, 930-936.	2	4	5	0	1	1	7	0	2	1	0	3	9	0	2	1