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*Genome Res.* 2018 28: 689-698 originally published online April 12, 2018  
Access the most recent version at doi:[10.1101/gr.229062.117](https://doi.org/10.1101/gr.229062.117)

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## Research

# HNRNPA1 promotes recognition of splice site decoys by U2AF2 in vivo

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Alternative pre-mRNA splicing plays a major role in expanding the transcript output of human genes. This process is regulated, in part, by the interplay of *trans*-acting RNA binding proteins (RBPs) with myriad *cis*-regulatory elements scattered throughout pre-mRNAs. These molecular recognition events are critical for defining the protein-coding sequences (exons) within pre-mRNAs and directing spliceosome assembly on noncoding regions (introns). One of the earliest events in this process is recognition of the 3' splice site (3'ss) by U2 small nuclear RNA auxiliary factor 2 (U2AF2). Splicing regulators, such as the heterogeneous nuclear ribonucleoprotein A1 (HNRNPA1), influence spliceosome assembly both in vitro and in vivo, but their mechanisms of action remain poorly described on a global scale. HNRNPA1 also promotes proofreading of 3'ss sequences through a direct interaction with the U2AF heterodimer. To determine how HNRNPA1 regulates U2AF–RNA interactions in vivo, we analyzed U2AF2 RNA binding specificity using individual-nucleotide resolution cross-linking immunoprecipitation (iCLIP) in control and HNRNPA1 overexpression cells. We observed changes in the distribution of U2AF2 crosslinking sites relative to the 3'ss of alternative cassette exons but not constitutive exons upon HNRNPA1 overexpression. A subset of these events shows a concomitant increase of U2AF2 crosslinking at distal intronic regions, suggesting a shift of U2AF2 to “decoy” binding sites. Of the many noncanonical U2AF2 binding sites, *Alu*-derived RNA sequences represented one of the most abundant classes of HNRNPA1-dependent decoys. We propose that one way HNRNPA1 regulates exon definition is to modulate the interaction of U2AF2 with decoy or bona fide 3'ss.

[Supplemental material is available for this article.]

Precursor messenger RNA (pre-mRNA) splicing is catalyzed by a large macromolecular complex composed of five uridine-rich small nuclear ribonucleoprotein particles (U snRNPs) and myriad protein factors (Wahl et al. 2009). This process is required to excise intervening sequences (introns) from the pre-mRNA and ligate protein-coding sequences (exons). The 5' and 3' ends of introns are defined by nearly invariant GU and AG dinucleotides as well as the branch point sequence, respectively. Combinatorial protein–RNA interactions play important roles in defining the splice sites and branch point during spliceosome assembly. The 5' splice site (GU) is recognized by serine and arginine-rich splicing factors (SR Proteins) and through base-pairing interactions with the U1 snRNP (Eperon et al. 1993; Jamison et al. 1995). Similarly, the 3' splice site (3'ss) is decoded by a combination of SR proteins and the U2 snRNP auxiliary factor (U2AF). After early (E) complex assembly, the branch point sequencing is specified through the interaction of U2AF, splicing factor 1 (SF1), and via base-pairing with the U2 snRNP (Ruskin et al. 1988; Berglund et al. 1997). The responsibility for 3'ss recognition is shared by the two subunits of U2AF (Merendino et al. 1999; Wu et al. 1999; Zorio and Blumenthal 1999). The small and large subunits of U2AF, encoded by *U2AF1* and *U2AF2*, recognize the AG dinucleotide and the

upstream polypyrimidine tract, respectively (Merendino et al. 1999; Wu et al. 1999; Zorio and Blumenthal 1999). These early steps in spliceosome assembly play critical roles in defining exon–intron boundaries.

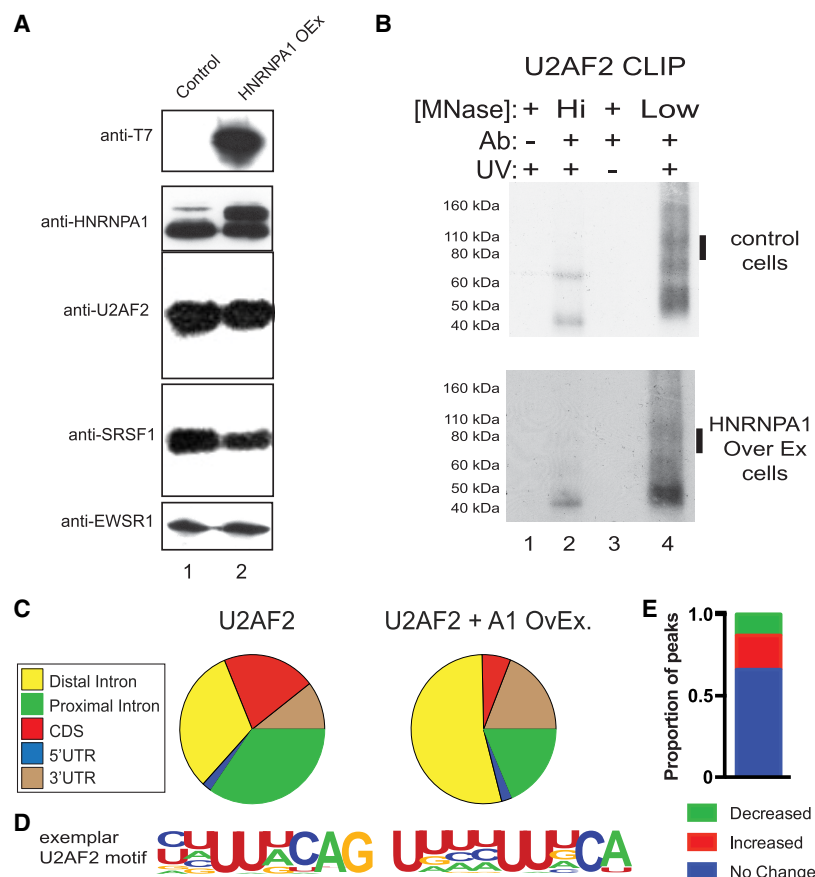
HNRNPA1 is a well-characterized regulator of alternative splicing. One of the primary functions of HNRNPA1 is to prevent inclusion of specific exons (Mayeda and Krainer 1992; Caceres et al. 1994; Yang et al. 1994). Several models for HNRNPA1-dependent exon skipping have been described in the literature. For example, HNRNPA1 can compete with RBPs for binding to juxtaposed regulatory elements that would be normally occupied by splicing enhancer proteins such as SRSF1 (Eperon et al. 2000; Zahler et al. 2004). Another potential mechanism involves oligomerization and spreading of HNRNPA1 from an exonic splicing silencer across a regulated exon, thus antagonizing binding of splicing enhancers and spliceosomal factors, such as U2AF2 (Zhu et al. 2001; Okunola and Krainer 2009). Finally, HNRNPA1 binding to intronic splicing silencers on either side of an alternative exon can dimerize, causing “looping out” of an exon from the pre-mRNA, and promote its exclusion from the final mature transcript (Blanchette and Chabot 1999). A related alternative splicing

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Article published online before print. Article, supplemental material, and publication date are at <http://www.genome.org/cgi/doi/10.1101/gr.229062.117>.

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**Figure 1.** Crosslinking immunoprecipitation of U2AF2 under HNRNPA1 modulation. (A) Western blot analysis of SRSF1, U2AF2, HNRNPA1, the T7 epitope, and EWSR1 in control (lane 1) and HNRNPA1 overexpression HEK293 cells. (B) Examples of iCLIP autoradiographs for U2AF2 either in control or following overexpression of HNRNPA1. Protein–RNA complex shifts are UV-, antibody-, and micrococcal-nuclease-sensitive. Bars denote the region of nitrocellulose blot excised for RNA isolation for iCLIP library preparation. Micrococcal nuclease treatment at 15 U (high) and 0.015 U (low). (C) CLIPper analysis of iCLIP RNA distribution for U2AF2 in control and HNRNPA1 overexpression conditions. (D) Top HOMER consensus binding motifs for U2AF2 in control and HNRNPA1 overexpression conditions in peaks that do not overlap repetitive elements. (E) Proportion of U2AF2 binding site peaks lost (green), gained (red), or unchanged (blue) in HNRNPA1 overexpression cells.

factor, PTBP1, also promotes exon skipping via a looping mechanism (Chou et al. 2000; Lamichhane et al. 2010). Perhaps most relevant to this work, HNRNPA1 promotes proofreading of the 3'ss by U2AF2. In this case HNRNPA1 enables the U2AF heterodimer to reject suboptimal splice sites (Tavanez et al. 2012). In light of these diverse molecular mechanisms of HNRNPA1-dependent splicing regulation, there is a critical need to test their generality on a global scale.

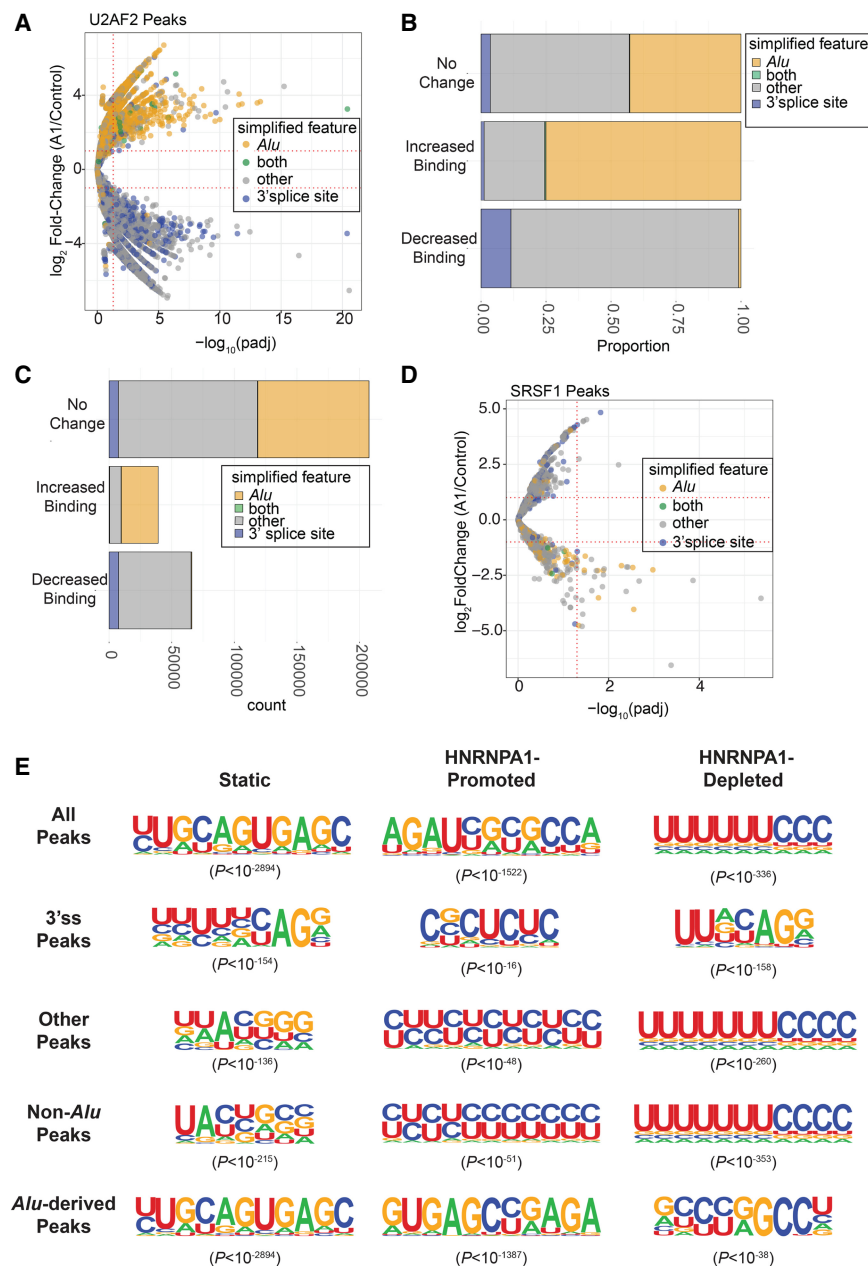
In this study, we investigate how HNRNPA1 influences the association of U2AF2 with 3'ss on a transcriptome-wide scale. We used individual nucleotide resolution crosslinking immunoprecipitation (iCLIP) and high-throughput sequencing to map U2AF2–RNA interactions in control or HNRNPA1 overexpression cell lines. As HNRNPA1 has been shown to be overexpressed in a variety of cancer types (Ushigome et al. 2005; Boukakis et al. 2010; Zhou et al. 2013) and directly involved in specific splicing event that regulate disease phenotype (David et al. 2010; Chen et al. 2012; Bonomi et al. 2013; Loh et al. 2015), this in-cell model will allow us a global look at the effects of HNRNPA1 modulation on splicing and RBP distribution.

## Results

To determine how HNRNPA1 influences the association of U2AF2 with 3'ss on a global scale, we established an HNRNPA1-inducible expression system in HEK293 cells. We then assayed U2AF2 and HNRNPA1 protein–RNA interactions in control or HNRNPA1 overexpression cells using individual nucleotide resolution crosslinking immunoprecipitation and high throughput sequencing (iCLIP-seq) (Konig et al. 2010). We favored the overexpression approach because HNRNPA1 protein levels are elevated in many human cancers (Pino et al. 2003; Ushigome et al. 2005; Chen et al. 2010; Loh et al. 2015; Yu et al. 2015). Induction of HNRNPA1 results in an approximately twofold increase compared to the endogenous protein (and relative to EWSR1) and has no appreciable effect on SRSF1 or U2AF2 steady-state protein levels (Fig. 1A). We used iCLIP to purify HNRNPA1-, SRSF1-, and U2AF2–RNA complexes from control and HNRNPA1 overexpressing cells (Fig. 1B; Supplemental Fig. S2A,B). In all cases, the immunoprecipitated material was both UV- and antibody-dependent, nuclease-sensitive, and produced robust sequencing libraries (Supplemental Tables 1–3). After identification of binding site peaks using CLIPper (Supplemental Table 4; Lovci et al. 2013), we observed differences in the distribution of U2AF2 peaks across genomic locations between control and HNRNPA1 overexpression cell lines (Fig. 1C). Most notably, the proportion of peaks located in coding exons (CDS) or in exon-proximal intronic regions was

reduced, whereas intronic peaks located >500 nucleotides (nt) from exons (distal intron) increased. A similar trend was observed for SRSF1 peaks (Supplemental Fig. 1). In both control cells and HNRNPA1 overexpression cells, intragenic, non-repeat-associated U2AF2 peaks are characterized by a pyrimidine-rich motif, closely resembling authentic 3'ss (Figs. 1D, 2E). Although the sequences are distinct from each other, SRSF1 and HNRNPA1 motifs also appear to be similar between control and HNRNPA1 overexpression cells (Supplemental Fig. 1A,B).

To understand how HNRNPA1 overexpression alters U2AF2–RNA interactions, we identified differentially occupied binding sites in the two cell lines. The union of U2AF2 binding sites from control and HNRNPA1 overexpression cells consisted of nearly 307,000 peaks (Supplemental Tables 5, 6). The majority of these sites (66.3%) were unaffected by HNRNPA1 overexpression (Figs. 1E, 2D). In contrast, 21.5% and 12.2% of U2AF2 binding sites exhibited significant decreases or increases in crosslinking, respectively, upon HNRNPA1 overexpression (Supplemental Table 5). We used BEDTools along with GENCODE and RepeatMasker (Tarailo-Graovac and Chen 2009; Quinlan and Hall 2010;

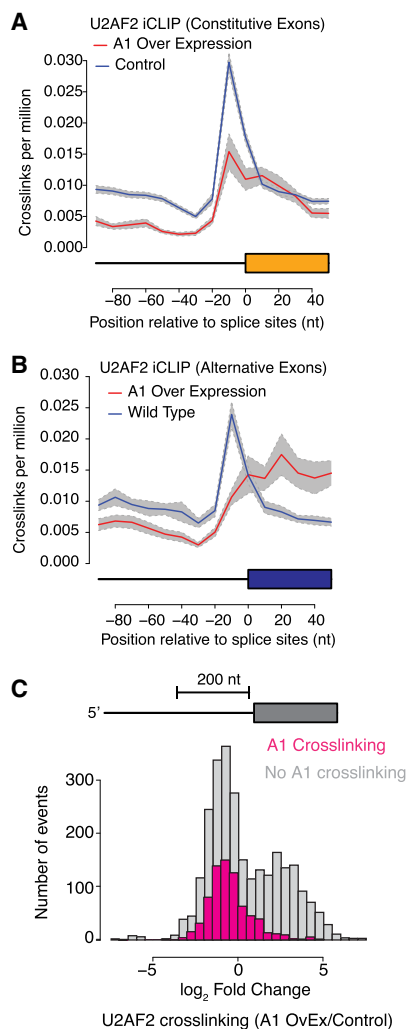


**Figure 2.** Analysis of global changes in U2AF2 binding site and consensus motifs in control and HNRNPA1 overexpression cells. (A) Volcano plot representing the  $\log_2$ (fold-change) in U2AF2 crosslinking peaks (HNRNPA1 overexpression/control) found within *Alu* elements, 3' splice site regions, both *Alu* and 3' splice site regions (both), or other regions outside of these (other). These fold-changes are plotted against their corresponding  $-\log_{10}$ (adjusted *P*-value) associated with each fold-change. Horizontal dotted lines are associated with a  $\log_2$ (fold-change) of  $\pm 2$  and between HNRNPA1 overexpression versus control samples. Vertical dotted lines are associated with  $-\log_{10}$ (adjusted *P*-value) corresponding to 0.05. (B) Stacked bar graph showing the proportion of U2AF2 crosslinking peaks showing no change (top), increased crosslinking peaks (Increased Binding), or decreased crosslinking peaks (Decreased Binding) found within *Alu* elements, 3' splice site regions, both *Alu* and 3' splice site regions (both), or other regions outside of these (other). (C) Stacked bar graph showing the total count of U2AF2 crosslinking peaks showing no change (top), increased crosslinking peaks (Increased Binding), or decreased crosslinking peaks (Decreased Binding) found within *Alu* elements, 3' splice site regions, both *Alu* and 3' splice site regions (both), or other regions outside of these (other). (D) Volcano plot representing the  $\log_2$ (fold-change) in SRSF1 crosslinking peaks found within *Alu* elements, 3' splice site regions, both *Alu* and 3' splice site regions (both), or other regions outside of these (other). (E) HOMER-generated exemplar U2AF2 motifs associated with static (left), HNRNPA1-disrupted (middle), and HNRNPA1-promoted (right) crosslinking peaks at 3' splice sites with HNRNPA1 overexpression. Associated *P*-values for each motif are provided in parentheses.

Wright et al. 2016) to annotate peaks with significantly different U2AF2 crosslinking in control and HNRNPA1 overexpression cells. U2AF2 peaks that were lost after HNRNPA1 overexpression were enriched for 3'ss annotations ( $P < 2.2 \times 10^{-16}$ ,  $\chi^2$  test of independence), whereas peaks gained showed a strong preference for *Alu*-derived RNA sequences ( $P < 2.2 \times 10^{-16}$ ,  $\chi^2$  test of independence) (Fig. 2A–C; Supplemental Fig. 2A). Like U2AF2 peaks, SRSF1 binding sites were also perturbed by HNRNPA1 overexpression, but they exhibit the opposite trend with respect to 3'ss (Supplemental Table 7) and *Alu*-derived RNA sequences (Fig. 2D; Supplemental Fig. 2B,C).

To determine if HNRNPA1 alters the sequence landscape of U2AF2–RNA interactions, we searched for over-represented sequences in both stable and differential peaks in control and HNRNPA1 overexpression cells. We classified U2AF2 peaks into three groups: those overlapping *Alu*-derived sequences, those overlapping 3'ss and “other” sequences, for peaks not part of either class. For each group, we identified enriched sequences from lost, gained, or stable (static) peaks, resulting in nine different classes of U2AF2 binding sites. For most classes, we observe variations of pyrimidine-rich sequences that resemble 3'ss. For example, Figure 2E shows the top three exemplar motifs from stable, HNRNPA1-promoted, or HNRNPA1-disrupted U2AF2 peaks annotated as 3'ss. For the 13,986 stable peaks at 3'ss, ~26% contained the most highly enriched motif shown in Figure 2E ( $P < 10^{-155}$ ). In contrast, at 3'ss peaks that were disrupted in HNRNPA1 overexpression cells, a distinct pyrimidine-rich motif was discovered in ~31% of 13,318 peaks ( $P < 10^{-159}$ ). A much smaller subset of peaks exhibited increased U2AF2 crosslinking in the overexpression cells relative to control. Of these 867 binding sites, 9.6% contained the strong polypyrimidine motif ( $P < 10^{-16}$ ), which closely resembled the static sites. Although polypyrimidine-rich motifs are present in the majority of 3'ss and “other” binding sites, peaks annotated as overlapping *Alu* elements are less pyrimidine-rich (Fig. 2E). These data suggest that *Alu*-derived sequences are an abundant class of noncanonical binding motif that become accessible to U2AF2 in HNRNPA1 overexpression cells.

HNRNPA1 influences association of splicing factors such as U2AF2 with 3'ss



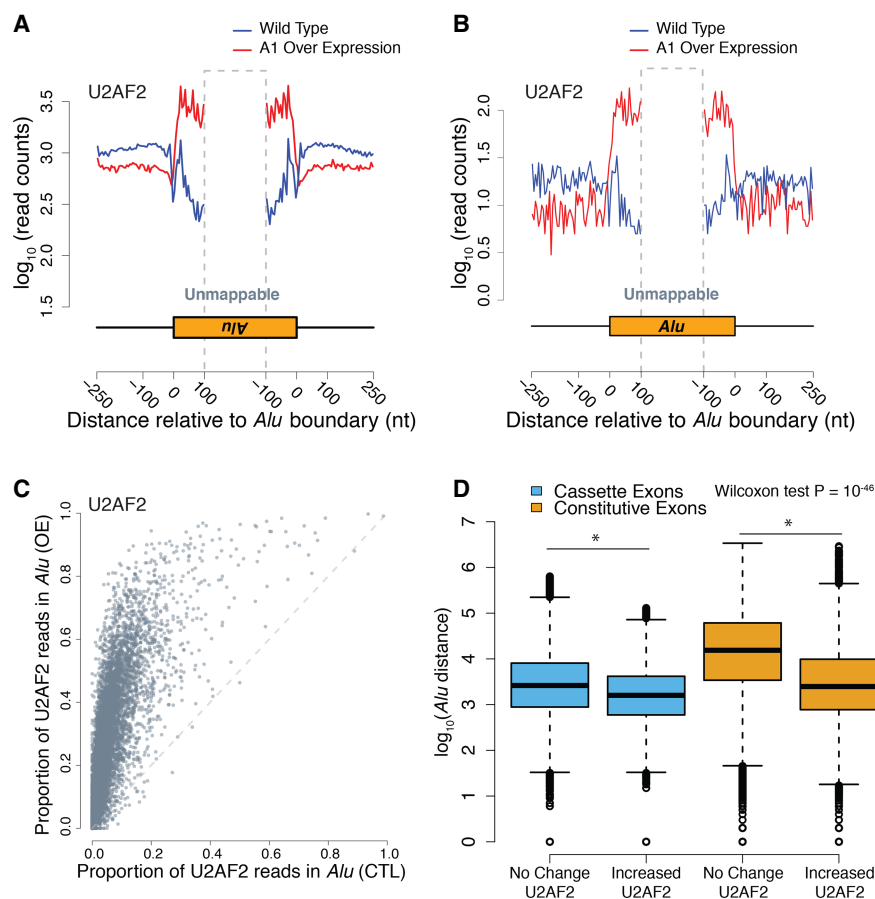
**Figure 3.** HNRNPA1 induced redistribution of U2AF2 crosslinking near 3'ss. (A,B) Normalized crosslinking distribution for U2AF2 in wild-type (blue line) and HNRNPA1 overexpression cell lines (red line) with 95% confidence interval (gray area). Data are divided between constitutive (A) and alternative cassette (B) exons. (C) Distribution of U2AF2 crosslinking changes within 200-bp intron regions near 3'ss of alternative cassette exons. Gray bars correspond to annotated alternative splicing events with no evidence of HNRNPA1 crosslinking in either condition, and pink represents annotated events with detectable HNRNPA1 crosslinking.

on a case-by-case basis (Zhu et al. 2001; Tavanez et al. 2012). However, its effects on splicing-factor RNA targeting on a global scale are not well defined. In order to assess the effects of HNRNPA1 on transcriptome-wide splicing, we determined how titration of HNRNPA1 affected the distribution of SRSF1- and U2AF2-RNA crosslinks relative to 3'ss of constitutive or alternative cassette exons. As suggested by the peak analysis (Supplemental Fig. 3), there are no differences in HNRNPA1 crosslinking sites between control and overexpression cells (Supplemental Fig. 4A,B, left panels, blue and red lines, respectively). SRSF1 crosslinking to exonic sequences was modestly reduced in the HNRNPA1 overexpression cells compared to control, but the positional distribution of the SRSF1 sites relative to the 3'ss was largely unchanged for constitutive and skipped exons (Supplemental Fig. 4A,B, right panels, blue and red lines, respectively). U2AF2 crosslinking distribution relative to the 3'ss was substantially altered in

HNRNPA1 overexpressing cells compared to the control, where a characteristic peak is observed over the 3'ss of both constitutive and skipped exons (Fig. 3A,B, blue line). In contrast, in cells overexpressing HNRNPA1, U2AF2 crosslinking density near alternative exons shifts downstream from the 3'ss and the peak is substantially reduced (Fig. 3B right panel, red line). To further determine if there is a direct relationship between HNRNPA1 binding and changes in U2AF2 or SRSF1 association with transcripts, we examined regions flanking the 3'ss of skipped exons with HNRNPA1 crosslinking in either condition (Fig. 3C; Supplemental Fig. 4C; Supplemental Tables 5–7). In regions with no detectable HNRNPA1 crosslinks, the change in U2AF2 crosslinking exhibits a bimodal distribution, which corresponds to regions flanking the 3'ss that show either increased or decreased U2AF2 crosslinking in HNRNPA1 overexpression cells relative to control cells (Fig. 3C, blue). In contrast, U2AF2 crosslinking to the vicinity of the 3'ss is significantly reduced when a direct association of HNRNPA1 is also evident (Fig. 3C, pink). For example, in both *SRSF6* (serine/arginine-rich splicing factor 6 also known as SRP55) and *PIEZO1* (piezo type mechanosensitive ion channel component 1), we found that U2AF2 crosslinking near the 3'ss is reduced in the cell lines overexpressing HNRNPA1 (Supplemental Fig. 5).

Previous work by Zarnack et al. (2013) demonstrated that HNRNP proteins, such as HNRNPC, compete with U2AF2 for binding to antisense *Alu* elements, in order to repress their exonization. We asked if HNRNPA1 similarly repressed U2AF2 crosslinking to *Alu* elements by measuring crosslink density across the boundaries of intronic *Alu* elements in control and HNRNPA1 overexpression cells. We observed a dramatic increase in U2AF2 crosslinking to *Alu*-containing RNA transcripts compared to control cells (Fig. 4A,B; Supplemental Tables 8–11). Like prior reports (Supplemental Table 8), U2AF2 crosslink density was nearly 10-fold greater in RNA derived from antisense *Alu* elements compared to sense (cf. Fig. 4A to B; Zarnack et al. 2013; Shao et al. 2014). To visualize HNRNPA1-dependent accumulation of U2AF2 in *Alu* elements, we calculated the coverage of all peaks across a consensus sequence assembled from all the *Alu* subtypes in the human genome (see Methods). For example, for subtype *AluSc*, HNRNPA1 overexpression induces U2AF2 crosslinking near the 3' end of the element compared to control cells (Supplemental Fig. 6A). Conversely, HNRNPA1 crosslinking globally decreases over *Alu* elements with overexpression (Supplemental Fig. 6B,C; Supplemental Table 9). In contrast to U2AF2, crosslinking of SRSF1 to antisense *Alu* elements shows no appreciable changes (Supplemental Fig. 6D,E; Supplemental Table 9), suggesting that the effect of HNRNPA1 is specific to U2AF2. In order to assess any indirect effects of HNRNPA1 expression on known regulators of U2AF2-*Alu* binding, we also measured protein levels and cell localization of HNRNPC under control and overexpression of HNRNPA1, which showed no discernable changes (Supplemental Fig. 7).

To determine if *Alu* elements exhibiting HNRNPA1-dependent increases in U2AF2 crosslinking are located in *cis* relative to annotated skipped exons (*Alu* elements upstream of or downstream from a splicing event) or in *trans* (another locus) (Supplemental Fig. 8), we compared the proportion of U2AF2 crosslinks within *Alu*-elements relative to sequences flanking annotated as alternative cassette (skipped) exons in control or HNRNPA1 overexpression cells. The scatter plot shown in Figure 4C (data in Supplemental Tables 10 and 11) demonstrates that the proportion of U2AF2 crosslinks present in *Alu* elements increases significantly upon HNRNPA1 overexpression, whereas



**Figure 4.** HNRNPA1 overexpression correlates with global redistribution of U2AF2 signal to *Alu* RNA elements. (A) Aggregated crosslinking sites on antisense *Alu* elements and nearby regions for U2AF2. Blue represents wild-type binding of the given RNA binding protein, and red represents HNRNPA1 overexpression of the  $\log_{10}$  number of iCLIP read counts across all antisense-*Alu* elements. (B) Aggregated crosslinking sites on sense *Alu* elements and nearby regions for U2AF2. (C) Scatter plot of all human cassette exons measuring the proportion of U2AF2 iCLIP crosslinking sites found within *Alu* elements within the cassette exon event over the total number of crosslinks found within the event. Proportions from control and HNRNPA1 overexpression samples are compared for each individual cassette exon event. (D) Box plot representing the distance of *Alu* elements from cassette exons (blue) and constitutive exons (orange) that show no change in U2AF2 crosslinking versus those that show an increase in U2AF2 crosslinking.

the proportion of HNRNPA1 crosslinks is decreased (Supplemental Fig. 6C; data in Supplemental Tables 12 and 13). Additionally, distribution of distal intronic U2AF2 peaks showing increases with HNRNPA1 overexpression show a significant enrichment for *Alu* elements compared to a null distribution (Supplemental Fig. 9). In contrast, the proportion of SRSF1 crosslinks to *Alu* elements is refractory to changes in HNRNPA1 expression levels (Supplemental Fig. 6E; data in Supplemental Tables 14 and 15). These data demonstrate a global change in U2AF2-*Alu* association and refute the hypothesis that a few spurious *Alu* elements are responsible for the signal observed in Figure 4A. To determine if *cis-Alu* elements are involved in HNRNPA1-regulated alternative splicing, we manually curated the list of 83 splicing events that also had detectable U2AF2 crosslinking. We found that 41% of HNRNPA1-dependent exon skipping events exhibited redistribution of U2AF2 to adjacent *Alu* elements (Table 1). Taken together, these data demonstrate that overexpression of HNRNPA1 influences the association of U2AF2 with both sense and antisense *Alu* elements.

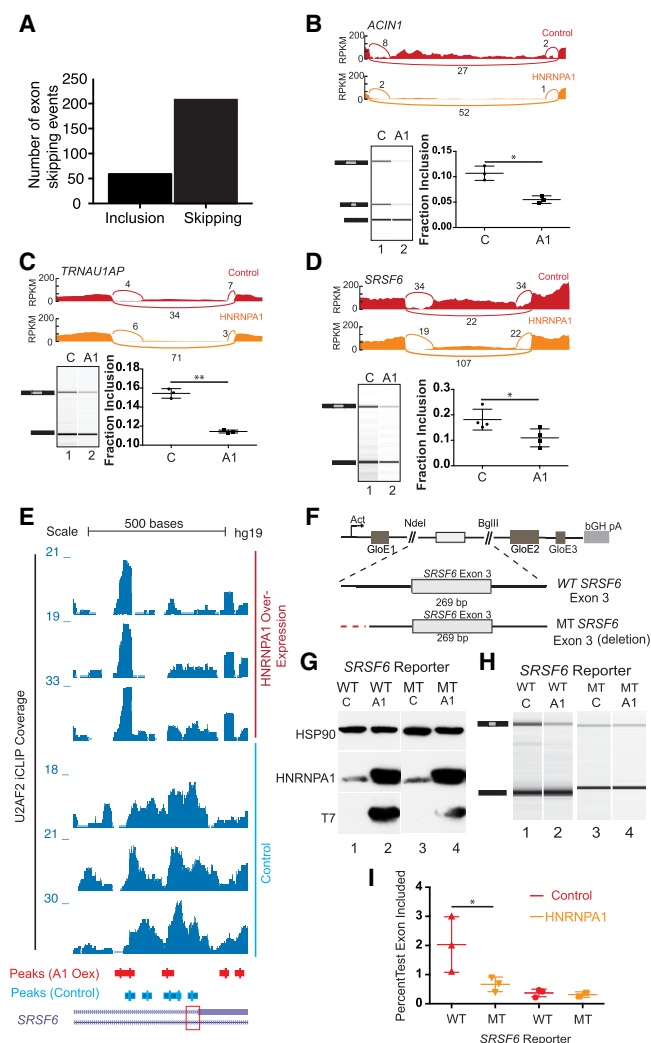
*Alu* elements influence alternative splicing, although the mechanisms are poorly understood (Sorek et al. 2002; Lev-Maor et al. 2008; Gal-Mark et al. 2009; Schwartz et al. 2009; Pastor and Pagani 2011). Splicing regulatory elements often exhibit position-dependent and context-dependent functions (Fu and Ares 2014). To determine if *Alu*-elements with HNRNPA1-dependent changes in U2AF2 crosslinking exhibit positional bias, we measured their distance relative to the 3'ss of constitutive or skipped exons. In general, we observed that *Alu* elements are closer to skipped exons than constitutive exons ( $P = 1.4 \times 10^{-46}$ , Wilcoxon rank-sum test) (Fig. 4D, cf. blue and orange boxes). But yet, those *Alu* elements with HNRNPA1-dependent increases in U2AF2 crosslinking are significantly closer to exons than those that are unchanged ( $P = 9.5 \times 10^{-92}$ ) (Fig. 4D). Taken together, our data suggest that *Alu*-elements may function as *cis*-regulatory elements that compete with authentic exons for binding to splicing factors.

To determine if changes in U2AF2 crosslinking correlated with HNRNPA1-dependent splicing regulation, we sequenced poly(A)<sup>+</sup> selected RNA libraries from control and HNRNPA1 overexpression cells (Supplemental Table 16). Of the 267 HNRNPA1-regulated cassette exons that exhibited at least a 33% change in splicing (Bayes factor >10), the majority exhibited increased levels of exon skipping upon HNRNPA1 overexpression (Fig. 5A; Supplemental Table 17). Eighty-three of the 267 differentially spliced exons also had detectable U2AF2 crosslinking proximal to 3'ss in either control or HNRNPA1-overexpressing

cell lines. Of those 83 exons, ~90% exhibited an HNRNPA1-dependent increase in exon skipping compared to ~70% of exons with no detected U2AF2 crosslinking ( $P < 0.0025$ , Fisher's exact test) (Table 1). To investigate the relationship between exon skipping with loss or gain of U2AF2 crosslinking, we increased the sensitivity of our analysis by using less stringent thresholds for differential splicing (difference >1%, Bayes factor >5) (Supplemental Table 18). This resulted in 574 events, the majority of which favored exon

**Table 1.** Summary of HNRNPA1-dependent changes in exon skipping with detectable U2AF2 crosslinking

	Total no. of events	U2AF2 crosslinking	No U2AF2 crosslinking
HNRNPA1-dependent exon inclusion	59	9	50
HNRNPA1-dependent exon skipping	208	74	134



**Figure 5.** Noncanonical U2AF2 binding sites influence HNRNPA1-dependent exon skipping. (A) Bar graph of all HNRNPA1-dependent splicing changes as detected by MISO analysis. Events determined from the comparison of RNA-seq data derived from HNRNPA1 overexpressing HEK293T cell versus control HEK293T cells. (B–D) Sashimi plots showing read and junction coverage for three genes: *ACIN1*, *SRSF6*, and *TRNAU1AP*. Virtual gel representation of an Agilent 2100 Bioanalyzer DNA 1000 assay of RT-PCR products corresponding to endogenous *ACIN1*, *SRSF6*, and *TRNAU1AP* from control and HNRNPA1 overexpression cells (lower left panel). Bar graph depicting mean exon inclusion for *TRNAU1AP* and *SRSF6* splicing reporters quantified using an Agilent 2100 Bioanalyzer with standard deviation bars. (\*)  $P < 0.05$ ; (\*\*)  $P < 0.01$  (lower right panel). (E) UCSC Genome Browser screenshot of U2AF2 iCLIP read coverage at the *SRSF6* exon 3 locus in control or HNRNPA1 overexpression cells (bottom and top three tracks, respectively). Peaks called by CLIPper are depicted below the coverage tracks. Red box denotes the 3' ss of *SRSF6* exon 3. (F) Splicing reporter constructs created representing wild-type (Wt) or mutant (Mt) versions of alternative exons in *SRSF6*. Noncanonical U2AF2 binding sites in *SRSF6* were mutated (red lines) by deletion. GloE1, GloE2, and GloE3 designate exons 1–3 of beta-globin. The polyadenylation signal from the bovine growth hormone 1 gene is indicated by bGH pA. (G) Representative western blots probed with antibodies against HSP90, HNRNPA1, and the T7 epitope. Lysates were prepared from HEK293T cells transiently transfected with either wild-type (WT) or mutant (MT) *SRSF6* reporter construct and control or T7 epitope-tagged HNRNPA1. (H) Virtual gel representation of an Agilent 2100 Bioanalyzer DNA 1000 assay of RT-PCR products from *SRSF6* splicing construct transfections. (I) Bar graph depicting mean exon inclusion for *SRSF6* splicing reporters quantified using an Agilent 2100 Bioanalyzer with standard deviation bars. (\*)  $P < 0.05$ .

**Table 2.** Summary of HNRNPA1-dependent changes in exon skipping and U2AF2 positioning

	HNRNPA1 crosslinking >1 cpm		No detectable HNRNPA1 crosslinking	
	Increased U2AF2	Decreased U2AF2	Increased U2AF2	Decreased U2AF2
Exon inclusion	1	0	11	18
Exon skipping	3	11	74	64

skipping (424 events). Exon skipping events that also had evidence of HNRNPA1 crosslinking in either condition were enriched for loss of U2AF2 crosslinking as compared to events with no detectable HNRNPA1 crosslinking ( $P < 0.026$  Fisher's exact test) (Table 2). In some cases, HNRNPA1-sensitive exons exhibited redistribution of U2AF2 signal from near 3' ss to distal, upstream crosslinking sites, suggesting their possible function in HNRNPA1-mediated splicing regulation (Fig. 5E; Supplemental Figs. 10–20). We use transient transfection of T7 epitope-tagged HNRNPA1 and RT-PCR to validate several endogenous splicing events exhibiting HNRNPA1-dependent reduction in U2AF2-3' ss crosslinking (Fig. 5B–D, upper panels). As expected, we observed a significant reduction in exon inclusion for *ACIN1*, *SRSF6*, and *TRNAU1AP* in HNRNPA1 overexpression cells relative to the control (Fig. 5B–D, lower panels).

To determine if noncanonical U2AF2 binding sites are involved in HNRNPA1-dependent exon skipping, we created matched pairs of hemoglobin subunit beta (*HBB*) splicing reporter gene constructs (Rothrock et al. 2003) containing the wild-type or mutated intronic elements upstream of the HNRNPA1-sensitive exons found in the *SRSF6* pre-mRNA (Fig. 5E). The *SRSF6* splicing reporter constructs were cotransfected into HEK293T cells with epitope-tagged HNRNPA1 expression plasmid or a control plasmid (Fig. 5F). Because inclusion of the test exons may induce nonsense-mediated decay (NMD) by inducing an in-frame premature termination codon (PTC), as is the case with *SRSF6*, we assayed splicing in the presence of the translation inhibitor emetine dihydrochloride, a potent inhibitor of NMD in vivo (Noensie and Dietz 2001; Lareau et al. 2007; Ni et al. 2007). As shown in Figure 5H, RT-PCR of wild-type *SRSF6* splicing reporters show expected decrease in exon inclusion in response to HNRNPA1 overexpression compared to endogenous levels (Fig. 5G). Deletion of the distal intronic U2AF2 binding site resulted in an attenuation of exon inclusion and a loss of HNRNPA1-dependent exon skipping. Quantification of amplicon ratios demonstrated that mutation of the putative HNRNPA1-induced, U2AF2-associated intronic elements disrupt splicing sensitivity to HNRNPA1 overexpression ( $P < 0.0029$ , respectively, ratio paired *t*-test) (Fig. 5I).

## Discussion

In this study, we have used iCLIP to study the impact of perturbation of alternative splicing factor levels on core components of the spliceosome. These experiments allow for quantitative analysis of competitive RNA binding and splice site recognition on a global scale. In cells overexpressing HNRNPA1, we observed a pronounced shift in U2AF2 binding sites relative to control cells. Based on U2AF2 peak distributions, this HNRNPA1-dependent redistribution involves loss of proximal-intron and recognition of distal-intronic peaks (Figs. 1, 2A–C). A similar pattern was

observed at the single nucleotide level, where U2AF2 crosslinking density near alternative 3' splice sites was reduced in HNRNPA1 overexpression cells and distributed across exons (Fig. 3). In addition to evidence for global changes in U2AF2 binding position, we also observed variation in sequence motifs enriched at U2AF2 peaks in both cell lines (Figs. 1, 2E). Splicing reporter assays revealed that upstream polypyrimidine tracks identified by iCLIP were involved in HNRNPA1-dependent splicing regulation (Fig. 5). These data also support the hypothesis that distant U2AF2 binding sites may function as splice site decoys that contribute to HNRNPA1-dependent splicing silencing.

HNRNPA1 represses splicing through diverse mechanisms. We think our data suggest that HNRNPA1 alters the competition between bona fide and decoy 3' splice sites. Because HNRNPA1 forms a ternary complex with the U2AF heterodimer and the 3' splice site (Tavanez et al. 2012), HNRNPA1-dependent redistribution of U2AF2 from bona fide splice sites to "decoy" sites might arise from direct competition for these splicing factors at 3' splice sites. Tavanez et al. (2012) also demonstrated that HNRNPA1 is required for discrimination of AG and CG dinucleotides at 3' splice sites. HNRNPA1 displaces U2AF from non-canonical CG-containing splice sites but allows it to productively engage AG-containing splice sites. We observed a global decrease in U2AF2 crosslinking near alternative 3' splice sites, but little change near constitutive splice sites. Given that alternative exons are typically flanked by weak splice sites, it seems unlikely that this proofreading mechanism contributes to regulation of alternative splicing. Tavanez et al. (2012) also noted that depletion of HNRNPA1 from HeLa cells resulted in increased U2AF association at spurious sites, including within the 3' UTRs of intronless messages when HNRNPA1 is knocked down. We also observed a global increase in U2AF2 crosslinking to 3' UTRs but not CDS regions when HNRNPA1 is overexpressed (Supplemental Table 3). It will be important to understand if RNA determinants (sequence, structure, splice site strength) distinguish between a proofreading function for HNRNPA1 and a simple competition with U2AF (Jain et al. 2017).

Our findings are well aligned with a recent census of U2AF2 binding sites in HeLa cells, which documented a position-dependent code for U2AF2 in splicing regulation (Shao et al. 2014). Perhaps most relevant to the work presented here is their observation that noncanonical U2AF2 binding sites located upstream or within alternative exons correlate with exon skipping (Lim et al. 2011; Shao et al. 2014; Cho et al. 2015). We find strong evidence for an HNRNPA1-dependent shift in U2AF2 binding sites from exon-proximal to -distal intronic positions. We have demonstrated that a noncanonical U2AF2 binding site in *SRSF6* intron 2 is required for HNRNPA1-dependent splicing regulation (Fig. 5E). Additionally, in the context of a splicing reporter, deletion of this element appears to dramatically reduce exon inclusion. It is possible that HNRNPA1-dependent eviction of U2AF2 at 3' splice sites could initiate remodeling at other functional RNA elements, such as nearby intronic splicing enhancers.

Our results suggest a role for HNRNPA1 in promoting recognition of noncanonical sites by U2AF2. One interesting observation is that although U2AF2 binding sites are predominantly pyrimidine-rich motifs (Singh et al. 1995), we observe variation in over-represented sequence motifs at different classes of binding sites across the transcriptome, suggesting that U2AF2 occupies low-affinity binding sites when HNRNPA1 is overexpressed (Fig. 2E). This result is consistent with the hypothesis that U2AF2 and HNRNPA1 compete for closely related sequences (Zhu et al. 2001; Okunola and Krainer 2009; Jain et al. 2017). Alternatively,

HNRNPA1 may modulate U2AF2 RNA binding specificity by altering the conformation of its RNA recognition motifs (Mackereth et al. 2011; Tavanez et al. 2012; Agrawal et al. 2016; Voithenberg et al. 2016). It is possible that the large-scale reorganization of U2AF2 binding to *Alu*-derived RNA sequences triggered by HNRNPA1 overexpression is an ultrasensitive response driven by molecular titration of U2AF-3' splice site interactions by HNRNPA1 (Zhang et al. 2013).

*Alu* elements influence gene expression in diverse ways (Hasler and Strub 2006; Chen and Carmichael 2009; Gong and Maquat 2011; Pastor and Pagani 2011; Kelley et al. 2014; Tajnik et al. 2015). Recently, Zarnack et al. (2013) demonstrated that HNRNPC competes with U2AF2 to repress inclusion of antisense *Alu*-derived exons in mRNA. We find that HNRNPA1 overexpression correlates with increased U2AF2 association with both sense and antisense *Alu*-derived RNA sequences. We did not observe any change in HNRNPC protein expression or localization (Supplemental Fig. 7). Taken together, our data demonstrate that *Alu*-derived sequences may function as RNA regulatory elements that respond to changes to the intracellular concentration of splicing factors. It is possible that, as primate-specific elements, *Alu* elements may influence the evolution of splicing regulation by modulating recognition of bona fide exons. Our results suggest the intriguing hypothesis that *Alu*-derived RNA elements may contribute to species-specific differences in alternative splicing throughout the primate lineage.

## Methods

### iCLIP analysis of U2AF2, SRSF1, and HNRNPA1

iCLIP was performed as previously described (Konig et al. 2010; Huppertz et al. 2014). Briefly, Flp-In T-REX HEK293T cells (Invitrogen) lacking or containing a stable, inducible T7-tagged version of HNRNPA1 were treated with tetracycline for 24 h and then irradiated with UV-C light to form covalent crosslinks between proteins and nucleic acids *in vivo*. After cell lysis, RNA was partially fragmented using low concentrations of micrococcal nuclease, and U2AF65<sup>-</sup>, SRSF1<sup>-</sup>, or HNRNPA1-RNA complexes were immunopurified with U2AF65, (MC3; Santa Cruz Biotechnology), SRSF1 (96; Santa Cruz Biotechnology), and HNRNPA1 (4B10; Santa Cruz Biotechnology) antibodies immobilized on protein A-coated magnetic beads (Life Technologies), respectively. After stringent washing and dephosphorylation (Fast AP, Fermentas), RNAs were ligated at their 3' ends with a preadenylated RNA adaptor (Bioo Scientific) and radioactively labeled to allow visualization. Samples were run using MOPS-based protein gel electrophoresis (in-house recipe) and transferred to a nitrocellulose membrane. Protein-RNA complexes migrating 15–80 kDa above free protein were cut from the membrane, and RNA was recovered from the membrane by proteinase K digestion under denaturing (3.5 M urea) conditions. The oligonucleotides for reverse transcription contained two inversely oriented adaptor regions adapted from the Bioo NEXTflex small RNA library preparation kit (Bioo Scientific), separated by a BamHI restriction site as well as a barcode region at their 5' end containing a 4-nt experiment-specific barcode within a 5-nt random barcode to mark individual cDNA molecules. cDNA molecules were size-purified using denaturing PAGE gel electrophoresis, circularized by CirLigase II (Epicenter), annealed to an oligonucleotide complementary to the restriction site, and cut using BamHI (NEB). Linearized cDNAs were then PCR-amplified using ImmoMix PCR master mix (Bioline) with primers (Bioo) complementary to the adaptor regions and were subjected to high-throughput sequencing using Illumina HiSeq.

A more detailed description of the iCLIP protocol has been published (Huppertz et al. 2014).

### Mapping and analysis of iCLIP sequencing data

Single-end reads generated by Illumina HiSeq were inspected for the presence of adaptor sequences. Reads containing sequences corresponding to the 3' RNA adaptor were retained if they were at least 30 bp long after the adaptor sequence was trimmed off. The first 9 bp in each read from the iCLIP library preparation, containing an internal barcode comprising 4 bp for replicate identification and 5 bp of random nucleotides for use in duplicate mapping removal, were also removed before mapping. Trimmed reads were checked for mapping to a repeat filter comprising RepeatMasker elements (Tarailo-Graovac and Chen 2009) in the human genome using Bowtie 2 (Langmead and Salzberg 2012). Reads that passed the repeat filter were mapped to the transcriptome and genome with TopHat2 (Kim et al. 2013). If reads mapped equally well to multiple loci, a single mapping was selected randomly by TopHat2. Duplicate mappings from each replicate were reduced to one per position if they had the same genomic endpoints and if they originated from reads with the same set of random 5-bp nucleotides. Following mapping and duplicate removal, individual reads were truncated to their 5' ends to represent the site of crosslinking consistent with the iCLIP methodology. For all samples, only such crosslinking sites found to have nonzero mapping counts in two out of three replicates (or two out of two duplicates where applicable) were considered to be biologically reproducible candidates for further analysis. The counts at such reproducible crosslinking sites were summed over all replicates to create an aggregated data set for each cell condition and CLIP. To determine the background from the iCLIP data sets, the two cell conditions (control and HNRNPA1-overexpressing) were temporarily further aggregated for each CLIP (U2AF, SF2, A1), and those binding sites that had nonzero counts in all three temporary aggregate data sets were determined. A 41-nt mask was created by extending 20 nt upstream of and 20 nt downstream from each such three-way common binding site. The aggregated data set of binding sites for each cell condition and CLIP was then filtered using this mask, keeping only sites outside the mask that also had a mapping count of at least three in the aggregate data. These aggregated and filtered data were used for downstream analyses. This aggregation and filtering strategy was adapted from previously described iCLIP analysis pipelines (Friedersdorf and Keene 2014; Flynn et al. 2015). For use as input to CLIPper (Lovci et al. 2013), the filtered (single nucleotide) binding sites were expanded by 15 nt upstream and 15 nt downstream.

CLIPper (CLIP-seq peak enrichment; <https://github.com/YeoLab/CLIPper>) was used to determine genomic distribution of RNA crosslinking peaks as well as identify clusters representing binding sites for HNRNPA1, U2AF2, and SRSF1 for each condition as previously described (Lovci et al. 2013). For each condition, the resulting iCLIP peak data of the replicates were merged. The peaks were annotated to the human genome (hg19) and then divided into categories based on their genomic locations, including CDS, intron, and UTR. The peaks in each category were further subsetted based on whether they overlapped with *Alu* elements. For the motif analysis, 50-bp sequences were extracted from the peak regions (crosslinking site  $\pm$  25 bp). Strand-specific MEME-ChIP and HOMER analyses were performed on these sequences to find 6–10 bp long, or 5-, 7-, 9-, and 11-bp enriched motifs (Bailey et al. 2009; Heinz et al. 2010).

For differential crosslinking analysis, CLIPper peaks from both control and HNRNPA1-overexpressing cells were merged using BEDTools intersect, and peak coverage by crosslink-centered

reads (i.e., reads constructed by expanding the 5' end upstream and downstream by 15 nt) was assessed for all replicates using BEDTools multicov (Quinlan and Hall 2010). Differential status was subsequently determined using DESeq2 (Love et al. 2014). Peaks were determined to be increasing (FDR < 0.05,  $\log_2(\text{fold-change}) > 1$ ) or decreasing (FDR < 0.05,  $\log_2(\text{fold-change}) < -1$ ) based on FDR and  $\log_2(\text{fold-change})$ . These categories were further subdivided based on overlap of *Alu* elements, 3'ss (i.e., the 200-nt intronic region proximal to the 3'ss), and other. Enriched motifs were identified as described above using HOMER (Heinz et al. 2010).

### RBP binding analysis

We extracted 40,769 cassette exons from MISO human genome (hg19) alternative events annotation version 2. We extracted 200,880 constitutive exons from RefSeq gene annotation by excluding the exons that overlap with cassette exons. Gene differential expression analysis was performed using edgeR (Robinson et al. 2010). We used 40,952 constitutive exons that were not significantly differentially expressed (FDR > 0.05) in further analysis.

For each RNA binding protein in each cell line, the iCLIP reads of all the replicates were merged together (Flynn et al. 2015). The start positions of the reads were considered as crosslinking sites. The number of reads near the 3'ss (100 bp into the intron, 50 bp into the exon) of each exon was calculated based on a 10-bp window. The raw read counts were normalized by the total library size.

The changes in binding of U2AF2 and SF2 near 3'ss were further analyzed with edgeR. Read counts were calculated for 200-bp intron regions near the 3'ss of the cassette exons. For each RBP, the regions with more than one count per million (CPM) in at least half of the replicates in either of the cell line were used for binding change analysis.

### RBP binding near *Alu* elements

We extracted 315,974 antisense *Alu* elements from RepeatMasker. The merged iCLIP data for each condition were down-sampled to 1M reads. The total number of sense strand reads were calculated for *Alu* and nearby regions (250 bp from *Alu* boundary). For each cassette exon events (cassette exon + up-/downstream introns + up-/downstream exons), the number of reads in antisense *Alu* elements, and the total number of reads in the whole event were calculated separately. The proportion of reads that fall into antisense *Alu* elements for each event was used to represent the RBP binding change in *Alu* regions.

### Randomization test for establishing significance of *Alu*-element overlap by distal intronic increasing peaks

U2AF2 iCLIP peaks exhibiting increasing binding following HNRNPA1 overexpression were extracted and subsetted for those found within distal intronic regions using a series of different distance constraints (minimum 200, 500, or 2000 bases from the nearest exon). Any intronic region found to overlap an alternative exon was removed from consideration (introns flanking the alternative exon were included). The peaks were then randomly shuffled within the same sets of distal intronic regions using BEDTools shuffle, and the number of shuffled peaks overlapping *Alu* elements was determined. The shuffling process was repeated 1000 times to construct a null distribution of expected *Alu* overlaps. Finally, the actual number of *Alu*-overlapping peaks was compared to the randomized null distribution. As the observed counts exceeded the entirety of the null distribution of counts for each definition of distal intron, the *P*-value has an upper bound of

0.001 for each comparison. The analysis was repeated for peaks found in corresponding proximal intronic regions and exons.

### mRNA-seq of control or HNRNPA1-overexpressing HEK293 cells

RNA was isolated from whole-cell lysates of control and HNRNPA1-overexpressing Flp-In T-REx HEK 293T cells using TRI-Reagent LS (Sigma). Poly(A)<sup>+</sup> sequencing libraries were prepared using the TrueSeq RNA library prep kit (Illumina). Each condition was analyzed in duplicate using the HiSeq 2000.

### Quantification of alternative splicing by RNA-seq

poly(A)<sup>+</sup> transcriptome sequencing reads were mapped to the human reference genome (hg19) with TopHat2. Mapped reads of duplicates were merged together for splicing analysis. Splicing change was analyzed with MISO (Katz et al. 2010). The MISO result was filtered with the following parameters: --num-inc 1 --num-exc 1 --num-sum-exc 10 --delta-psi 0.20 --bayes-factor 10. After filtering, 267 skipped exon events were left for further analysis. We chose to map our data to hg19 to facilitate analysis with the MISO alternative splicing event database, which is based on hg19 annotation. Because our study focuses on alternative splicing of protein-coding genes, remapping the data to GRCh38 will not have a significant impact on rigor or reproducibility.

### Mapping to an *Alu* consensus sequence

*Alu* element annotations were obtained from the hg19 UCSC Genome Browser RepeatMasker track. A strategy similar to the one described in Jacobs et al. (2014) was used to show the CLIPper peak density over all *Alu* elements as follows: After removal of the longest 2%, the top 50 longest human *Alu* sequences were aligned with MUSCLE and used to construct a consensus sequence (Edgar 2004). CLIPper peaks were mapped from the genomic position to the consensus sequence position using a BLAT alignment of the repeat to the consensus, and the coverage of summits per bp of the consensus *AluSc* was plotted in Supplemental Figure 5c (Kent 2002). A Genome Browser session displaying the Repeat Masker Data can be found at the following URL: [http://genome.ucsc.edu/cgi-bin/hgTracks?hgS\\_doOtherUser=submit&hgS\\_otherUserName=Max&hgS\\_otherUserSessionName=pubRepeats2Sanford](http://genome.ucsc.edu/cgi-bin/hgTracks?hgS_doOtherUser=submit&hgS_otherUserName=Max&hgS_otherUserSessionName=pubRepeats2Sanford).

### Data access

All iCLIP and RNA-seq data generated in this study have been submitted to the NCBI Gene Expression Omnibus GEO (GEO; <https://www.ncbi.nlm.nih.gov/geo/>) under accession number GSE83923. Access to other publicly available data sets from GEO used in this study is detailed in Supplemental Table S8.

### Acknowledgments

We thank Professors Javier Caceres (MRC HGU, Edinburgh), Manny Ares (UCSC), and Al Zahler (UCSC) for discussion of the project. We thank Julia Philipp (UCSC) for thoughtful comments on the manuscript. This project was supported by National Institutes of Health grants GM1090146 (National Institute of General Medical Sciences; J.R.S.), HG007336 (National Human Genome Research Institute; M.T.), U41HG002371 (National Human Genome Research Institute; M.H.), GM008646 (National Institute of General Medical Sciences; NIH Training Grant Support for J.M.D.), and California Institute of Regenerative Medicine grant GC1R-06673-A (J.R.S.).

**Author contributions:** J.M.H., H.L., Y.L., and J.R.S. designed the experiments. J.M.H., H.L., A.J.W., G.K., J.M.D., M.H., and S.K. performed the experiments and data analysis. J.M.H., H.L., A.J.W., G.K., J.M.D., M.H., Y.L., and J.R.S. wrote and edited the manuscript. All authors read and approved the final manuscript.

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Received August 13, 2017; accepted in revised form March 22, 2018.