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Genome sequencing of linezolid-resistant *Streptococcus pneumoniae* mutants reveals novel mechanisms of resistance

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Linezolid is a member of a novel class of antibiotics, with resistance already being reported. We used whole-genome sequencing on three independent *Streptococcus pneumoniae* strains made resistant to linezolid in vitro in a step-by-step fashion. Analysis of the genome assemblies revealed mutations in the 23S rRNA gene in all mutants including, notably, G2576T, a previously recognized resistance mutation. Mutations in an additional 31 genes were also found in at least one of the three sequenced genomes. We concentrated on three new mutations that were found in at least two independent mutants. All three mutations were experimentally confirmed to be involved in antibiotic resistance. Mutations upstream of the ABC transporter genes sprI02I and sprI887 were correlated with increased expression of these genes and neighboring genes of the same operon. Gene inactivation supported a role for these ABC transporters in resistance to linezolid and other antibiotics. The hypothetical protein sprO333 contains an RNA methyltransferase domain, and mutations within that domain were found in all *S. pneumoniae* linezolid-resistant strains. Primer extension experiments indicated that sprO333 methylates G2445 of the 23S rRNA and mutations in sprO333 abolished this methylation. Reintroduction of a nonmutated version of sprO333 in resistant bacteria reestablished G2445 methylation and led to cells being more sensitive to linezolid and other antibiotics. Interestingly, the sprO333 ortholog was also mutated in a linezolid-resistant clinical *Staphylococcus aureus* isolate. Whole-genome sequencing and comparative analyses of *S. pneumoniae* resistant isolates was useful for discovering novel resistance mutations.

[Supplemental material is available online at www.genome.org. The sequence data from this study have been submitted to GenBank under accession nos. ABZC00000000 and ABZT00000000.]

Linezolid contains an oxazolidinone ring and represents a novel class of synthetic antibiotics. Linezolid is highly effective against a number of clinically important gram-positive pathogens such as *Staphylococcus aureus* and its methicillin-resistant version (MRSA), *Streptococcus pneumoniae*, enterococci, and their vancomycin-resistant versions (VRE), and several others (for review, see Vara Prasad 2007). Linezolid binds to the 50S subunit of the bacterial ribosome via interactions with the central loop segment of domain V of the 23S rRNA to block the formation of protein synthesis initiation complexes. Recent cross-linking experiments have shown that oxazolidinone antibiotics interact with the A site of the bacterial ribosome and possibly interfere with the placement of the aminoacyl-tRNA (Leach et al. 2007). This has been further substantiated by the crystal structure of linezolid bound to the 50S ribosome subunit, where the antibiotic was suggested to perturb the correct positioning of tRNAs on the ribosome (Wilson et al. 2008).

S. pneumoniae is the most common human respiratory pathogen, causing mainly pneumonia, acute otitis media, and bacterial meningitis. This pathogen is also frequently involved in life-threatening infections that are being further aggravated by the appearance and spread of drug-resistance to several classes of antimicrobial agents. For example, a recent survey of U.S. isolates indicated that 25% of *S. pneumoniae* were resistant to at least two

antibiotics, and a third of these isolates were resistant to at least four antibiotics (Thornsberry et al. 2008), but all studies confirmed that linezolid is still very active against *S. pneumoniae* (Jones et al. 2007). However, it was shown that linezolid-resistant strains (minimal inhibitory concentration [MIC] \geq 8 μ g/mL) of *S. pneumoniae* can be generated in vitro (Carsenti-Dellamonica et al. 2005), that clinical *S. pneumoniae* strains not susceptible to linezolid (MIC \geq 4 μ g/mL) have been reported (Wolter et al. 2005), and that one resistant clinical isolate of *Streptococcus oralis* has been described (Mutmick et al. 2003). With the use of linezolid steadily increasing in the Western world, the potential for an increase in resistance to linezolid is likely to be inevitable.

Resistance to linezolid was described in the enterococci (Prystowsky et al. 2001) and the staphylococci (Tsiodras et al. 2001) and is usually due to point mutations in key genes. It was first studied in gram-positive pathogens selected for resistance in vitro, and mutations were noted in domain V of the 23S rRNA. Several mutations have been pinpointed in the 23S rRNA, but the most common mutation is G2576T when using the *Escherichia coli* numbering system (for review, see Meka and Gold 2004). The same mutations were also observed in clinical isolates of *S. aureus* (Tsiodras et al. 2001; Meka et al. 2004), *Staphylococcus epidermidis* (Hong et al. 2007; Kelly et al. 2008), and in enterococci (Gonzales et al. 2001; Sinclair et al. 2003; Bourgeois-Nicolaos et al. 2007) resistant to linezolid. There are four to six copies of the 23S rRNA genes in most gram-positive pathogens, and the level of resistance generally correlates with the number of mutated copies of 23S rRNA, as shown in the enterococci (Marshall et al. 2002; Ruggero

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et al. 2003) and in the staphylococci (Tsakris et al. 2007; Besier et al. 2008). Domain V of the 23S rRNA is also the binding site of other antibiotics such as chloramphenicol, and linezolid-resistant bacteria are consequently often cross-resistant to these other classes of antibiotics.

In addition to mutations in the 23S rRNA, a 6-bp deletion in the ribosomal protein L4 has been described in two clinical isolates of *S. pneumoniae* that were resistant to chloramphenicol and not susceptible to linezolid (Wolter et al. 2005). Recently, an integrated plasmid 23S rRNA methyltransferase coded by the *cfr* gene (for chloramphenicol-florfenicol resistance) was shown to confer resistance to linezolid in *S. aureus* (Toh et al. 2007; Mendes et al. 2008). There are also other linezolid-resistant strains (selected in vitro or clinical isolates) that do not seem to carry any of the main known mutations (Sander et al. 2002; Carsenti-Dellamonica et al. 2005; Richter et al. 2007); thus, it is possible that other mutations or the acquisition of additional genes may also contribute to linezolid resistance. Recently, with the emergence of new genome analysis technologies such as comparative genome sequencing (CGS) and massively parallel DNA sequencing techniques, whole-genome sequencing has become a powerful method to detect mutations linked to drug resistance or mode of action of antibiotics (Albert et al. 2005; Andries et al. 2005; Manjunatha et al. 2006; Mwangi et al. 2007). We report the use of whole-genome sequencing of three *S. pneumoniae* strains selected independently for in vitro linezolid resistance in a step-by-step fashion to find known (23S rRNA) and novel (RNA methyltransferase, ABC proteins) genes involved in resistance to this novel drug.

Results

Generation of mutants and cross-resistance

Two independent mutants for each of two *S. pneumoniae* strains (R6 and CCRI 1974) were generated by iterative selection for linezolid resistance in vitro. Cells were selected for progressive resistance to 1, 2, 4, 8, 16, and then 32 $\mu\text{g}/\text{mL}$ of linezolid. The MICs of the progenitor-susceptible isolates were 0.5 $\mu\text{g}/\text{mL}$ for R6 and 0.75 $\mu\text{g}/\text{mL}$ for CCRI 1974. The MICs of the most highly resistant isolates were 32 $\mu\text{g}/\text{mL}$ and were named R6M1 and R6M2 when derived from *S. pneumoniae* R6, and CCRI 1974M1 and 1974M2 when derived from *S. pneumoniae* CCRI 1974. These linezolid-resistant isolates were cross-resistant to chloramphenicol and florfenicol (Table 1), which are antibiotics that also target the 23S rRNA. The mutants were not cross-resistant to vancomycin or to erythromycin (Table 1). Vancomycin does not target the 23S

rRNA, and while erythromycin does interact with the 23S rRNA within the peptidyl transferase center, it is at a distinct site from linezolid (Schlunzen et al. 2001).

Whole-genome sequencing of *S. pneumoniae* linezolid-resistant strains

The two independent R6 mutants R6M1 and R6M2 were resequenced using the comparative genome sequencing (CGS) method developed by NimbleGen (<http://www.nimblegen.com/>). The CGS technique uses tiled DNA microarray hybridizations and rapidly surveys entire microbial genomes, identifying the locations of SNPs, insertions, or deletions. The mutations are further characterized by a subsequent step of array sequencing. These mutations were then confirmed by PCR sequencing of the regions suspected to be mutated. Both highly resistant R6 mutants R6M1 and R6M2 had a point mutation in each of their four 23S rRNA gene copies at position G2576T (Table 2). In addition to the 23S rRNA gene, CGS pinpointed nine additional mutations in the highly resistant mutant R6M1 and 10 additional mutations in R6M2 (Table 2). Six mutations in R6M1 were located in coding regions and were nonsynonymous. A single synonymous mutation (spr0525) and two deletions causing frameshifts were observed in the genes coding for hypothetical protein spr0333 and the ribosomal large subunit pseudouridine synthase D (spr1824) (Table 2). Nonsynonymous mutations in coding sequences were also the most prevalent in R6M2. Interestingly, two of these mutations were in the same open reading frame coding for the ABC protein spr1887. The only mutation found in noncoding sequences was also found 46 bp upstream of the spr1887 ATG (Table 2). A nonsense mutation in the argininosuccinate synthase gene spr0102 was detected by CGS in R6M2 (Table 2). Only two mutations were found in common between R6M1 and R6M2, which were the 23S rRNA gene copies and hypothetical protein spr0333. To extend this analysis and to test for the putative recurrence of resistance mutations to eventually find novel resistance mutations, we sequenced an independent linezolid-resistant isolate derived from the clinical isolate CCRI 1974. By multilocus sequence typing (MLST) analysis (see Methods), it was determined that 1974 was of the sequence type "124" while R6's sequence type was "595." Genome-wide assessments of SNPs as a marker of genetic diversity also confirmed that 1974 and R6 were different (see below).

Since the sequence of wild-type *S. pneumoniae* 1974 was not known, it was not possible to use the NimbleGen CGS technique. We instead used the massively parallel 454 Life Sciences (Roche) GS-FLX DNA sequencing platform to generate a 20 \times sequence coverage genome assembly for both the wild-type CCRI 1974 progenitor and the 1974M2 isolate. The draft genome assemblies are available at NCBI/DDBJ/EMBL/GenBank under the project accession numbers ABZC000000000 and ABZT000000000. A genome-wide alignment between R6 and 1974 indicated that the overall genome colinearity has been disrupted by a dispersed set of 18 insertions/deletions representing 4.2% of the CCRI 1974 genome. We used candidate SNPs as a further surrogate for divergence, and the CCRI 1974 strain genome assembly alignment comparisons generated 11,000 to 15,000 candidate SNPs when compared with R6, TIGR4, or other *S. pneumoniae* strains recently sequenced (Hiller et al. 2007). This number is similar to the 13,000 SNPs that we have measured between the R6 and TIGR4 genomes.

Comparative sequence analysis of 1974M2 and its sensitive parent revealed 25 mutations that were confirmed by PCR

Table 1. Susceptibility levels of *S. pneumoniae* isolates

<i>S. pneumoniae</i> strains	MIC ($\mu\text{g}/\text{mL}$)				
	LZD	CHL	FFC	ERY	VAN
R6 WT	0.5	3	1.6	0.064	0.38
R6M1	32	24	6	0.064	0.38
R6M2	32	32	12	0.047	0.38
1974WT	0.75	3	3.2	0.064	0.5
1974 M1	32	24	12	0.047	0.5
1974 M2	32	24	20	0.047	0.38

Average of at least three independent measures. (WT) Wild-type; (LZD) linezolid; (CHL) chloramphenicol; (FFC) florfenicol; (ERY) erythromycin; (VAN) vancomycin.

Table 2. List of mutations found in *S. pneumoniae* isolates selected for linezolid resistance

Name of gene	Locus name R6 ^a	<i>S. pneumoniae</i> strains			
		R6M1	R6M2	1974M2	1974M1 ^b
23S rDNA	spr_rnaD23S	G2576T	G2576T	T1743T A2503G	G2576T
23S rDNA	spr_rnaC23S	G2576T	G2576T	A1743T A2503G	G2576T
23S rDNA	spr_rnaB23S	G2576T	G2576T	A1743T A2503G	G2576G
23S rDNA	spr_rnaA23S	G2576T	G2576T	A1743T A2503G	G2576T
Hypothetical protein	spr0333	282–288 deletion	G623A C208Y ^c	G626T G209V	G503T S168I
ABC transporter, protein	spr1021			G-29T ^d	G-29T ^d
ABC transporter, protein	spr1021			C1419G H473Q	
ABC transporter, protein	spr1887		G-46T ^d	G-46T ^d	G-32T ^d
ABC transporter, protein	spr1887		G593A G198D G498T L166F		
Acyl-carrier-protein S-malonyltransferase	spr0380	G896T S297I			
ABC transporter, protein	spr0525	C234T R78R			
Phosphoenolpyruvate carboxylase	spr0947	A406C T136P			
Glycosyltransferase	spr0981	G347A G116E			
Hypothetical protein	spr1130	G1562A R521H			
Phosphoglucosamine mutase	spr1417	G887C R296P			
Aspartate aminotransferase	spr1808	A806G N269S			
Ribosomal large subunit pseudouridine synthase D	spr1824	C208 deletion			
Argininosuccinate synthase	spr0102		G116A W39stop		
Phosphotransferase system sugar-specific EII component	spr0421		C113A P38H		
ABC transporter, protein	spr1121		G514T D172Y		
Hypothetical protein	spr1195		G921A M307I		
Hypothetical protein	spr1316		G337A E113K		
Hypothetical protein	spr1465		C317T P106L		
Magnesium transporter	spr0170			G-3T ^d	
Competence-induced protein Ccs4	spr0182			T1380G N460K	
50S ribosomal protein L4	spr0189			G211A G71R	
Single-strand-DNA-specific exonuclease RecJ	spr0537			C1400T T467M	
Elongation factor Tu family protein	spr0598			A-52C ^d	
Spermidine synthase	spr0819			G146T G49V	
Translation initiation factor IF-3	spr0861			C454A H152N	
Hypothetical protein	spr1236			C277T Q93stop	
Putative oxidoreductase	spr1325			A393C E131D	
ABC transporter, substrate-binding protein	spr1534			C50A A17D	
Sialidase A precursor	spr1536			G430T D144Y	
Sucrose operon repressor	spr1569			G191T G64V	
Capsular polysaccharide biosynthesis protein	spr1654			C1032A H344Q	
Xanthine phosphoribosyltransferase	spr1662			C-216A ^d	

^aAll *S. pneumoniae* loci number (spr#) are according to the annotation of R6.

^bThe genome of this mutant was not sequenced. Only the genes found mutated in 1974M2 were PCR amplified and sequenced in 1974M1.

^cWhen the mutations are within coding regions, the change in amino acids is also indicated in italics.

^dIn noncoding sequence. The number preceded by “-” indicates the position upstream of the ATG.

amplification and conventional DNA sequencing. Sequence analysis highlighted mutations in the 23S rRNA genes of 1974M2, although they differed from the two R6 mutants, with mutations at position 1743 for three out of four 23S rRNA gene copies and at position 2503 for the four copies (Table 2). Eighteen additional mutations were found and confirmed in 1974M2 when compared with the wild-type sequence. Five of the 18 mutations were in noncoding regions. Interestingly, the same mutation (G to T) 46 bp upstream of the ABC gene spr1887 found in R6M2 was also present in 1974M2 (Table 2). The 13 other mutations were in coding regions, with 12 nonsynonymous substitutions and one nonsense mutation. The hypothetical protein spr0333 mutated in both R6 mutants was also mutated in 1974M2 (Table 2). We also PCR amplified and sequenced all the genes that were found to be mutated in 1974M2 in the independent mutant 1974M1. We found that the 23S rRNA gene was mutated in 1974M1 at the G2576T position in three of the four 23S rRNA gene copies (Table 2). Mutations in three additional genes (not necessarily at the same position as in 1974M2) were common between 1974M1 and

1974M2. One was in the coding region of the hypothetical protein spr0333, and the two other mutations were upstream of two operons, each encoding for ABC proteins (spr1021 and spr1887) (Table 2).

Mutations in known loci associated with linezolid resistance

The sequencing of three independent linezolid-resistant mutants and PCR analysis of targeted regions of a fourth mutant (1974M1) highlighted mutations in two genes already linked to linezolid resistance but also in 27 genes never previously associated with resistance. In the category of known mutations, the G2576 position within the domain V region of the 23S rRNA is central to linezolid resistance. In staphylococci and enterococci, the number of mutated copies of the 23S rRNA is generally correlated with linezolid MIC (see above), and this correlation was generally true for the four *S. pneumoniae* isolates studied in this report. In the R6M1 and R6M2 mutants, three copies of the 23S rRNA were mutated at position 2576 in cells with a MIC of 2 μg/mL, and

the fourth copy was mutated in cells resistant to 4 $\mu\text{g}/\text{mL}$ of linezolid (Supplemental Table 2). Three copies were also mutated at the same position in 1974M1 with a MIC of 2 $\mu\text{g}/\text{mL}$, but the fourth copy remained nonmutated even in the most resistant cells (Supplemental Table 2). The position G2576 was not mutated in 1974M2. Instead, positions 1743 and 2503 were mutated and three copies of the domain V 23S rRNA had the A2503G mutation at 16 $\mu\text{g}/\text{mL}$, while the fourth copy was mutated when cells reached a MIC of 32 $\mu\text{g}/\text{mL}$ (Supplemental Table 2).

Small deletions in the 50S ribosomal protein L4 were observed in clinical strains of *S. pneumoniae* with decreased susceptibility to linezolid (Wolter et al. 2005). It is salient to point out that in mutant 1974M2, whole-genome sequencing revealed that the L4 protein is mutated to G71R, a conserved region of L4.

ABC transporters and linezolid resistance

In addition to the known loci associated previously with linezolid resistance, we found 31 additional genes that were mutated in at least one of the three resistant strains that were sequenced. As an initial strategy to confirm the role of these novel mutations in resistance, we concentrated on genes that were mutated in at least two independent mutants. We reasoned that a recurrent mutation may increase the likelihood that it is associated with resistance, whereas a unique mutation may simply be a random replication error due to stress. Members of the ABC proteins can act as drug efflux pumps in some organisms (Ouellette et al. 1994), and five ABC protein genes were mutated in the linezolid-resistant mutants. This included *spr0525* in R6M1, *spr1534* in 1974M2, and *spr1121* in R6M2. However, mutations in both the upstream and coding regions of two ABC protein genes, *spr1021* and *spr1887*, were noted in three of the four linezolid-resistant isolates studied (Table 2). Consistent with our strategy of recurrent mutations, we attempted to assess the role of the latter two ABC genes in linezolid resistance. The mutations upstream of these two genes occurred early during the selection process in both 1974M2 and R6M2 but later in 1974M1 (Supplemental Table 2). Both genes are part of predicted operons (Fig. 1), and we used qRT-PCR to test whether a correlation existed between the mutation upstream of these

operons and their expression. The ABC *spr1021* gene is part of a three-gene operon (Fig. 1A), and the expression of the three genes was found to be increased seven- to 13-fold in the 1974M1 and 1974M2 mutants (Fig. 1A). The expression of these genes was not altered in the R6M2 that had no mutation upstream of this operon (Fig. 1A). The genes *spr1020* and *spr1024* were not overexpressed in 1974M2 (Fig. 1A), indicating that they are not part of the same operon as *spr1021*.

G to T transversions were found either at position 32 or 46 upstream of the operon containing *spr1887*. We similarly tested by qRT-PCR whether these mutations were correlated with increased expression of these genes. This was indeed the case, with a seven- to 13-fold increase in expression (Fig. 1B). The *spr1884* and *spr1888* genes were not overexpressed as determined by qRT-PCR (Fig. 1B).

The increased expression of these ABC protein genes may be linked to resistance, but since they are co-overexpressed with other genes, we assessed the role in resistance of each of these genes by gene inactivation in *S. pneumoniae*. The three genes of the *spr1021* operon (*spr1021*, an ABC protein; *spr1022*, a putative transcriptional regulator; *spr1023*, a putative transmembrane protein) were individually inactivated by insertional duplication mutagenesis using the nonreplicative plasmid pFF6 (Supplemental Table 1) containing a kanamycin resistance marker. Inactivation of the three genes in the wild-type background did not lead to any changes in susceptibility to linezolid (results not shown). However, when the three genes were inactivated independently in *S. pneumoniae* 1974M2, a phenotype of a threefold decrease in linezolid resistance was found in cells in which *spr1021* was inactivated (Table 3). No phenotype was observed in 1974M2 in which *spr1022* or *spr1023* was inactivated (not shown). Interestingly, cells in which *spr1021* was inactivated not only were more susceptible to linezolid but also were less cross-resistant to chloramphenicol and florfenicol (Table 3). We have attempted to overproduce *spr1021* (wild-type or mutated versions) in wild-type cells. Resistance was minimal and not significant when compared with controls (results not shown).

The *spr1887* gene is part of an operon containing two ABC protein genes (*spr1885* and *spr1887*). These ABC proteins, also known as PatA and PatB, have been linked to resistance to a number of drugs, including fluoroquinolones (Marrer et al. 2006b). This prompted us to test and subsequently verify that both the 1974M1 and 1974M2 mutants, with increased *patA* and *patB* expression (Fig. 1), were also cross-resistant to ciprofloxacin (Table 3). We inactivated both genes in both the CCRI1974 and in the 1974M2 background. In the wild-type background, we failed to show a decrease in susceptibility to linezolid (results not shown). However, a small but reproducible decrease in linezolid resistance was observed when any of the two genes was inactivated in the 1974M2 background (Table 3). Interestingly, resistance to several other drugs targeting the 23S rRNA was decreased significantly when either *spr1887* or *spr1885* were inactivated in 1974M2 (Table 3). Consistent with the known role of PatA and PatB in ciprofloxacin resistance

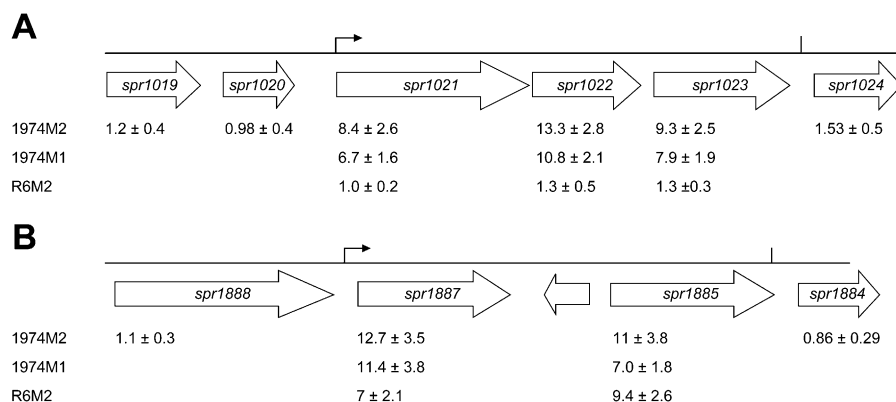


Figure 1. Monitoring mRNA expression by qRT-PCR in linezolid-resistant *Streptococcus pneumoniae*. Two genomic regions are shown, one containing the ABC protein gene *spr1021* (A) and one containing the ABC protein genes *spr1887* and *spr1885* (B). Between *spr1887* and *spr1885* there is a putative ORF, although it is not clear whether it is real. The *spr*# is according to the nomenclature of strain R6. The RNAs were normalized with 16S rRNA. The expression ratios shown are the cycle threshold (Ct) ratios between the resistant cells and the parent wild-type cells for each individual gene shown. Possible sites of transcription initiation (arrows) and termination (vertical bars) of the operons are indicated. Values represent an average of three independent experiments.

Table 3. Functional analysis of *S. pneumoniae* genes in antibiotic resistance

<i>S. pneumoniae</i> strains	Plasmid	MIC ($\mu\text{g/mL}$)			
		LZD	CHL	FFC	CIP
1974WT	—	0.75	3	3	0.25
1974M2	—	32	24	20	2
1974 M2	pJF_1021KO	12	12	8	1
1974 M2	pJF_1887KO	20	6	8	0.25
1974 M2	pJF_1885KO	24	8	8	0.38
1974 M1	pDL289	32	24	12	1.5
1974 M1	pJF_0333	12	12	6	1.5
R6 WT	—	0.5	3	1.6	0.38
R6 WT	pJF_0333KO	0.5	—	—	—
R6 M1	pDL289	32	—	—	—
R6 M1	pJF_0333	16	—	—	—

Average of at least three independent measures. (LZD) linezolid; (CHL) chloramphenicol; (FFC) florfenicol; (CIP) ciprofloxacin; (KO) knock-out construct.

(Marrer et al. 2006b), inactivation of either of these ABC genes led to a 10-fold decrease in ciprofloxacin resistance back to wild-type levels (Table 3).

RNA methyltransferase and linezolid resistance

Other than genes corresponding to the 23S rRNA, only one other gene was mutated in all four mutants, which corresponded to the hypothetical protein spr0333 (Table 2). Further analysis revealed that spr0333 corresponded to a protein of 385 amino acids containing PFAM domains for S-adenosylmethionine (SAM)-dependent methyltransferases (PFAM UPF0020). The spr0333 protein exhibited 32% sequence identity with a domain of the 702-amino-acid *E. coli* gene product YcbY, which encodes an RNA methyltransferase responsible for the N²-methylation of G2445 in the 23S rRNA (Lesnyak et al. 2006). Orthologs of spr0333 were found in many bacterial species, including several pathogens (Fig. 2). Mutations in spr0333 were at different positions and included nonsynonymous mutations and a frame-shift deletion (Table 2), but all were in the conserved SAM-methyltransferase domain. These mutations may render spr0333 inactive and may be linked to linezolid resistance. Methylation of ribosomal RNA is a frequent mechanism of acquired resistance to antibiotics such as erythromycin (for review, see Roberts 2008). It has recently been argued that one role of endogenous RNA methylases may be in 23S rRNA protection against natural xenobiotics (Toh and Mankin 2008). Therefore, we tested by primer extension whether spr0333 had 23S rRNA methylation activity, and in particular whether residue G2445 (*E. coli* numbering) was modified to m²G in the 23S rRNA of *S. pneumoniae*. N²-methylguanosine

blocks AMV reverse transcriptase during primer extension experiments, thus causing a stop one nucleotide before the modified guanosine. A reverse transcription stop was observed at position G2445 when the 23S rRNA template was extracted from the wild-type cells CCRI1974 (Fig. 3B, lane 1). Mutant 1974M1 had a point mutation leading to a mutated spr0333 protein with S168I (Table 2). When the 23S rRNA was derived from 1974M1 and used as a template in reverse transcription reactions, we did not observe a stop at position 2445 (Fig. 3B, lane 2). To ascertain that the lack of methylation as measured by primer extension was due to a mutated version of spr0333, we transformed pJF0333 (the spr0333 gene cloned into a shuttle vector replicating in *S. pneumoniae*, Supplemental Table 1) in the strain 1974M1. When the mutant was transformed with the empty vector, we still did not observe a stop at position 2445 (Fig. 3, lane 4), but in mutant cells transformed with pJF0333, cells gained back the methylation of ribonucleotide G2445 as shown by the stop of reverse transcription on the 23 S rRNA template (Fig. 3, lane 3).

The regain of methylation at position G2445 in 1974M1 was associated with an increased susceptibility to linezolid, but also to other antibiotics (Table 3). As susceptibility testing is simpler than primer extension, we also transformed pJF0333 in R6M1 (which had a 7-bp deletion in the spr0333 gene), and these cells were also shown to be more susceptible to linezolid (Table 3). The R6M2 with a C208Y mutation and 1974M2 with a G209V mutation in the spr0333 protein were also more susceptible to linezolid when these cells were transformed with pJF0333 (results not shown), demonstrating that different mutations in spr0333 are directly linked to linezolid resistance. Mutations in spr0333 were a relatively late event, being detected in cells resistant to either 16

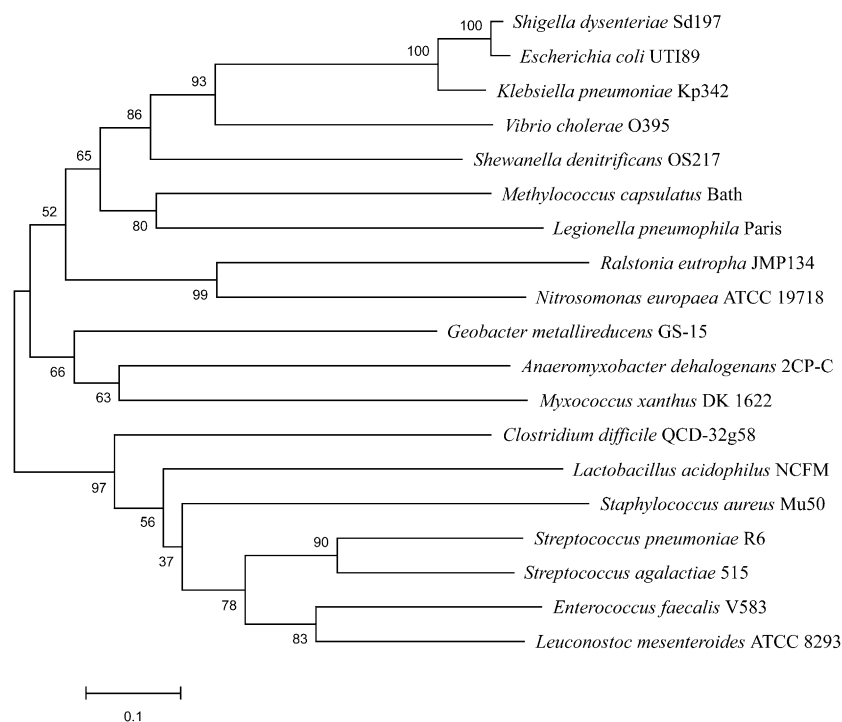


Figure 2. Phylogenetic analysis of a class of RNA methyltransferases related to spr0333. Amino acid sequences of the RNA methyltransferase proteins were aligned using the ClustalW algorithm, and evolutionary trees were constructed using the Neighbor-Joining method with Poisson correction as implemented in the MEGA3.1 package for the most conserved part of the proteins. The reliabilities of each branch point were assessed by the analysis of 1000 bootstrap replicates. A similar tree was obtained also when the whole protein was used for carrying the phylogenetic analysis (not shown).

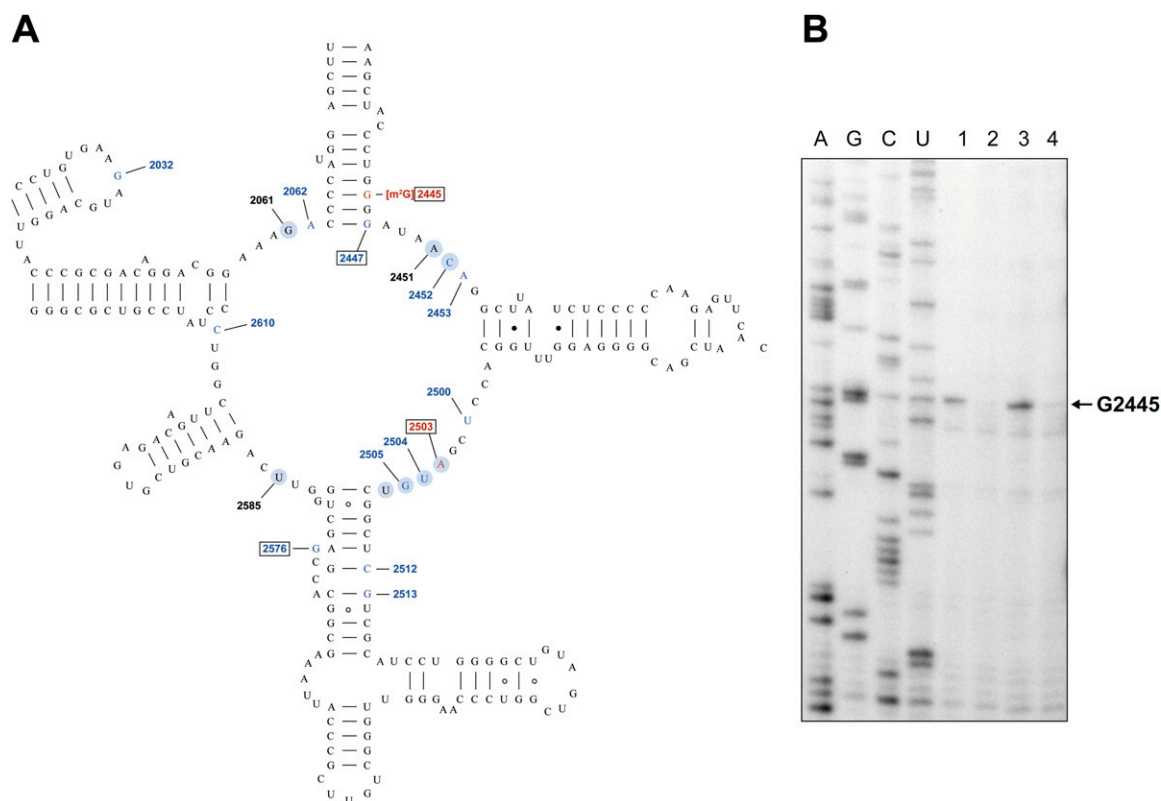


Figure 3. Mutations and modifications in the 23S rRNA related to linezolid resistance. (A) Secondary-structure model of part of the V domain of the 23S rRNA of *S. pneumoniae*. (Blue) Mutations in the 23S rRNA linked to linezolid resistance; (boxed residues) positions 2445, 2447, 2503, and 2576 found in this study; (red) new mutations or resistance mechanisms; (shaded) nucleotides that directly interact with linezolid as determined from several structural studies cited in the text. (B) Reverse transcription analysis of the *S. pneumoniae* 23S rRNA. U, A, C, G correspond with sequencing lanes. Primer extension results of the 23S rRNA from the control susceptible strain 1974 (lane 1), 1974M1 (lane 2), 1974M1 with pJF0333 (lane 3), 1974M1 with vector pDL289 (lane 4) are shown. The block (when present) in the reverse transcription reaction at position G2445 is indicated.

or 32 $\mu\text{g}/\text{mL}$ linezolid (Supplemental Table 2), suggesting that this mutation may occur after other mutations such as in the 23S rRNA. Consistent with this hypothesis, we did not observe a change in susceptibility when the spr0333 gene was inactivated in wild-type cells (Table 3).

Mutations in a chromosomally encoded RNA methylase and clinical resistance to linezolid

The complete genome sequencing of several isolates has pinpointed a novel linezolid resistance marker. There are few if any reports of *S. pneumoniae* linezolid-resistant isolates, but there have been reports of linezolid-resistant staphylococci and enterococci.

The staphylococci and the enterococci have a spr0333 ortholog (Fig. 2). In the NARSA collection (www.narsa.net) of vancomycin-intermediate MRSA (VISA) that we have studied in the past (Drummel-Smith et al. 2007), we confirmed that three isolates, NRS 119, 127, and 271, were linezolid-resistant (Table 4). Sequencing of the V region of the 23S rRNA gene of these isolates indicated the mutation G2576T in both NRS271 and NRS119, but a G2447T mutation for NRS127 (Table 4). The spr0333 ortholog in *S. aureus* is SAV1444 (nomenclature of *S. aureus* Mu50) (Kuroda et al. 2001), and it shares ~45% identity with the *S. pneumoniae* ortholog. In the isolate NRS119, which had the highest MIC against linezolid, the sequence of the SAV1444 gene revealed a 39-bp deletion in the SAM-methylation domain (Table 4). No mutations were observed

Table 4. Mutations in linezolid-resistant *Staphylococcus aureus* clinical isolates

<i>Staphylococcus aureus</i> strains	Linezolid MIC ($\mu\text{g}/\text{mL}$)	Multilocus sequence type	Mutations in SAV1444 (homolog of spr0333)	Mutation in 23S rDNA
Clinical isolates				
NRS127	1.5	ST5	No mutation	G2447T
NRS271	14	ST22	No mutation	G2576T ^a
NRS119	48	ST507	39-bp deletion (931–969)	G2576T ^a
Resistant mutants selected in vitro				
NRS127M1	128	ST5	No mutation	G2447T
NRS271M1	256	ST22	60-bp deletion (775–834)	G2576T ^a

^aThese mutations were also reported by Tsiodras et al. (2001) and Wilson et al. (2003).

in the two other VISA isolates that were less resistant to linezolid. NRS271 and NRS127 were selected in vitro for increased linezolid resistance, and their SAV1444 gene was resequenced. A 60-bp deletion was observed in the in vitro linezolid-selected NRS271 (Table 4).

Discussion

Complete bacterial genome sequencing is emerging as a cost-effective approach for identifying candidate genes and determining the mode of action and resistance to drugs (Albert et al. 2005; Andries et al. 2005; Fournier et al. 2006; Manjunatha et al. 2006; Mwangi et al. 2007; Howden et al. 2008). We used whole-genome sequencing of linezolid-resistant *S. pneumoniae* strains to find novel resistance mutations. Our strategy was based on sequencing several isolates selected for resistance in vitro. In vitro selected isolates were studied because (1) *S. pneumoniae* linezolid-resistant clinical isolates are not yet available; (2) the polymorphism between field isolates is such that it would be difficult to discriminate between resistance mutations and natural polymorphisms; (3) resistance to linezolid has been shown so far to be due mostly to point mutations that were shared between in vitro selected organisms and clinical isolates. Linezolid is a novel class of antibiotics to which resistance has only recently been described and involves primarily mutations in the 23S rRNA gene and most notably the mutation G2576T (for review, see Meka and Gold 2004). Indeed, this mutation was found in three out of the four *S. pneumoniae* (Table 2) and two out of three *S. aureus* (Table 4) strains resistant to linezolid. Resistant cells not having this key mutation usually have other mutations in the 23S rRNA gene (Fig. 3A), and this was also found to be the case for 1974M2 (Table 2) or NRS127 (Table 4). Mutation G2447T was previously linked to linezolid resistance (Xiong et al. 2000). The mutations A2503G and A1743T in 1974M2 are associated with linezolid resistance for the first time. A recent crystal structure of linezolid bound to the 50S ribosomal subunit has shown interaction between the antibiotic and A2503 (Leach et al. 2007; Ippolito et al. 2008; Wilson et al. 2008), and mutation of this residue was observed in some chloramphenicol-resistant mutants (Gregory et al. 2005). Interestingly, methylation of the A2503 residue by the acquired *cfr* gene also led to linezolid resistance (Toh et al. 2007). Mutation at G2447 was also correlated with resistance to several antibiotics, including chloramphenicol (Gregory et al. 2005), tiamulin (Pringle et al. 2004), and anisomycin (Blaha et al. 2008) in addition to linezolid (this study) (Xiong et al. 2000). The A1743 residue is not in the V region of the 23S rRNA, and further work would be required to test whether it is involved in linezolid resistance.

The sequencing of several independent mutants from genetically different backgrounds has allowed the identification of independently recurring mutations. We hypothesized that recurrent mutations in independent mutants increase the likelihood that a particular gene is involved in resistance. In addition to the 23S rRNA, few mutations were common to at least two independent mutants. ABC proteins are involved in multidrug resistance in several microbial eukaryotic pathogens (Ouellette and Légaré 2002), but also in bacteria (Lubelski et al. 2007), and mutations within putative promoter regions of operons coding for ABC proteins were observed in more than one mutant (Fig. 1).

S. pneumoniae encodes 14 ABC proteins, and disruption of 13 of them was possible. However, the disruption of only two ABC genes, spr1887 (*patA*) and spr1885 (*patB*), was shown to lead to

more susceptible bacteria (Robertson et al. 2005). The expression of these two genes was also shown to be up-regulated in cells treated with fluoroquinolones (Marrer et al. 2006a). The spr1887 and spr1885 proteins are the closest homologs of the well-studied multidrug-resistant proteins LmrC and LmrD of *Lactococcus lactis* (Lubelski et al. 2007). The increased expression of their genes in linezolid-resistant mutants (Fig. 1) indicated a direct involvement in linezolid resistance. Our gene disruption experiments of resistant cells provided evidence that PatA and PatB are associated with resistance to linezolid, although their role seems to be minor (Table 3). It has been suggested that they form heterodimers (Lubelski et al. 2007), and inactivation of both genes in the same cell may lead to further resistance. In addition to their role in linezolid resistance, analysis of the phenotype of either *patA* or *patB* disruptants clearly implicate the two ABC proteins in a multidrug resistance phenotype due to increased RNA expression (Fig. 1; Table 3).

Inactivation of spr1021 in a wild-type context did not lead to decreased susceptibility to drugs (Table 3), confirming the results of others (Robertson et al. 2005). However, the disruption of the same ABC gene in the linezolid-resistant background 1974M2 led to increased susceptibility to linezolid and also to chloramphenicol and florfenicol (Table 3). Since the expression of the mutated gene was increased by sevenfold, inactivation of the gene may explain the gain in sensitivity of the mutant. Indeed, a similar observation was noted for the major facilitator efflux pump PmrA in *S. pneumoniae*, where inactivation in wild-type cells did not increase susceptibility (Robertson et al. 2005) but did render resistant cells more susceptible to drugs (Gill et al. 1999). Alternatively, mutations in primary targets (23S rRNA) may lead to an increased concentration of free unbound linezolid that needs to be extruded by an efflux system. Disruption of the latter system would lead to the decreased resistance as observed (Table 3). The work highlights how mutations in putative promoter regions can increase the expression of efflux pumps (PatA, PatB, spr1021), hence contributing to resistance to clinically useful drugs including linezolid.

In addition to describing a novel role for ABC proteins in linezolid resistance, our complete genome sequencing approach has led to the identification of a novel resistance mechanism and to finding a cellular function for a hypothetical protein of *S. pneumoniae*. The spr0333 protein corresponds to an rRNA methyltransferase modifying the G2445 residue. This protein is only distantly related to the RlmL (YcbY) protein of *E. coli* (Fig. 2), which modifies the G2445 residue of the 23S rRNA in *E. coli* (Lesnyak et al. 2006). Inactivation of the *rlmL* gene in *E. coli* did not lead to changes in drug susceptibilities including linezolid (Toh and Mankin 2008), and, similarly, inactivation of spr0333 in susceptible *S. pneumoniae* R6 also did not change resistance to linezolid (Table 3). However, in all *S. pneumoniae* cells highly resistant to linezolid, spr0333 was mutated (Table 2), and this was even seen in one *S. aureus* clinical isolate (Table 4). The role of spr0333 in 23S rRNA methylation was tested experimentally (Fig. 3). Transformation of spr0333 in resistant cells rescued the methylation defect at position G2445 and restored susceptibility to linezolid but also to other antibiotics (Table 3). Thus, the loss of methylation is clearly involved in resistance but first requires other mutations, most likely in the 23S rRNA (e.g., position G2576T). Indeed, mutation in spr0333 occurred late during the selection process (Supplemental Table 2), and we propose a hierarchy in mutation selection. It is possible that some specific mutations would be required before the methylation status at position 2445 would be implicated in linezolid resistance. Indeed,

it was intriguing that we could not detect a mutation in spr0333 while selecting for increased resistance in *S. aureus* NRS127, which had another mutation close to position 2445 in the 23S rRNA (G2447) (Table 4). This would suggest the possibility of additional resistance mechanisms.

While we concentrated on recurrent mutations, the sequence analysis has revealed other strain-specific mutations, several of which could also contribute to resistance to linezolid. They are certainly worth investigating in additional studies, but it is possible that some of these mutations, despite further experimentation, may not be directly involved in the resistance phenotype. Two sequencing techniques were used in this study: microarray hybridizations and massively parallel DNA sequencing (MPS). Although we used a 454 Life Sciences (Roche) system, and other technologies are available (Illumina GAII and ABI SOLiD), de novo sequencing is still a sizable investment; however, costs are decreasing and it is becoming a routine technique (Kahvejian et al. 2008). Resequencing (microarray-based or low-coverage MPS) is less expensive, but the requirement of a prior reference sequence limits the species that can be analyzed. However, with improved bioinformatic tools and increasingly longer read lengths, including paired ends, it will likely become routine also to sequence de novo bacterial isolates using MPS short-read systems (Hernandez et al. 2008; Holt et al. 2008). This cost-effective de novo sequencing will allow the sequencing of clinical isolates. While several natural polymorphisms would be detected (Joyce et al. 2002), recurrence of a mutation in independent clinical isolates would provide a strong argument to pursue the more labor-intensive biological validation. This could prevent one of the main limitations in our current approach that requires a sequenced parent strain. These approaches are already leading to novel factors or concepts in bacterial virulence (Fouts et al. 2005; Gill et al. 2005; Highlander et al. 2007; Holt et al. 2008; Kennedy et al. 2008) and as suggested (Fournier et al. 2007; Francois et al. 2007; Kahvejian et al. 2008) will certainly contribute to a better understanding of the mutations and adaptations required to develop and maintain resistance. This study and other ongoing studies in the laboratory are certainly in agreement with the premise that whole-genome sequencing will be useful for understanding resistance mechanisms.

Methods

Bacterial strains, growth conditions, and MIC determinations

Key strains are listed in Supplemental Table 1. *S. pneumoniae* R6, whose genome sequence has been determined (Hoskins et al. 2001), and one clinical isolate, *S. pneumoniae* CCRI 1974, were chosen for the laboratory-induced development of resistance to linezolid. Pneumococci were grown in brain heart infusion broth (BHI, Difco) supplemented with 0.5% yeast extract, or in blood agar containing 5% defibrinated sheep's blood as described previously (Munoz et al. 1992). Cultures were incubated for 16–24 h in a 5% CO₂ atmosphere at 37°C. In order to select resistant cells, we subcultured R6 or CCRI 1974 in medium containing increasing concentration gradients of linezolid as described previously for other antibiotics (Martineau et al. 2000; Soualhine et al. 2005). Mutants were selected in a stepwise fashion by picking colonies growing at the highest antibiotic concentration at each stage. Six selection cycles were required to obtain two resistant mutants (M1 and M2) for each strain of *S. pneumoniae*. Minimal inhibitory concentration (MIC) to linezolid was determined by the E-test or the macrodilution method according to the guidelines of the

Clinical and Laboratory Standards Institute (CLSI). For the E-test, colonies from a fresh overnight plate were resuspended in phosphate saline buffer (PBS) to 0.5 McFarland units. The suspension was layered on Mueller–Hinton agar plates and then overlaid with E-test strips (AB Biodisk). For macrodilution, cation-adjusted Mueller–Hinton broth with lysed horse blood containing twofold concentration increments of linezolid was inoculated with a 5×10^5 cfu/mL bacterial suspension. The MIC was recorded as the lowest dilution showing no growth. All measurements were done at least in triplicate after 12–16 h of incubation, depending on the strain.

Multilocus sequence typing (MLST)

The MLST analysis was performed as described at <http://www.mlst.net>. The *aroE*, *gdh*, *gki*, *recP*, *spi*, *xpt*, and *ddl* genes for the *S. pneumoniae* strains and the *arcC*, *aroE*, *glpF*, *gmk*, *pta*, *tpi*, and *yqiL* genes for the *S. aureus* strains were PCR amplified and sequenced. The sequence type (ST) of strains was obtained from the MLST database on the basis of the resulting allelic profile.

Whole-genome sequencing

Genomic DNAs were prepared from mid-log phase cultures of *S. pneumoniae* strains using the Wizard Genomic DNA Purification Kit (Promega) according to the manufacturer's instructions. Mutants R6M1 and R6M2 were sequenced using the NimbleGen whole-genome sequencing approach (<http://www.nimblegen.com/>). Briefly, DNA of each mutant isolate and the DNA of the progenitor isolate (which were differentially labeled with fluorescent markers) were co-hybridized on DNA tiling microarrays. Regions hybridizing differently were sequenced by a second round of sequencing hybridization arrays. The sequencing of both mutants and their analyses were performed by NimbleGen (Albert et al. 2005). The CCRI 1974 wild-type strain and the CCRI 1974 M2 mutant were sequenced by the massively parallel sequencing using the 454 Life Sciences (Roche) GS-FLX system. Genome sequencing, assemblies, and comparative analyses were performed at the McGill University and Génome Québec Innovation Center. The draft genome assemblies for CCRI 1974 and CCRI 1974 M2 have been submitted to NCBI/DDBJ/EMBL/GenBank under the project accession numbers ABZC000000000 and ABZT000000000. The versions described in this study are the first versions, ABZC01000000 and ABZT01000000. The CCRI 1974 shotgun sequencing led to an assembly of 121 contigs with an aggregate genome size of 2,000,683 bp. The 1974M2 isolate sequencing project produced an assembly of similar quality (130 contigs, 1,998,515 bp). Whenever possible, the order and orientation of assembled contigs was done in accordance with the genome assembly of the *S. pneumoniae* TIGR4 isolate (accession number NC_003028). Mutations, deduced either from array hybridizations or massively parallel sequencing, were confirmed by PCR amplification and conventional DNA sequencing. The primers were designed on the basis of the published *S. pneumoniae* TIGR4 strain (Tettelin et al. 2001). Specific primers targeting the four individual 23S rRNA genes were designed.

RNA isolation and qRT-PCR

Wild-type and linezolid-resistant mutant *S. pneumoniae* strains were grown in BHI in the absence of antibiotics. Bacterial cells were harvested for RNA isolation during the logarithmic growth phase at an OD_{600nm} of 0.35–0.45. Total RNAs from sensitive and resistant derivatives were isolated using the Qiagen RNeasy Mini

Kit (Qiagen) according to the manufacturer's instructions. Contaminating DNA was digested with DNase I (Ambion). The quality and integrity of the starting RNA material were assessed with an Agilent Technologies 2100 BioAnalyzer using the RNA 6000 Nano LabChIP kit (Agilent). The cDNAs were generated from total RNAs using a random hexamer primer following the protocol for Superscript II (Invitrogen). Real-time quantitative RT-PCR assays were carried out in a BioRad Cycler using SYBR Green I (Molecular Probes). The reactions were carried out in a final volume of 20 μ L containing specific primers, and iQ SYBR Green Supermix (BioRad). All real-time RT-PCR data were normalized with the real-time amplification of the 16S rRNA. The expression data are shown relative to the data for the wild-type strain.

DNA constructs

The genetic constructs used in this study are described in Supplemental Table 1. Gene inactivation was done by insertional duplication mutagenesis using the nonreplicative plasmid pFF6, a pGEMT Easy (Promega) derivative where the ampicillin resistance marker was replaced by the kanamycin resistance gene marker of pDL289 (Buckley et al. 1995), which is expressed through the *amiC* promoter (F Fani, J Feng, and M Ouellette, unpubl.). PCR fragments of the selected genes were derived from strain *S. pneumoniae* R6 or CCRI 1974 and cloned into the multiple cloning sites of pFF6. For episomal expression, the entire coding sequences of genes of interest were PCR amplified with primers containing BamHI and XbaI and cloned into the *S. pneumoniae*-*E. coli* shuttle vector pDL289 (Buckley et al. 1995), a kind gift of D. Cvitkovitch (University of Toronto). The plasmid DNAs (2 μ g) were introduced in *S. pneumoniae* in which competence was induced using a competence peptide as described (Soualhiine et al. 2005). Transformants were selected in media containing 500 μ g/mL kanamycin. Proper integration leading to gene inactivation was confirmed by PCR.

Primer extension analysis

The primer (5'-TACAGCCCCAGGATGCGAC-3'), corresponding to the reverse complement of nucleotides 2512–2531 of *S. pneumoniae* 23S rRNA, was [³²P]-labeled. Total *S. pneumoniae* RNA (1 μ g) and the labeled primer were annealed by heating the tubes at 60°C followed by cooling for 10 min at room temperature. The AMV reverse transcriptase (Promega) was added and incubated for 30 min at 42°C. The cDNA products of the primer extension reactions were separated on 8% polyacrylamide sequencing gels. The gels were transferred to Whatman paper, dried, and revealed using autoradiography.

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References

- Albert TJ, Dailidene D, Dailide G, Norton JE, Kalia A, Richmond TA, Molla M, Singh J, Green RD, Berg DE. 2005. Mutation discovery in bacterial genomes: Metronidazole resistance in *Helicobacter pylori*. *Nat Methods* **2**: 951–953.
- Andries K, Verhasselt P, Guillemont J, Gohlmann HW, Neefs JM, Winkler H, Van Gestel J, Timmerman P, Zhu M, Lee E, et al. 2005. A diarylquinoline drug active on the ATP synthase of *Mycobacterium tuberculosis*. *Science* **307**: 223–227.
- Besier S, Ludwig A, Zander J, Brade V, Wichelhaus TA. 2008. Linezolid resistance in *Staphylococcus aureus*: Gene dosage effect, stability, fitness costs, and cross-resistances. *Antimicrob Agents Chemother* **52**: 1570–1572.
- Blahe G, Gurel G, Schroeder SJ, Moore PB, Steitz TA. 2008. Mutations outside the anisomycin-binding site can make ribosomes drug-resistant. *J Mol Biol* **379**: 505–519.
- Bourgeois-Nicolaos N, Massias L, Couson B, Butel MJ, Andremont A, Doucet-Populaire F. 2007. Dose dependence of emergence of resistance to linezolid in *Enterococcus faecalis* in vivo. *J Infect Dis* **195**: 1480–1488.
- Buckley ND, Lee LN, LeBlanc DJ. 1995. Use of a novel mobilizable vector to inactivate the *scrA* gene of *Streptococcus sobrinus* by allelic replacement. *J Bacteriol* **177**: 5028–5034.
- Carsenti-Dellamonica H, Galimand M, Vandenbos E, Pradier C, Roger PM, Dunais B, Sabah M, Mancini G, Dellamonica P. 2005. In vitro selection of mutants of *Streptococcus pneumoniae* resistant to macrolides and linezolid: Relationship with susceptibility to penicillin G or macrolides. *J Antimicrob Chemother* **56**: 633–642.
- Drummelsmith J, Winstall E, Bergeron MG, Poirier GG, Ouellette M. 2007. Comparative proteomics analyses reveal a potential biomarker for the detection of vancomycin-intermediate *Staphylococcus aureus* strains. *J Proteome Res* **6**: 4690–4702.
- Fournier PE, Vallenet D, Barbe V, Audic S, Ogata H, Poirel L, Richet H, Robert C, Mangenot S, Abergel C, et al. 2006. Comparative genomics of multidrug resistance in *Acinetobacter baumannii*. *PLoS Genet* **2**: e7. doi: 10.1371/journal.pgen.0020007.
- Fournier PE, Drancourt M, Raoult D. 2007. Bacterial genome sequencing and its use in infectious diseases. *Lancet Infect Dis* **7**: 711–723.
- Fouts DE, Mongodin EF, Mandrell RE, Miller WG, Rasko DA, Ravel J, Brinkac LM, DeBoy RT, Parker CT, Daugherty SC, et al. 2005. Major structural differences and novel potential virulence mechanisms from the genomes of multiple *Campylobacter* species. *PLoS Biol* **3**: e15. doi: 10.1371/journal.pbio.0030015.
- Francois P, Hernandez D, Schrenzel J. 2007. Genome content determination in methicillin-resistant *Staphylococcus aureus*. *Future Microbiol* **2**: 187–198.
- Gill MJ, Brenwald NP, Wise R. 1999. Identification of an efflux pump gene, *pmrA*, associated with fluoroquinolone resistance in *Streptococcus pneumoniae*. *Antimicrob Agents Chemother* **43**: 187–189.
- Gill SR, Fouts DE, Archer GL, Mongodin EF, Deboy RT, Ravel J, Paulsen IT, Kolonay JF, Brinkac L, Beanan M, et al. 2005. Insights on evolution of virulence and resistance from the complete genome analysis of an early methicillin-resistant *Staphylococcus aureus* strain and a biofilm-producing methicillin-resistant *Staphylococcus epidermidis* strain. *J Bacteriol* **187**: 2426–2438.
- Gonzales RD, Schreckenberger PC, Graham MB, Kelkar S, DenBesten K, Quinn JP. 2001. Infections due to vancomycin-resistant *Enterococcus faecium* resistant to linezolid. *Lancet* **357**: 1179.
- Gregory ST, Carr JF, Rodriguez-Correa D, Dahlberg AE. 2005. Mutational analysis of 16S and 23S rRNA genes of *Thermus thermophilus*. *J Bacteriol* **187**: 4804–4812.
- Hernandez D, Francois P, Farinelli L, Osteras M, Schrenzel J. 2008. De novo bacterial genome sequencing: Millions of very short reads assembled on a desktop computer. *Genome Res* **18**: 802–809.
- Highlander SK, Hulten KG, Qin X, Jiang H, Yerrapragada S, Mason EO Jr, Shang Y, Williams TM, Fortunov RM, Liu Y, et al. 2007. Subtle genetic changes enhance virulence of methicillin resistant and sensitive *Staphylococcus aureus*. *BMC Microbiol* **7**: 99. doi: 10.1186/1471-2180-7-99.
- Hiller NL, Janto B, Hogg JS, Boissy R, Yu S, Powell E, Keefe R, Ehrlich NE, Shen K, Hayes J, et al. 2007. Comparative genomic analyses of seventeen *Streptococcus pneumoniae* strains: Insights into the pneumococcal supragenome. *J Bacteriol* **189**: 8186–8195.
- Holt KE, Parkhill J, Mazzoni CJ, Roumagnac P, Weill FX, Goodhead I, Rance R, Baker S, Maskell DJ, Wain J, et al. 2008. High-throughput sequencing provides insights into genome variation and evolution in *Salmonella typhi*. *Nat Genet* **40**: 987–993.
- Hong T, Li X, Wang J, Sloan C, Cicogna C. 2007. Sequential linezolid-resistant *Staphylococcus epidermidis* isolates with G2576T mutation. *J Clin Microbiol* **45**: 3277–3280.
- Hoskins J, Alborn WE Jr, Arnold J, Blaszcak LC, Burgett S, DeHoff BS, Estrem ST, Fritz L, Fu DJ, Fuller W, et al. 2001. Genome of the bacterium *Streptococcus pneumoniae* strain R6. *J Bacteriol* **183**: 5709–5717.

- Howden BP, Stinear TP, Allen DL, Johnson PD, Ward PB, Davies JK. 2008. Genomic analysis reveals a point mutation in the two-component sensor gene *graS* that leads to intermediate vancomycin resistance in clinical *Staphylococcus aureus*. *Antimicrob Agents Chemother* **52**: 3755–3762.
- Ippolito JA, Kanyo ZF, Wang D, Franceschi FJ, Moore PB, Steitz TA, Duffy EM. 2008. Crystal structure of the oxazolidinone antibiotic linezolid bound to the 50S ribosomal subunit. *J Med Chem* **51**: 3353–3356.
- Jones RN, Fritsche TR, Sader HS, Ross JE. 2007. Zyvox annual appraisal of potency and spectrum program results for 2006: An activity and spectrum analysis of linezolid using clinical isolates from 16 countries. *Diagn Microbiol Infect Dis* **59**: 199–209.
- Joyce EA, Chan K, Salama NR, Falkow S. 2002. Redefining bacterial populations: A post-genomic reformation. *Nat Rev Genet* **3**: 462–473.
- Kahvejian A, Quackenbush J, Thompson JF. 2008. What would you do if you could sequence everything? *Nat Biotechnol* **26**: 1125–1133.
- Kelly S, Collins J, Maguire M, Gowing C, Flanagan M, Donnelly M, Murphy PG. 2008. An outbreak of colonization with linezolid-resistant *Staphylococcus epidermidis* in an intensive therapy unit. *J Antimicrob Chemother* **61**: 901–907.
- Kennedy AD, Otto M, Braughton KR, Whitney AR, Chen L, Mathema B, Mediavilla JR, Byrne KA, Parkins LD, Tenover FC, et al. 2008. Epidemic community-associated methicillin-resistant *Staphylococcus aureus*: Recent clonal expansion and diversification. *Proc Natl Acad Sci* **105**: 1327–1332.
- Kuroda M, Ohta T, Uchiyama I, Baba T, Yuzawa H, Kobayashi I, Cui L, Oguchi A, Aoki K, Nagai Y, et al. 2001. Whole genome sequencing of methicillin-resistant *Staphylococcus aureus*. *Lancet* **357**: 1225–1240.
- Leach KL, Swaney SM, Colca JR, McDonald WG, Blinn JR, Thomasco LM, Gadwood RC, Shinabarger D, Xiong L, Mankin AS. 2007. The site of action of oxazolidinone antibiotics in living bacteria and in human mitochondria. *Mol Cell* **26**: 393–402.
- Lesnyak DV, Sergiev PV, Bogdanov AA, Dontsova OA. 2006. Identification of *Escherichia coli* m2G methyltransferases: I. The *ycbY* gene encodes a methyltransferase specific for G2445 of the 23S rRNA. *J Mol Biol* **364**: 20–25.
- Lubelski J, Konings WN, Driessen AJ. 2007. Distribution and physiology of ABC-type transporters contributing to multidrug resistance in bacteria. *Microbiol Mol Biol Rev* **71**: 463–476.
- Manjunatha UH, Boshoff H, Dowd CS, Zhang L, Albert TJ, Norton JE, Daniels L, Dick T, Pang SS, Barry CE III. 2006. Identification of a nitroimidazo-oxazine-specific protein involved in PA-824 resistance in *Mycobacterium tuberculosis*. *Proc Natl Acad Sci* **103**: 431–436.
- Marrer E, Satoh AT, Johnson MM, Piddock LJ, Page MG. 2006a. Global transcriptome analysis of the virulence of a fluoroquinolone-resistant *Streptococcus pneumoniae* mutant and its parent to ciprofloxacin. *Antimicrob Agents Chemother* **50**: 269–278.
- Marrer E, Schad K, Satoh AT, Page MG, Johnson MM, Piddock LJ. 2006b. Involvement of the putative ATP-dependent efflux proteins PatA and PatB in fluoroquinolone resistance of a multidrug-resistant mutant of *Streptococcus pneumoniae*. *Antimicrob Agents Chemother* **50**: 685–693.
- Marshall SH, Donskey CJ, Hutton-Thomas R, Salata RA, Rice LB. 2002. Gene dosage and linezolid resistance in *Enterococcus faecium* and *Enterococcus faecalis*. *Antimicrob Agents Chemother* **46**: 3334–3336.
- Martineau F, Picard FJ, Lansac N, Menard C, Roy PH, Ouellette M, Bergeron MG. 2000. Correlation between the resistance genotype determined by multiplex PCR assays and the antibiotic susceptibility patterns of *Staphylococcus aureus* and *Staphylococcus epidermidis*. *Antimicrob Agents Chemother* **44**: 231–238.
- Meka VG, Gold HS. 2004. Antimicrobial resistance to linezolid. *Clin Infect Dis* **39**: 1010–1015.
- Meka VG, Pillai SK, Sakoulas G, Wennersten C, Venkataraman L, DeGirolami PC, Eliopoulos GM, Moellering RC Jr, Gold HS. 2004. Linezolid resistance in sequential *Staphylococcus aureus* isolates associated with a T2500A mutation in the 23S rRNA gene and loss of a single copy of rRNA. *J Infect Dis* **190**: 311–317.
- Mendes RE, Deshpande LM, Castanheira M, Dipersio J, Saubolle M, Jones RN. 2008. First report of *cfi*-mediated resistance to linezolid in human staphylococcal clinical isolates recovered in the United States. *Antimicrob Agents Chemother* **52**: 2244–2246.
- Munoz R, Dowson CG, Daniels M, Coffey TJ, Martin C, Hakenbeck R, Spratt BG. 1992. Genetics of resistance to third-generation cephalosporins in clinical isolates of *Streptococcus pneumoniae*. *Mol Microbiol* **6**: 2461–2465.
- Mutnick AH, Enne V, Jones RN. 2003. Linezolid resistance since 2001: SENTRY Antimicrobial Surveillance Program. *Ann Pharmacother* **37**: 769–774.
- Mwangi MM, Wu SW, Zhou Y, Sieradzki K, de Lencastre H, Richardson P, Bruce D, Rubin E, Myers E, Siggia ED, et al. 2007. Tracking the in vivo evolution of multidrug resistance in *Staphylococcus aureus* by whole-genome sequencing. *Proc Natl Acad Sci* **104**: 9451–9456.
- Ouellette M, Légaré D. 2002. Clinical relevance of drug resistance mediated by ABC-transporters in parasites of humans. In *ABC transporters* (eds. B. Holland et al.), pp. 317–333. Academic Press, New York.
- Ouellette M, Legare D, Papadopoulou B. 1994. Microbial multidrug-resistance ABC transporters. *Trends Microbiol* **2**: 407–411.
- Pringle M, Poehlsgaard J, Vester B, Long KS. 2004. Mutations in ribosomal protein L3 and 23S ribosomal RNA at the peptidyl transferase centre are associated with reduced susceptibility to tiamulin in *Brachyspira* spp. isolates. *Mol Microbiol* **54**: 1295–1306.
- Prystowsky J, Siddiqui F, Chosay J, Shinabarger DL, Millichap J, Peterson LR, Noskin GA. 2001. Resistance to linezolid: Characterization of mutations in rRNA and comparison of their occurrences in vancomycin-resistant enterococci. *Antimicrob Agents Chemother* **45**: 2154–2156.
- Richter E, Rusch-Gerdes S, Hillemann D. 2007. First linezolid-resistant clinical isolates of *Mycobacterium tuberculosis*. *Antimicrob Agents Chemother* **51**: 1534–1536.
- Roberts MC. 2008. Update on macrolide-lincosamide-streptogramin, ketolide, and oxazolidinone resistance genes. *FEMS Microbiol Lett* **282**: 147–159.
- Robertson GT, Doyle TB, Lynch AS. 2005. Use of an efflux-deficient streptococcus pneumoniae strain panel to identify ABC-class multidrug transporters involved in intrinsic resistance to antimicrobial agents. *Antimicrob Agents Chemother* **49**: 4781–4783.
- Ruggero KA, Schroeder LK, Schreckenberger PC, Mankin AS, Quinn JP. 2003. Nosocomial superinfections due to linezolid-resistant *Enterococcus faecalis*: Evidence for a gene dosage effect on linezolid MICs. *Diagn Microbiol Infect Dis* **47**: 511–513.
- Sander P, Belova L, Kidan YG, Pfister P, Mankin AS, Bottger EC. 2002. Ribosomal and non-ribosomal resistance to oxazolidinones: Species-specific idiosyncrasy of ribosomal alterations. *Mol Microbiol* **46**: 1295–1304.
- Schlunzen F, Zarivach R, Harms J, Bashan A, Tocilj A, Albrecht R, Yonath A, Franceschi F. 2001. Structural basis for the interaction of antibiotics with the peptidyl transferase centre in eubacteria. *Nature* **413**: 814–821.
- Sinclair A, Arnold C, Woodford N. 2003. Rapid detection and estimation by pyrosequencing of 23S rRNA genes with a single nucleotide polymorphism conferring linezolid resistance in *Enterococci*. *Antimicrob Agents Chemother* **47**: 3620–3622.
- Soualhine H, Brochu V, Menard F, Papadopoulou B, Weiss K, Bergeron MG, Legare D, Drummelsmith J, Ouellette M. 2005. A proteomic analysis of penicillin resistance in *Streptococcus pneumoniae* reveals a novel role for PstS, a subunit of the phosphate ABC transporter. *Mol Microbiol* **58**: 1430–1440.
- Tettelin H, Nelson KE, Paulsen IT, Eisen JA, Read TD, Peterson S, Heidelberg J, DeBoy RT, Haft DH, Dodson RJ, et al. 2001. Complete genome sequence of a virulent isolate of *Streptococcus pneumoniae*. *Science* **293**: 498–506.
- Thornsberry C, Brown NP, Draghi DC, Evangelista AT, Yee YC, Sahm DF. 2008. Antimicrobial activity among multidrug-resistant *Streptococcus pneumoniae* isolated in the United States, 2001–2005. *Postgrad Med* **120**: 32–38.
- Toh SM, Mankin AS. 2008. An indigenous posttranscriptional modification in the ribosomal peptidyl transferase center confers resistance to an array of protein synthesis inhibitors. *J Mol Biol* **380**: 593–597.
- Toh SM, Xiong L, Arias CA, Villegas MV, Lolans K, Quinn J, Mankin AS. 2007. Acquisition of a natural resistance gene renders a clinical strain of methicillin-resistant *Staphylococcus aureus* resistant to the synthetic antibiotic linezolid. *Mol Microbiol* **64**: 1506–1514.
- Tsakris A, Pillai SK, Gold HS, Thauvin-Eliopoulos C, Venkataraman L, Wennersten C, Moellering RC Jr, Eliopoulos GM. 2007. Persistence of rRNA operon mutated copies and rapid re-emergence of linezolid resistance in *Staphylococcus aureus*. *J Antimicrob Chemother* **60**: 649–651.
- Tsiodras S, Gold HS, Sakoulas G, Eliopoulos GM, Wennersten C, Venkataraman L, Moellering RC, Ferraro MJ. 2001. Linezolid resistance in a clinical isolate of *Staphylococcus aureus*. *Lancet* **358**: 207–208.
- Vara Prasad JV. 2007. New oxazolidinones. *Curr Opin Microbiol* **10**: 454–460.
- Wilson DN, Schlunzen F, Harms JM, Starosta AL, Connell SR, Fucini P. 2008. The oxazolidinone antibiotics perturb the ribosomal peptidyl-transferase center and effect tRNA positioning. *Proc Natl Acad Sci* **105**: 13339–13344.
- Wilson P, Andrews JA, Charlesworth R, Walesby R, Singer M, Farrell DJ, Robbins M. 2003. Linezolid resistance in clinical isolates of *Staphylococcus aureus*. *J Antimicrob Chemother* **51**: 186–188.
- Wolter N, Smith AM, Farrell DJ, Schaffner W, Moore M, Whitney CG, Jorgensen JH, Klugman KP. 2005. Novel mechanism of resistance to oxazolidinones, macrolides, and chloramphenicol in ribosomal protein L4 of the pneumococcus. *Antimicrob Agents Chemother* **49**: 3554–3557.
- Xiong L, Kloss P, Douthwaite S, Andersen NM, Swaney S, Shinabarger DL, Mankin AS. 2000. Oxazolidinone resistance mutations in 23S rRNA of *Escherichia coli* reveal the central region of domain V as the primary site of drug action. *J Bacteriol* **182**: 5325–5331.

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