

Production of lentiviruses

Version LG-23/08/2005

Adapted protocol from Dirac and Bernards (2003) J Biol Chem 278:11731-11734

Please check <http://www.nki.nl/nkidep/vansteensel> for updated versions of this protocol.

Cell culture

- Grow 293T cells in 10 cm tissue culture dishes
 - 293T cells are cultured in DMEM + 10% FCS + P/S
- Transfect cells at 80-90% confluency

Transfection (per 10 cm tissue culture dish)

- Cells are transfected by calcium-phosphate co-precipitation
- Prepare the DNA/CaCl₂/dH₂O mix:

◦ Envelope plasmid:	pMD-G	3.5 µg
◦ Packaging construct:	pCMV-ΔR8.2	6.5 µg
◦ Rev-encoding plasmid:	pRSV-Rev	2.5 µg
◦ pL-transfer construct:		10.0 µg
◦ 2.5M CaCl ₂ :		50.0 µl
◦ dH ₂ O:		Adjust to 500 µl
- Add 500 µl 2×HBS (pH 7.00), dropwise, while vortexing
- Incubate the precipitate 5 min. at RT
- Add 1 ml precipitate to the tissue culture dish, dropwise
- Gently shake the plate to mix the precipitate with the medium
- Transfer the transfected 293T cells to the lentivirus lab
- Incubate the cells o/n, 37°C, 5% CO₂
- Replace medium with 6ml fresh medium
- Incubate the cells o/n, 37°C, 5% CO₂

Harvest virus

- Harvest virus on 3 consecutive days:
 - Take off medium
 - Add 6 ml fresh medium to the dish
 - Filter through 0.45 μm filter
 - Store filtered medium at 4°C
- After the final harvest:
 - Combine filtered media
 - Aliquot in 1 ml fractions
 - Store at -70°C

Notes

- All plasmids are ampicillin-resistant
- 2×HBS (pH 7.00):
 - 140 mM NaCl (MW = 58.44)
 - 1.5 mM $\text{Na}_2\text{HPO}_4 \cdot \text{H}_2\text{O}$ (MW = 177.99)
 - 50 mM HEPES (MW = 238.3)
 - Dissolve in dH_2O
 - Set pH with 0.5 M NaOH
 - Filter-sterilise (0.22 μm filter)
 - Aliquot (5 ml fractions)
- Transfection by calcium-phosphate co-precipitation is very pH-sensitive. Make various batches of 2×HBS with different pHs, ranging from 6.80 to 7.20. Do a test-transfection with these 2×HBS batches with a GFP-expressing plasmid.
- 293T cells easily detach from the culture dish. Be very careful with pipetting medium etc.!