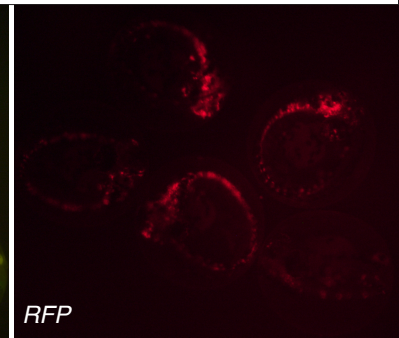
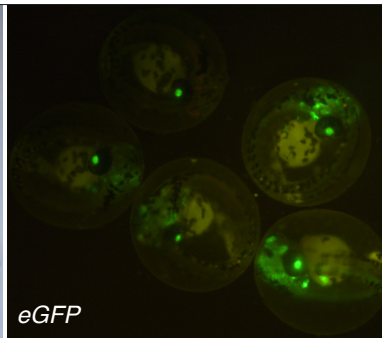
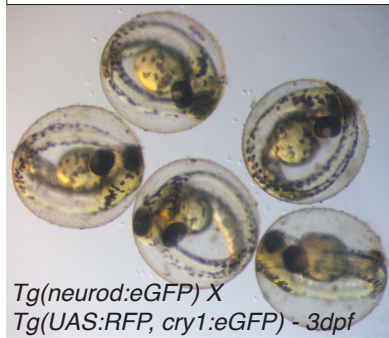
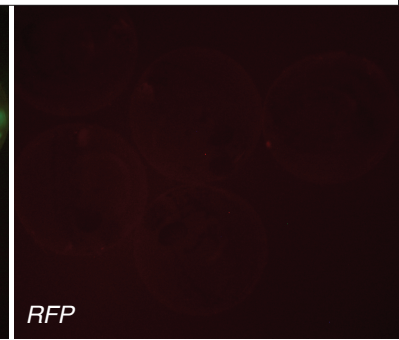
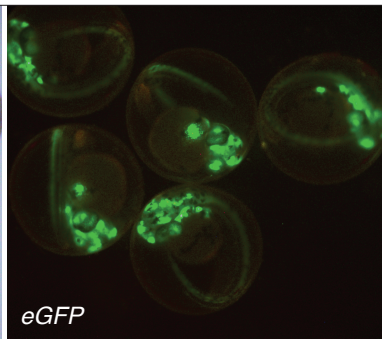
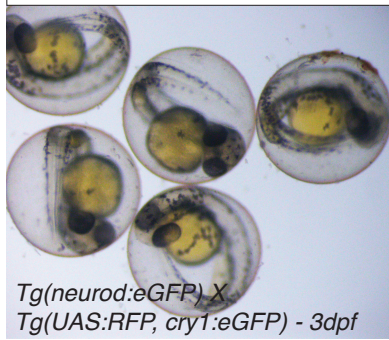


A

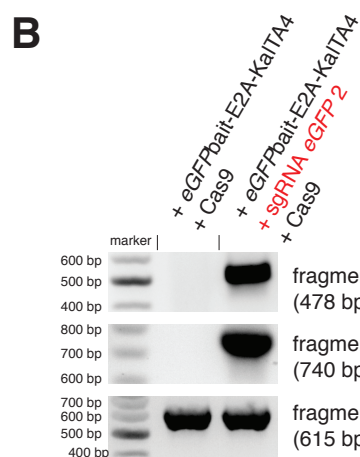
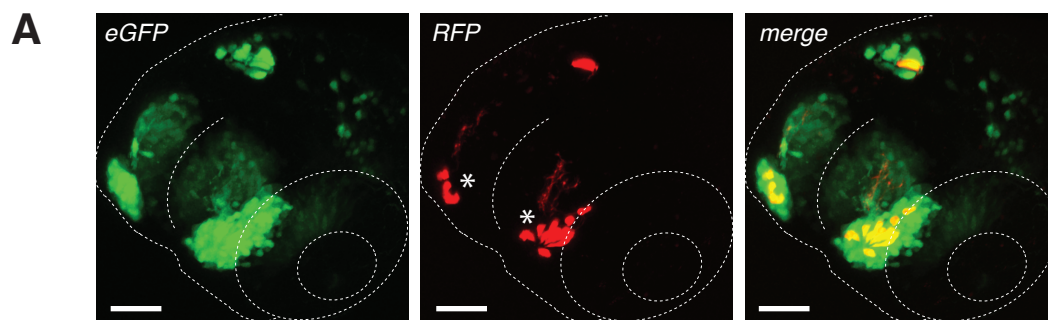
+ eGFPbait-E2A-KalTA4 + sgRNA eGFP 1 + Cas9

**B**

uninjected



Auer et al., Supplementary Figure 1



C

5' junction (fragment B+C):

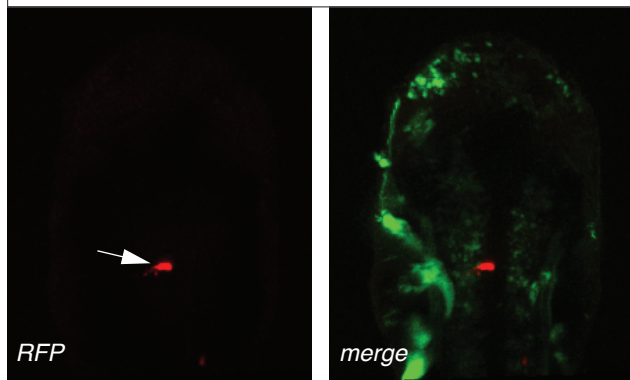
AAACGGCCACAAGTTCAGCGTGTCCGGCGAG	wt
AAACGGCCACAAGTTCAGCCGTTAGCTAATGC	+22 (8x)
ATCATGATAGAGTCCGGCGAG	Δ0 (2x)
AAACGGCCACAAGTTCAGCGTGTCCGGCGAG	

3' junction (fragment D+E):

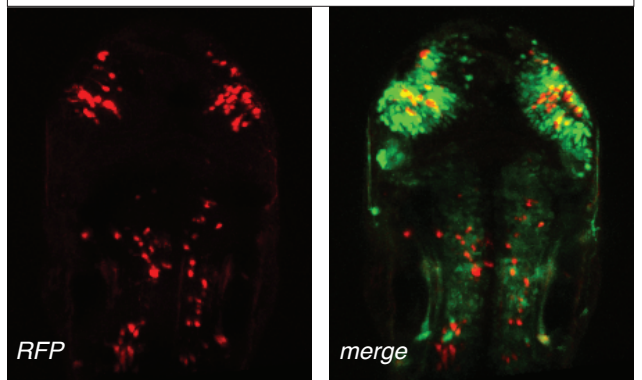
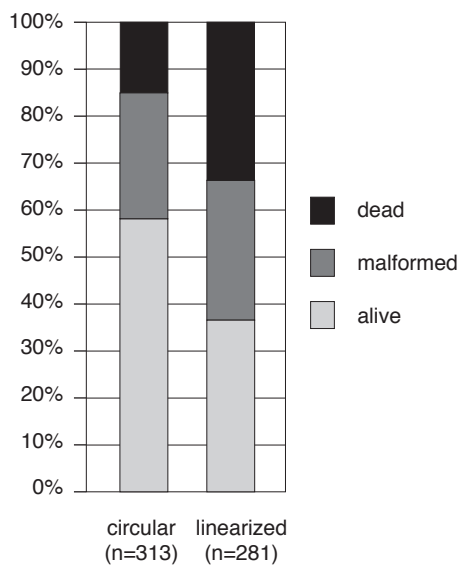
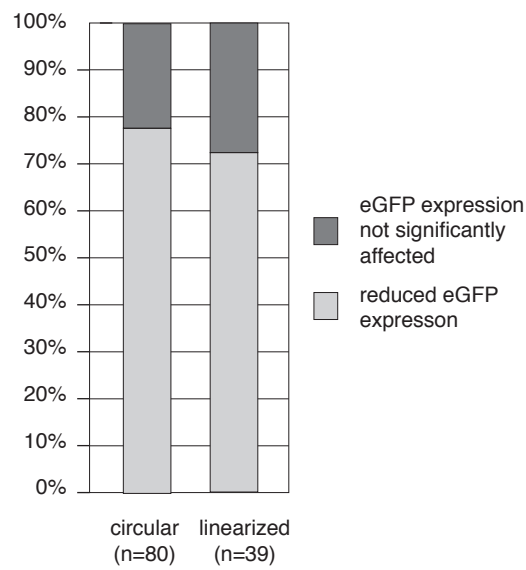
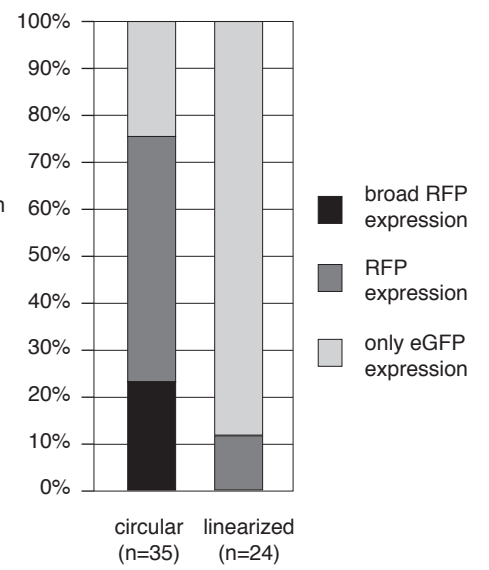
AAACGGCCACAAGTTCAGCGTGTCCGGCGAG	wt
AAACGGCCACAAGTTCAGCGAGGCTTCAGTAC	+7 (6x)
GCTGAG	Δ2 (4x)
AAACGGCCACAAGTTCAGC--GTCCGGCGAG	

A

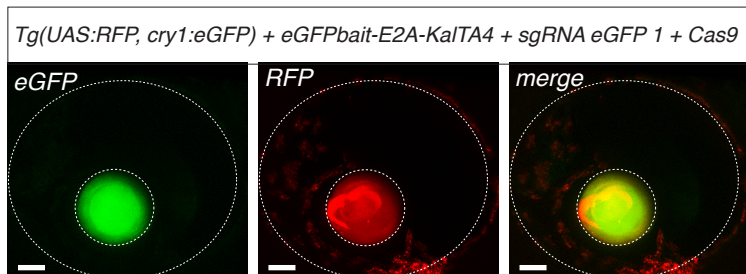
*Tg(neurod:eGFP) X Tg(UAS:RFP, cry1:eGFP) + sgRNA eGFP1 + Cas9 + **linearized** eGFPbait-E2A-KalTA4 donor*

**B**

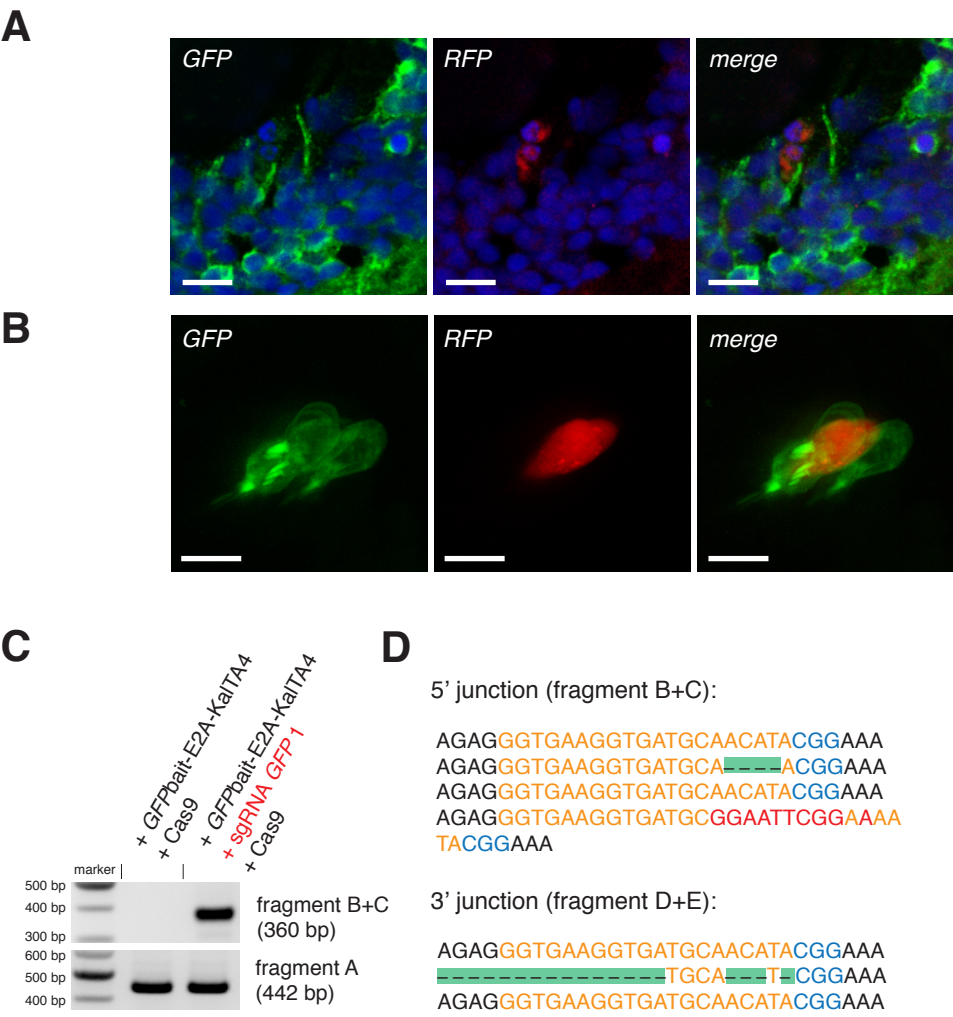
*Tg(neurod:eGFP) X Tg(UAS:RFP, cry1:eGFP) + sgRNA eGFP1 + Cas9 + **circular** eGFPbait-E2A-KalTA4 donor*

**C****D****E**

Auer et al., Supplementary Figure 3

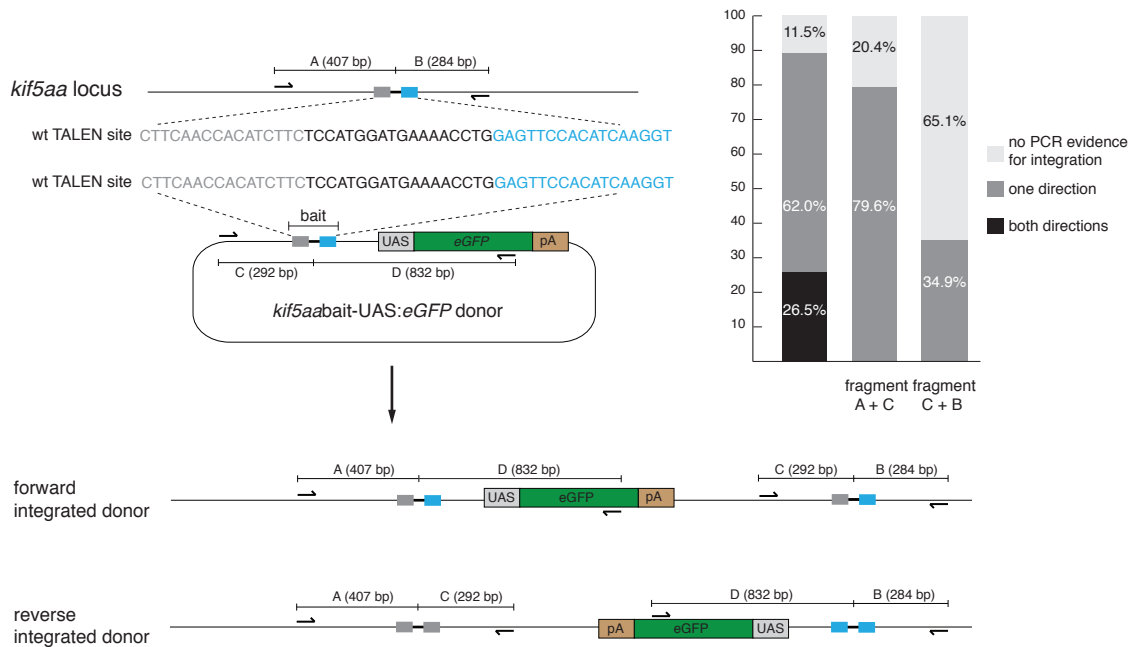


Auer et al., Supplementary Figure 4

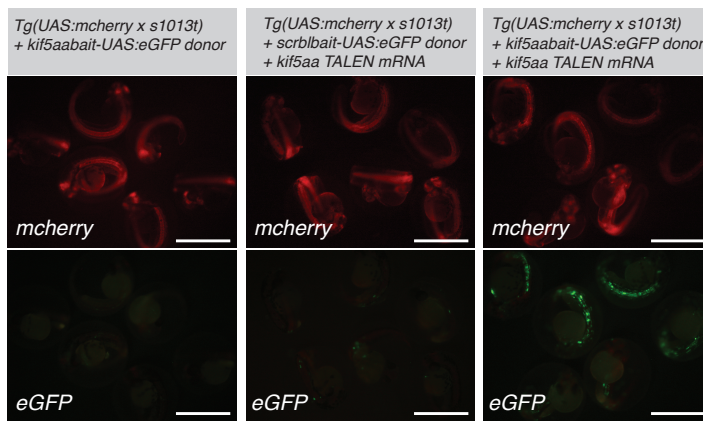


Auer et al., Supplementary Figure 5

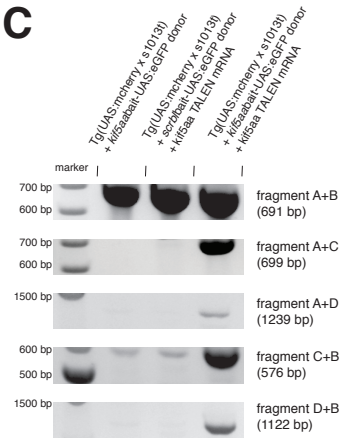
A



B



C



D

5' junctions:

CTTCAACCACATCTTCTCCATGGATGAAAACCTGGAGTTCACATCAAGGT
 CTTCAACCACATCTTCTGAAGCACCACCAACCAACCTAACCCCTAACCCCT
 AACCCTAACCCCTAACCCCTAACCCCTGGAGTTCACATCAAGGT
 CTTCAACC-----TGAGTTCACATCAAGGT
 CTTCAACCACATCTTCTCCATGG-----AACCTGAAGTTCACATCAAGGT
 CTTCAACCACATCTTCTCCA-----TGAGTTCACATCAAGGT
 CTTCAACCACATCTTCTCCATGG-----

wt

+39

Δ24

Δ5

Δ12

Δ86

CTTCAACCACATCTTCTCCATGGATGCATCCATGGAGAAGATGTGGTTGAAG
 CTTCAACCACATCTTCTCCATGGATGAATCATCCATGGAGAAGATGTGGTTGAAG
 CTTCAACCACATCTTCTCATGGATGATCTCCAAGGAGAAGACGTGGTGAAG
 CTTCAACCACATCTTCTCCATGG-----AGAAGATGTGGTTGAAG
 CTTCAACCACATCTTCTCCATGGAGGGTTAGGGTTAACCCATGGAGAAGATGTGGTTGAAG

wt

+3

Δ1

Δ12 (2x)

+9

3' junctions:

CTTCAACCACATCTTCTCCATGGATGAAAACCTGGAGTTCACATCAAGGT
 CTTCAACCACATCTTCTCCATGGACCTGGAGAGCACATCAACCATGAAC
 CTGGAGTTCACATCAAGGT
 CTTCAACCACATCTTCTCCATGG-----AAACCTGGAGTTCACATCAAGGT
 CTTCAACCACATCTTCTCCATGGATGAAAACCTGGAGTTCACATCAAGGT
 CTTCAACCACATCTTCTCCATGGATG-----CTGGAGTTCACATCAAGGT
 CTTCAACCACATCTTCTCCA-----TGAGTTCACATCAAGGT

wt

+19

Δ4

Δ0

Δ5

Δ12

ACCTTGATGTGGAACCTCCAGGTTTAAACCTGGAGTTCACATCAAGGT
 ACCTTGATGTGGAACCTCCAGGTT-----CTCCACATCAAGGT
 ATCCGTGATGTTTTACTCCATGTTTCTCAATGAAAACCTGGAGTTCACATCAAGGT
 ACCTTGATGTGGAACCTCCAG-----GAGTTCACATCAAGGT
 ACCTTGATGTGGAACCTCCAGGTT-----CACCTGGAGTTCACATCAAGGT

wt

Δ13 (2x)

+7

Δ13

Δ4

Auer et al., Supplementary Figure 7: Sequence at the *neurod:eGFP* locus after insertion of the *eGFP*bait-E2A-KalTA4 donor plasmid with perfect repair

HindIII
AACTATCATAAC**AAGCTT**TCAACACACCCTAGGTATGTGATCTGTTCTGTCTTTTATTAATGTGTACCTACTGTGTTTTACGCATTAATGTTGAACTATTTAGACAG
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TAGAGGATTTAAGATAAATGTTAAATCAAAGATCAAATTCAAATAACTGTTAGCTTAAATGTAGTGTCAACCAATTGCACATTTTCAGCCACTTAACTGAAAGAAG
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CGTTGAGCAAAAAAATTTCTTTTAAAGGCTGTCCTATATGTATATTACTTTATGTCTTACACAGTATGAACTATTTCAGACATGGGCTACATTTAAATCCTTCATT
ATTTCAAATAAAACGAGAGGTCTATCAGATTAGGGAAGGCTGATGATGTTACATAATAAAGCTGCATAAAGTAAGCAGCTATGATTCATCCACAGGTCATGTGT
TGTTGCATTTTCGGAGGTGCGCTCTAGTGCAATTCATCACTGTCTCTCATATTAATAAAAAAACTGTCACTGAATGAATGTGTAAAGGTCTGCTTTATCGCTTTGCA
GAGTTC**CGACAT**GGTGAGCAAGGGCGAGGAGCTGTT**CACCGGGTGGTGCCCATCCTGGTCGAGCTGGACGGCGACGTAACGGCCACAAGTTCAGCGTGT**
sgRNA eGFP 2
sgRNA eGFP 1
eGFP bait sequence from donor vector
triple glycine-E2A-triple glycine linker
theoretical cleavage site
CCGGCGAGGGCGAGGGCGATGCCACCTACGGCAAGCTGACCTGAAGTTCATCTGCACCACCGGCAAGCTGCCCGTGGCCACCCCTCGTGACCACC
CTGACCTACGGCGTGCAAGTCTCAGCCGCTACAGGGCGAATTCTGCAGATATCCAGGAGGAGGACAGTGTACTAATTATGCTCTCTTGAAATTGGCTGGAG
ATGTTGAGAGCAACCCAGGTCCCGGAGGAGGAAAAGTGTCTCATCCATCGAGCAAGCCTGCGACATTTGTCTGGCTTAAGAAGCTGAAATGCTCCAAGGAAAA
GCCGAAATGTGCCAAATGCCTGAAGAACAATTGGGAATGTCGTTACTCTCCCAAAACCAAGCGAAGTCCACTCACAAGGGCTCATCTGACCGAAGTGAGAGAGC
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GGCTTATTCGTCCAGGACAATGTGAACAAAGACGCTGTGACAGACCGATTGGCAAGTGTGAGACCGATATGCCTCTGACACTGAGACAGCACAGAATCAGCG
CTACTTCCTCAAGCGAAGAGTCTTCTAACAAGGGACAGAGACAGCTGACTGTTTCGAGCAGGTGACCCCGTCCCGGGCCGACGCCCTGGACGACGGCGACC
TGGACATGCTGCCTGCTGATGCTCTCGATGATTTCGATCTGGATATGCTCCCGGCCGACGCCCTGGACGACTACGACCTGGACATCCTCCCGGGTAACATAAGT
AAGGATCTCGACCGATCCTGAGAACTTCAGGGTGAGTTTGGGGACCCTTGATTGTTCTTTCTTTTCGCTATTGTAAATTTCATGTTATATGGAGGGGGCAAAGT
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GAGGCCCGCACCGATCGCCCTTCCCAACAGTTGCGCAGCCTGAATGGCGAATGGACGCGCCCTGTAGCGGCGCATTAAGCGCGGCGGGGTGTGGTGGTTACG
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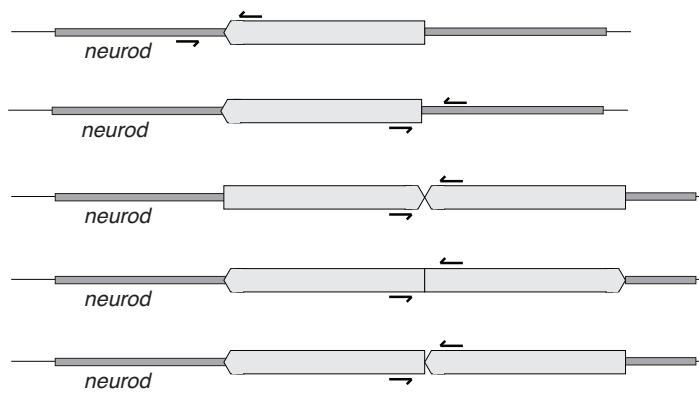
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 GACACCACGATGCCTGTAGCAATGGCAACAACGTTGCGCAAACTATTAAGTGGCGAACTACTTACTCTAGCTTCCCGGCAACAATTAATAGACTGGATGGAGGC
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 GCACTGGGGCCAGATGGTAAGCCCTCCCGTATCGTAGTTATCTACACGACGGGGAGTCAGGCAACTATGGATGAACGAAATAGACAGATCGCTGAGATAGGTG
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 AGAGCGCAGATACCAAATACTGTTCTTCTAGTGAGCCGTAGTTAGGCCACCACTTCAAGAACTCTGTAGCACCAGCTACATACCTCGCTCTGCTAATCCTGTTA
 CCAGTGGCTGCTGCCAGTGGCGATAAGTCGTGTCTTACCGGGTTGGACTCAAGACGATAGTTACCGGATAAGGCGCAGCGGTGCGGCTGAACGGGGGGTTGCG
 TGCACACAGCCCAGCTTGGAGCGAACGACCTACACCGAACTGAGATACCTACAGCGTGAGCTATGAGAAAGCGCCACGCTTCCCGAAGGGAGAAAGGCGGAC
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 GACTGGAAAGCGGGCAGTGAGCGCAACGCAATTAATGTGAGTTAGTCACTCATTAGGCACCCAGGCTTTACACTTTATGCTTCCGGCTCGTATGTTGTGTGG
 AATTGTGAGCGGATAACAATTTACACAGGAAACAGCTATGACCATGATTACGCCAAGCTATTTAGGTGACACTATAGAATACTCAAGCTATGCATC**AAGCTTGG**
 TACCGAGCTCGGATCCACTAGTAACGGCCGCCAGTGTGCTGGAATTCGCCCTTATAGTGGTACCATGGTGAGCAAGGGCGAGGAGCTGTTACCGGGGGTGGT
 GCCCATCCTGGTCGAGCTGGACGGCGACGTAAACGGCCACAAGTTCAGCGTGTCC**sgRNA eGFP 1**
 TCATCTGCACCACCGCAAGCTGCCCGTGCCCTGGCCACCCTCGTGACCACCCTGACCTACGGCGTGAGTGCTTCAGCCGCTACCCCGACCACATGAAGC
 AGCAGGACTTCTTCAAGTCCGCCATGCCGAAGGCTACGTCCAGGAGCGCACCATCTTCTTCAAGGACGACGGCAACTACAAGACCCGCGCCGAGGTGAAGT
 TCGAGGGCGACACCCTGGTGAACCGCATCGAGCTGAAGGGCATCGACTTCAAGGAGGACGGCAACATCCTGGGGCACAAGCTGGAGTACAACGAGCC
 ACAACGTCTATATCATGGCCGACAAGCAGAAGAAGGCGCATCAAGGTGAAGTTCAGATCCGCCACAACATCGAGGACGGCAGCGTGCAGCTCGCCGACCACTA
 CCAGCAGAACACCCCCATCGGCGACGGCCCCGTGCTGCTGCCCCGACAACCACTACCTGAGCACCCAGTCCGCCCTGAGCAAAGACCCCAACGAGAAGCGCG
 ATCACATGGTCCTGCTGGAGTTCGTGACCGCCGCCGGGATCACTCTCGGCATGGACGAGCTGTACAAGTCCGGACTCAGATCTCGAGCTCAAGCTTCGAATTC
 TGCAGTCGACGGTACCGCGGGCCCGGGATCCACCGGATCTAG

HindIII

neurod promoter
 eGFP open reading frame
 eGFP sgRNA 2
 eGFP sgRNA 1

eGFP bait sequence
 pCRII TOPO vector fragment
 triple glycine-E2A-triple glycine linker
 KalTA4 open reading frame

globin intron
 poly A
bold: HindIII restriction sites

A

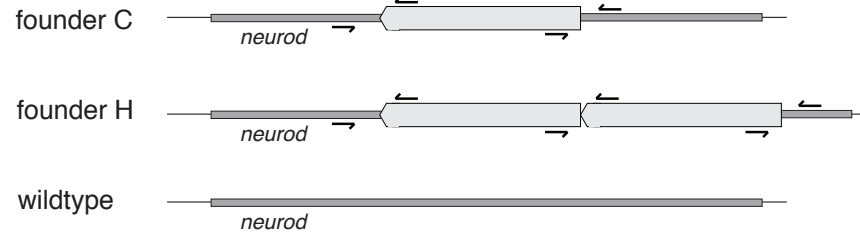
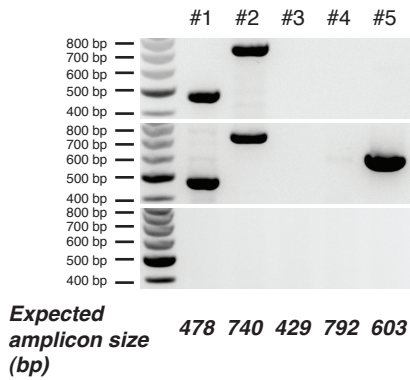
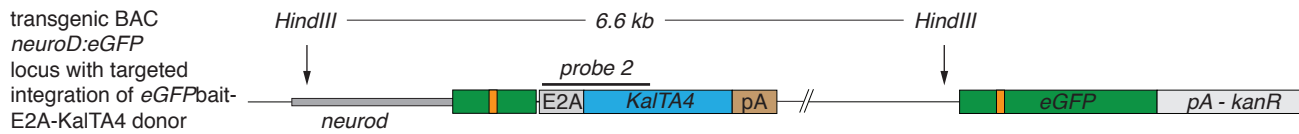
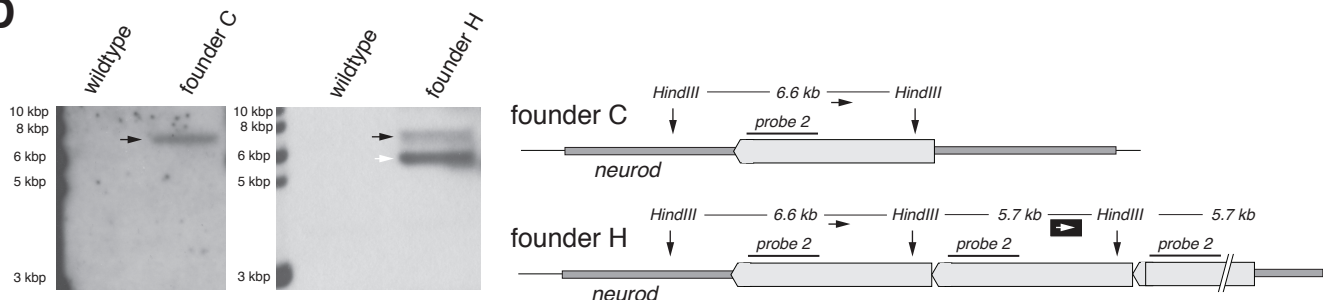
#1 = 5' junction of forward integrated donor plasmid

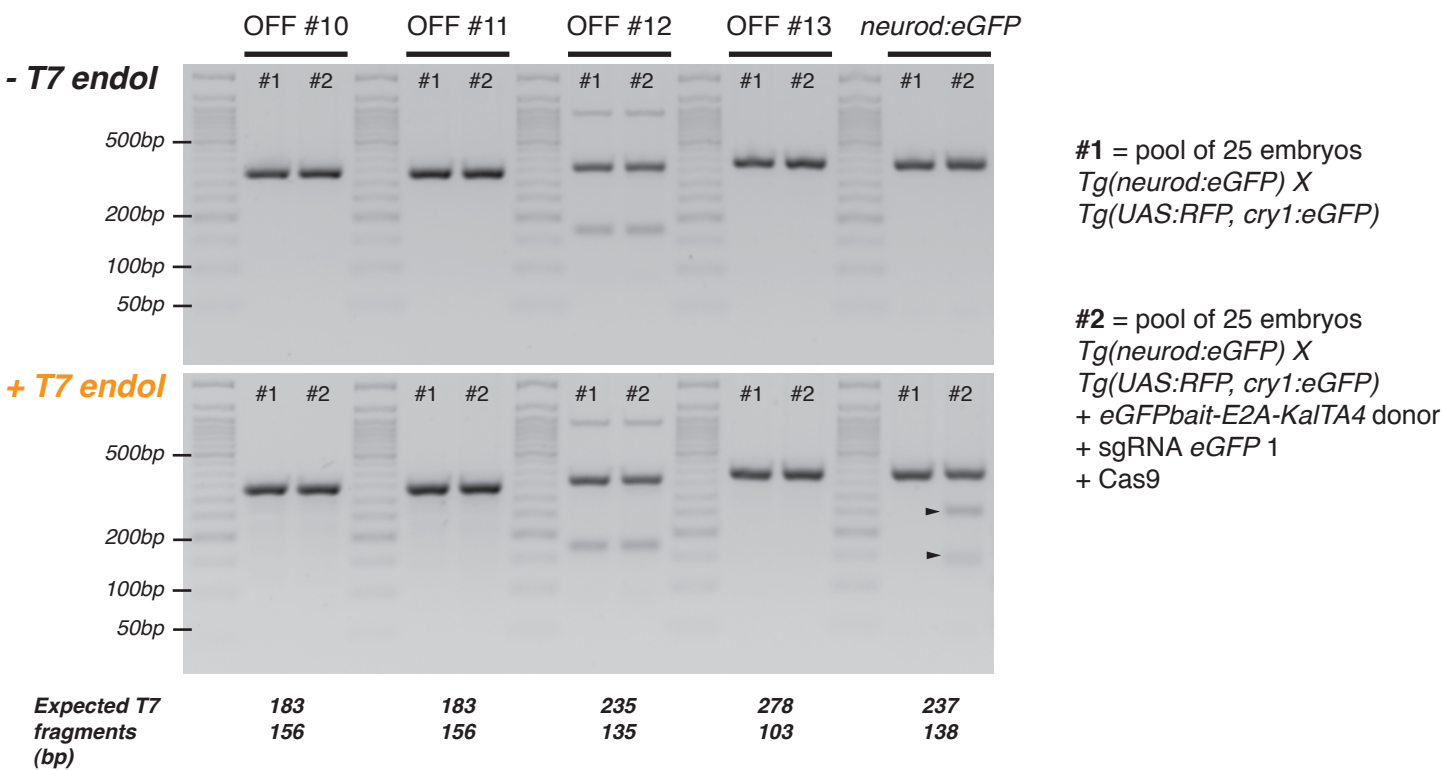
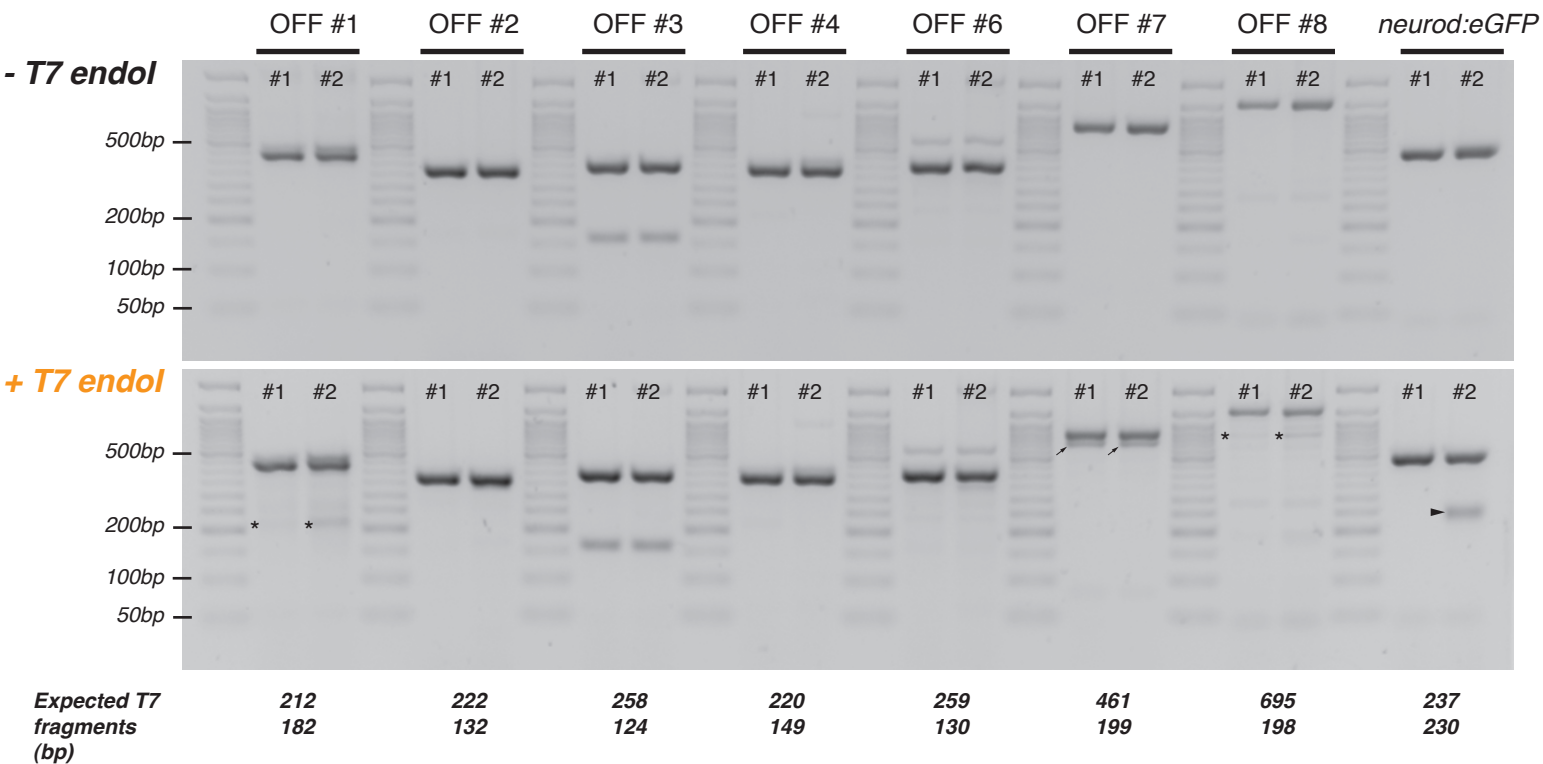
#2 = 3' junction of forward integrated donor plasmid

#3 = head-to-head concatemer of integrated donor plasmid

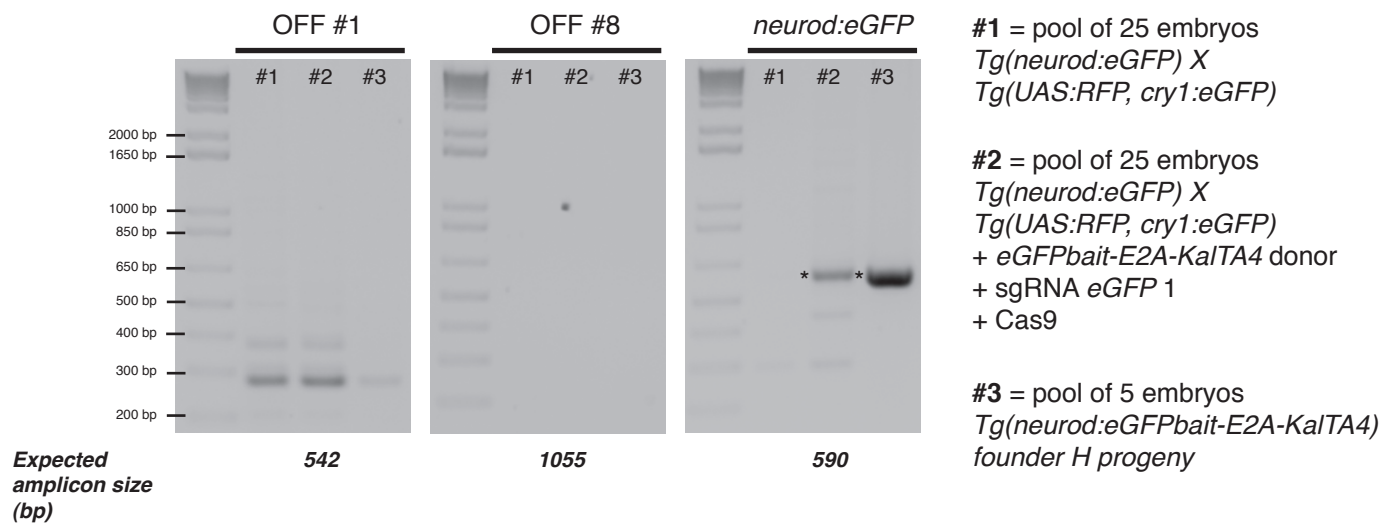
#4 = tail-to-tail concatemer of integrated donor plasmid

#5 = head-to-tail concatemer of integrated donor plasmid

B**C****D**



Auer et al., Supplementary Figure 9



Auer et al., Supplementary Figure 10

Auer et al., Supplementary Table 1: Frequencies and sequences of indel mutations at the *eGFP* and the *kif5aa* locus

sgRNA <i>eGFP</i> 1/Cas9 induced indels at the <i>eGFP</i> locus (examined in DNA of pool of 10 embryos): 66% of mutant sequences (10/15)		
CGAGGGCGAGGGCGATGCCACCTACGGCAAGCTGA	wt	
CGAGGGCGAGGGCGATG-----CGGCAAGCTGA	Δ7 (2x)	
CGAGGGCGAGGTTCTGGCGA-G--A-----GCAAGCTGA	Δ4 (+5, -9)	
CGAGGGCGAGGGCGATGC-----TA-----	Δ33 (3x)	
CGAGGGCGAGGGCGATGCC-----ACGGCAAGCTGA	Δ4 (2x)	
CGAGGGCG-----TGA	Δ24	
CGAGGGCGAGGGCGATGCCCTACGATATCCTACGGCAAGCTGA	+9	
CGAGGGCGAGGGCGATGCCACCTACGGCAAGCTGA	wt (5x)	
sgRNA <i>kif5aa</i> 1/Cas9 induced indels at the <i>kif5Aa</i> locus (examined in DNA of pool of 10 embryos): 22.2% of mutant sequences (4/18)		
CGACCCGCAGCAGATGGGCATCATTTCCCGCAT	wt	
CGA-----AAC-----CTTCATTTCCCGCAT	Δ12	
CGACCCG---CAGATGGGCATCATTTCCCGCAT	Δ3	
CGACCCGCATCATTTCAGATGGGCATCATTTCCCGCAT	+4	
CGACCC-----AGATGGGAATCATTTCCCGCAT	Δ5	
CGACCCGCAGCAGATGGGCATCATTTCCCGCAT	wt (14x)	
Kif5aa TALEN induced indels at the <i>kif5aa</i> locus (examined in DNA of pool of 10 embryos): 60% of mutant sequences		
CTTCAACCACATCTTCTCCATGGATGAAAACCTGGAGTTCCACATCAAGGT	wt	
CTTCAACCACATCTTCT-----CCTGGAGTTCCACATCAAGGT	Δ13	
CTTCAACCACATCTTCTCCA-----TGGAGTTCCACATCAAGGT	Δ12	
CTTCAACCACATCTTCT--AT-----CCTGGAGTTCCACATCAAGGT	Δ11	
CTTCAACCACATCTTCT-----CCTGGAGTTCCACATCAAGGT	Δ13	
CTTCAACCACATCTTCTCCATGGA-GAGAACCTGGAGTTCCACATCAAGGT	Δ1	
CTTCAACCACATCTTCTCCA-----TGGAGTTCCACATCAAGGT	Δ12	
CTTCAACCACATCTTCTCCATGGATGAAAACCTGGAGTTCCACATCAAGGT	wt (4x)	

Auer et al., Supplementary Table 2: Germline transmission of in-frame transgene knock-in in 5 founder fish for the *neurod:eGFP* locus

Founder fish	F1 individuals evaluated	Positive individuals	% of germline with an in-frame transgene knock-in event
founder C	244	3	1.2%
founder G	313	42	13.4%
founder H	272	93	34.2%
founder J	83	14	16.9%
founder K	97	6	6.2%
		<u>Average</u>	<u>14.4%</u>

Auer et al., Supplementary Table 3: Genomic location of potential off target sites of sgRNA *eGFP 1*

Nr.	Sequence	Number of Mis- matches	Genomic location (chromosome:start-stop:strand)
Off#1	GGTGAGGGCAATGCAATATACGG	5	1:40240686-40240708:1
Off#2	GGAGAGAGCGATGCTGCCTCCGG	5	1:49910353-49910375:1
Off#3	GACGAGATCAATGCCACCCACGG	5	10:9919676-9919698:1
Off#4	GGAGTGGCCGCTGCCTCCTACGG	5	10:35974734-35974756:1
Off#5	GGTGAGGGCAATACAACATACGG	5	10:44559443-44559465:1
Off#6	TGCGAGGGCTATGCCACCTTGGG	4	13:10485228-10485250:-1
Off#7	GGCCAGGGCGAGGGCACCGGCGG	5	16:43707826-43707848:-1
Off#8	GGCCAGGGCGAGGGCACCGCCGG	5	16:43733094-43733116:-1
Off#9	GGTGAGGGCGATACAATATACGG	4	14:47813279-47813301:1
Off#10	GGCGCTGGCGGTGTCACCTCCGG	5	2:29982044-29982066:1
Off#11	GGCGCTGGCGGTGTCACCTCCGG	5	Zv9_scaffold3491:89882-89904:-1
Off#12	GGCGAGTGCATGCCGCTTGCGG	5	21:4414337-4414359:-1
Off#13	GGCTAGCGCGATGTCATCTTCGG	5	9:45028926-45028948:-1
Off#14	GGTGAGGGCAATACAACATACGG	5	11:6308610-6308632:-1

Auer et al., Supplementary Table 4: TALEN and sgRNA target (5' to 3') and TALEN protein sequences.

Kif5aa-TALEN-L (target sequence TCTTCAACCACATCTTCT)
MAPKKKRKVYPYDVPDYAGYPYDVPDYAGSYPYDVPDYAAHGTVDLRTLGYSSQQQEKI KPKVRSTVAQHHEALVGHGFTTHAHIVALSQHPAALGTVAVKYQDMIAALPEATHEAIVGV GKQWSGARALEALLTVAGELRGPPLQLDTGQLLKIAKRGGVTAVEAVHAWRNALTGAPL NLTPEQVVAIASHDGGKQALETVQRLLPVLCQAHGLTPAQVVAIASNGGGKQALETVQRL LPVLCQAHGLTPEQVVAIASNGGGKQALETVQRLLPVLCQAHGLTPEQVVAIASHDGGGKQ ALETVQRLLPVLCQAHGLTPAQVVAIASNIGGKQALETVQRLLPVLCQAHGLTPDQVVAIA SNIGGKQALETVQRLLPVLCQAHGLTPDQVVAIASHDGGKQALETVQRLLPVLCQAHGLT PAQVVAIASHDGGKQALETVQRLLPVLCQAHGLTPAQVVAIASNIGGKQALETVQRLLPVL CQAHGLTPDQVVAIASHDGGKQALETVQRLLPVLCQAHGLTPAQVVAIASNIGGKQALET VQRLLPVLCQAHGLTPDQVVAIASNGGGKQALETVQRLLPVLCQAHGLTPEQVVAIASHD GGKQALETVQRLLPVLCQAHGLTPAQVVAIASNGGGKQALETVQRLLPVLCQAHGLTPEQ VVAIASNGGGKQALETVQRLLPVLCQAHGLTPEQVVAIASHDGGKQALETVQRLLPVLCQ AHGLTPAQVVAIASNGGGRPALESIVAQLSRPDPALAALTNDHLVALACLGGRPALDAVK KGLPHAPALIKRTNRRIPERTSHRVAGSQLVKSELEKKSELRHKLKYVPHEYIELIEIARN STQDRILEMKVMEFFMKVYGYRGKHLGGSRKPDGAIYTVGSPIDYGVIVDTKAYSGGYNLP IGQADEMQRYVEENQTRNKHINPNEWWKVYPSSVTEFKFLVSGHFKGNYKAQLTRLNH ITNCNGAVLSVEELLIGGEMIKAGTLTLEEVRKFNNGEINFRS
Kif5aa-TALEN-R (target sequence TACCTTGATGTGGAATC)
MAPKKKRKVYPYDVPDYAGYPYDVPDYAGSYPYDVPDYAAHGTVDLRTLGYSSQQQEKI KPKVRSTVAQHHEALVGHGFTTHAHIVALSQHPAALGTVAVKYQDMIAALPEATHEAIVGV GKQWSGARALEALLTVAGELRGPPLQLDTGQLLKIAKRGGVTAVEAVHAWRNALTGAPL NLTPEQVVAIASNIGGKQALETVQRLLPVLCQAHGLTPDQVVAIASHDGGKQALETVQRL PVLCQAHGLTPAQVVAIASHDGGKQALETVQRLLPVLCQAHGLTPAQVVAIASNGGGKQA LETVQRLLPVLCQAHGLTPEQVVAIASNGGGKQALETVQRLLPVLCQAHGLTPEQVVAIAS NNGGKQALETVQRLLPVLCQAHGLTPEQVVAIASNIGGKQALETVQRLLPVLCQAHGLTP DQVVAIASNGGGKQALETVQRLLPVLCQAHGLTPEQVVAIASNNGGKQALETVQRLLPVL

CQAHGLTPEQVVAIASNNGGKQALETVQRLLPVLCQAHGLTPEQVVAIASNNGGKQALET VQRLLPVLCQAHGLTPEQVVAIASNNGGKQALETVQRLLPVLCQAHGLTPEQVVAIASNIG GKQALETVQRLLPVLCQAHGLTPDQVVAIASNIGGKQALETVQRLLPVLCQAHGLTPDQV VAIASHDGGKQALETVQRLLPVLCQAHGLTPAQVVAIASNNGGKQALETVQRLLPVLCQA HGLTPEQVVAIASHDGGRPALLESIVAQLSRPDPALAALTNDHLVALACLGGRPALDAVKK GLPHAPALIKRTNRRIPERTSHRVAGSQLVKSELEEKSELRHKLKYVPHEYIELIEIARNS TQDRILEMKVMEFFMKVYGYRGKHLGGSRKPDGAIYTVGSPIDYGVIVDTKAYSGGYNLPI GQADEMQRYVEENQTRNKHINPNEWWKVYPSSVTEFKFLFVSGHFKGNYKAQLTRLNHI TNCNGAVLSVEELLIGGEMIKAGTLTLEEVRKFNNGEINFRS	
sgRNA <i>eGFP</i> 1	GGCGAGGGCGATGCCACCTA
sgRNA <i>eGFP</i> 2	GGCCACAAGTTCAGCGTGTC
sgRNA <i>GFP</i> 1	GGTGAAGGTGATGCAACATA
sgRNA <i>kif5aa</i> 1	GGAATGATGCCCATCTGCTGC

Auer et al., Supplementary Table 5: Primers used for insertion mapping and concatemer detection (5' to 3')

<i>Tg(neurod:eGFP) locus</i>	
map_neurod_fwd (binds to the <i>neurod</i> promoter)	GTAAGCAGCTATGATTCATCCACAGGTCATG
map_eGFP_rev (binds within the eGFP ORF)	GGCCATGATATAGACGTTGTGGCTGTTGTAGT TG
map_topo_fwd (binds within the topo vector)	CCGACTGGAAAGCGGGCAGTGAGCGCAACGC
map_E2A_rev (binds within the E2A-KalTA4 fragment)	TTCCTCCTCCGGGACCTGGGTTGCTC
<i>Tg(pou4f3:mGFP) locus</i>	
map_pou4f3_fwd (binds to the <i>pou4f3</i> promoter)	CATGTCAGGTCCACTCACCTGCAAGGTG
map_GFP_rev (binds within the GFP ORF)	GTCCTTGAAGAAGATGGTCCTC
map_topo_fwd (binds within the topo vector)	CCGACTGGAAAGCGGGCAGTGAGCGCAACGC,
map_E2A_rev (binds within the E2A-KalTA4 fragment)	TTCCTCCTCCGGGACCTGGGTTGCTC
<i>Tg(vsx2:eGFP) locus</i>	
map_vsx2_fwd (binds to the <i>vsx2</i> promoter)	CCTGTATCCTCTGGAATTCTAAATCGCTAAGT CGC
map_eGFP_rev (binds within the eGFP ORF)	GGCCATGATATAGACGTTGTGGCTGTTGTAGT TG
map_topo_fwd (binds within the topo vector)	CCGACTGGAAAGCGGGCAGTGAGCGCAACGC
map_E2A_rev (binds within the E2A-KalTA4 fragment)	TTCCTCCTCCGGGACCTGGGTTGCTC

<i>kif5aa locus</i>	
map_kif5aa_fwd (binds upstream of the kif5aa TALEN site)	GTGTCAGTTAAATGTCTCTGGTTTGACGATAG TG
map_kif5aa_rev (binds downstream of the kif5aa TALEN site)	GAACACAAGACAAGATATCCTGAAGGATGCTG GAC
map_topo_fwd (binds within the topo vector)	CCGACTGGAAAGCGGGCAGTGAGCGCAACGC
map_eGFP_rev (binds within the UAS-eGFP fragment)	GGCCATGATATAGACGTTGTGGCTGTTGTAGT TG

Auer et al., Supplementary Table 6: Primers used for T7 analysis (5' to 3') and donor plasmid integration analysis at off-target sites

Nr.	Fwd primer	Rev primer
Off#1	tgacatgggtatgacacgggta	ctgaatcactggctgcgtca
Off#2	ggcctgcttctgttctctagca	tcccgtatgtgaccattca
Off#3	agtcctggcgacttgctcac	gtccgctcagggaagactca
Off#4	tttccggcagattctcaaa	ctgtcaggacagcaggaca
Off#5	atgttctccccgtgttggtg	ctcatccctttgtgccacct
Off#6	acggacaacatccagcatcc	agcgctaccaagcagaccaa
Off#7	tcctgtggctgattgagagtga	ctaccaatcaaagtaaatttcag
Off#8	tcctgtggctgaatgagagtga	ttccaaaattcctttagtgcac
Off#9	tgttgcatgggttccttc	ttacacagcggatgcccttc
Off#10	gcgagtgcacccaaatctc	ttcgggtggcaacaatgatg
Off#11	gcgagtgcacccaaatctc	ttcgggtggcaacaatgatg
Off#12	cgcccagatttaaccacaa	agggatgagtgtccccaggt
Off#13	gttcgcgcttttcctgtct	tctgtccaccacgttgtga
Off#14	cgaaacaacaaggcttcctc	tgcggaattttacaggcagtg
neurod:eGFP (a)	catttcggaggtgcgctcta	gtgtcgccctcgaacttcac
neurod:eGFP (b)	cgctttgcagagttccgaca	gtgtcgccctcgaacttcac
intKaLTA4 (for mapping of E2A-KalTA4 insertion)	gctcgatggatgagagcagttt	