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A gene duplication affecting expression of the ovine *ASIP* gene is responsible for white and black sheep

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Agouti signaling protein (*ASIP*) functions to regulate pigmentation in mice, while its role in many other animals and in humans has not been fully determined. In this study, we identify a 190-kb tandem duplication encompassing the ovine *ASIP* and *AHCY* coding regions and the *ITCH* promoter region as the genetic cause of white coat color of dominant white/tan (A^{Wt}) agouti sheep. The duplication 5' breakpoint is located upstream of the *ASIP* coding sequence. Ubiquitous expression of a second copy of the *ASIP* coding sequence regulated by a duplicated copy of the nearby *ITCH* promoter causes the white sheep phenotype. A single copy *ASIP* gene with a silenced *ASIP* promoter occurs in recessive black sheep. In contrast, a single copy functional wild-type (A^+) *ASIP* is responsible for the ancient Barbary sheep coat color phenotype. The gene duplication was facilitated by homologous recombination between two non-LTR SINE sequences flanking the duplicated segment. This is the first sheep trait attributable to gene duplication and shows nonallelic homologous recombination and gene conversion events at the ovine *ASIP* locus could have an important role in the evolution of sheep pigmentation.

[Supplemental material is available online at www.genome.org. The sequence data from this study have been submitted to GenBank under accession nos. EU185093–EU185100 and EU420022–EU420031.]

The genomic organization of the agouti signaling protein gene (*ASIP*) is generally highly conserved in mammalian species, including the mouse (Bultman et al. 1992), human (Kwon et al. 1994), horse (Rieder et al. 2001), pig (Leeb et al. 2000), cow (Girardot et al. 2005), and dog (Kerns et al. 2004). In the mouse genome, various insertions and deletions affecting *ASIP* and nearby loci have resulted in deregulated *ASIP* expression causing yellow pigmentation, adult onset obesity, diabetes, tumor growth, and embryonic lethality (Wolff et al. 1999). Other mice mutations in both the coding and regulatory regions affect *ASIP* expression and function and subsequent coat pigmentation patterns (Bennett and Lamoreux 2003; Eppig et al. 2005). *ASIP* alleles causing coat color variation have also been characterized in domestic dogs (Kerns et al. 2004), cats (Eizirik et al. 2003), pigs (Drogemuller et al. 2006), horses (Rieder et al. 2001), rats (Kuramoto et al. 2001), and foxes (Vage et al. 1997). Although *ASIP* transcripts are present in various human tissues, including liver, kidney, heart, and adipose tissue, the role of *ASIP* in humans is not clear (Voisey and van Daal 2002) but may include both pigmentation (Kanetsky et al. 2002; Bonilla et al. 2005; Voisey et al. 2006) and energy homeostasis (Voisey et al. 2002).

In mammalian species, coat color is an important form of camouflage and can be an integral part of social communication and recognition (Sponenberg 1997). The standard wild-type sheep coat color is generally dark-bodied with a pale belly, similar to other mammalian wild-type coat color patterns (Sponenberg 1997). However, this wild-type coat color pattern is much rarer in domestic sheep, where coat color is an important breed characteristic and production trait. In domestic breeds, unlike in their wild ancestors, the lack of natural selection allows coat color variants to arise and segregate. As a result of artificial selection for white fibers, the white coat phenotype has reached a

high frequency in certain breeds and shows autosomal dominant inheritance. In mice the dark dorsal and pale ventral wild-type coat color pattern has been shown to be caused by the spatial expression of different transcripts from a single *ASIP* gene (Vrieling et al. 1994), presumably controlled by different regulatory elements.

The contribution of *ASIP* to coat color patterns of domestic sheep has been investigated via classical genetics since the early 1920s. The dominant white or tan (A^{Wt}) *ASIP* allele is responsible for the phaeomelanin (yellow/red) phenotype in modern sheep breeds, while the most recessive allele, non-agouti (A^a) results in eumelanin (black/brown) phenotypes (Brooker and Dolling 1969; Adalsteinsson 1970). Another *ASIP* allele, badgerface (A^b), is characterized by a pale dorsal phaeomelanin and darker ventral eumelanin pattern; it is recessive to A^{Wt} and dominant to A^a (Brooker and Dolling 1969). Independent loci controlling color directly and structural features of the hair/wool coat also modify the phaeomelanin colors such that wool areas are typically white (Sponenberg 1997). Despite strong selection for white wool color, the frequency of the recessive non-agouti (also known as self-color black) A^a allele in the Australian Merino flock was estimated at 0.03 (Hayman and Cooper 1965). While strong comparative and classical genetic evidence for *ASIP* involvement in sheep dominant white and recessively colored coat phenotypes exist, the molecular genetic cause has remained unresolved (Parsons et al. 1997; Parsons et al. 1999a,b).

To identify the molecular genetic cause of coat color variation in domestic sheep, we have used sequence analysis of genomic DNA, BAC clones, and RT-PCR products from Romanov, Texel, Merino, and ancient Barbary sheep to characterize the genomic structure of the *ASIP* locus. We report a tandem duplication of a 190-kb portion of the ovine genome is responsible for the dominant white coat color (A^{Wt}) allele of domestic sheep. An asymmetric competitive PCR assay was developed and used to show that the multiple copy *ASIP* alleles consistently segregate with the dominant white coat color in Merino sheep. RT-PCR experiments were used to demonstrate alternative splicing of *ASIP* transcripts and show expression in multiple tissues of white

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sheep and lack of expression of the single copy alleles in black sheep. 5'RACE experiments determined that unlike the silent single copy *ASIP* of recessive black Merino sheep, a single copy Barbary sheep *ASIP* with a functional promoter is responsible for the wild-type pale belly phenotype.

Results

The ovine *ASIP* gene intron and exon structure

By sequencing the 5379-base-pair (bp) ovine *ASIP* gene from Merino sheep, (GenBank accession no. EU420022), we determined the genomic organization of the ovine *ASIP* coding exons 2, 3, and 4 and introns 2 and 3 to be similar to that reported for the bovine, human, and mouse genes (Fig. 1). Each coding exon is flanked by a consensus splice donor and acceptor site. BLASTN of the *ASIP* sequence against GenBank databases showed greatest identity to the bovine *ASIP* sequence (95.2%). The ovine gene encodes a putative 133 amino acid protein, which is 98%, 76%, and 74% identical to the bovine (133 amino acid), mouse (131 amino acid), and human (132 amino acid) proteins, respectively.

Mutation screening

To identify possible recessive mutations causing pigmentation, we sequenced the entire *ASIP* coding and noncoding sequence of Romanov, Texel, and Merino sheep and identified four coding sequence mutations compared with the published ovine *ASIP* sequence (Fig. 1; Parsons et al. 1999b). A previously reported 5-bp deletion, g.100-105delAGGAA (denoted D₅), was present in exon 2 (Smit et al. 2002). A novel 9-bp deletion, g.10-19delAGCCGCCTC (denoted D₉), was also found to be present in exon 2 10 bp from the ATG start codon, and two SNPs were found in exon 4 (g.5051G>C and g.5172T>A). Both the D₅ dele-

tion and the g.5172T>A SNP in exon 4 would be predicted to independently cause functional changes to the agouti protein. The D₅ deletion would result in a frame shift followed by a premature stop codon 63 amino acids downstream of the start site, truncating the agouti protein before the functionally important cysteine signaling domain (amino acids 91–130). The g.5172T>A SNP would predict a change of cysteine (amino acid 123) to serine, which would disrupt the highly conserved signaling region of the protein. The D₉ deletion would result in the loss of a tripeptide (SRL), which may affect the function of the *ASIP* transport leader sequence but not the remainder of the protein, while the synonymous g.5051G>C SNP in exon 4 would not be expected to disrupt *ASIP* function.

Association analysis of the D₉ and D₅ deletions and A allele with recessive black Merino phenotypes

A coat color panel comprising 373 DNA samples from white ($n = 183$), self-color black ($n = 142$), and badgerface ($n = 48$) Australian Merino sheep was genotyped for the D₅ and D₉ indels and the g.5172T>A SNP. The self-color black Merinos displayed a typical agouti symmetrical pigmentation pattern of a dark body with a white blaze on the head and neck (Fig. 2A). A characteristic badgerface Merino with dark ventral and lighter dorsal symmetrical agouti pigmentation pattern is shown in Figure 2B. None of the white Merinos were homozygous for the D₉, D₅, or A alleles. Furthermore, 78 self-color black Merinos that were homozygous D₅ were also homozygous T at the g.5172T>A SNP, and only 11% of the recessive black sheep were homozygous A. Approximately 16% of the white sheep and less than 2% of the black sheep had the D₉ allele, and none of the animals had an allele with both of these deletions. During genotyping, we also deduced that 8% of the Merino sheep had all alleles—nondeleted alleles at the D₉ and D₅ positions (denoted N₉ and N₅) and the D₉ and D₅ alleles, confirming the previous report of another *agouti*-like locus (Smit et al. 2002). As none of the white Merinos were homozygous for the D₉, D₅, or A alleles, the presence of the functional N₉, N₅, and T haplotype thus seems necessary for a Merino sheep to be white. However, not all black animals (self-color and badgerface) in our panel could be explained by homozygosity of a recessive nonfunctional D₅ or A allele. PCR products spanning *ASIP* exons 2, 3, and 4 in four self-color black Merino sheep homozygous for the N₉, N₅, and T alleles were amplified and sequenced directly. No other coding mutations that might predict a nonfunctional mature agouti protein were present.

ASIP expression in Merino sheep

Allele-specific expression of functional *ASIP* transcripts in white sheep skin

The expression of alleles with the protein altering D₅ mutation and the D₉ mutation were investigated in nine white Merino sheep. PCR and polyacrylamide electrophoresis were used to examine the expression of the variant alleles (Fig. 3). Seven white sheep with the N₉N₅ and N₉D₅ alleles and one sheep (Fig. 3, lane 5) with all three alleles (N₉N₅, N₉D₅, and D₉N₅) showed that the nondeleted N₉N₅ and deleted D₉N₅ alleles were preferentially expressed in Merino skin as visualized by the band intensities. The N₉D₅ allele was not amplified by competitive RT-PCR (Fig. 3, arrow, lanes 1–9) and was considered comparatively lowly expressed or not expressed at all.

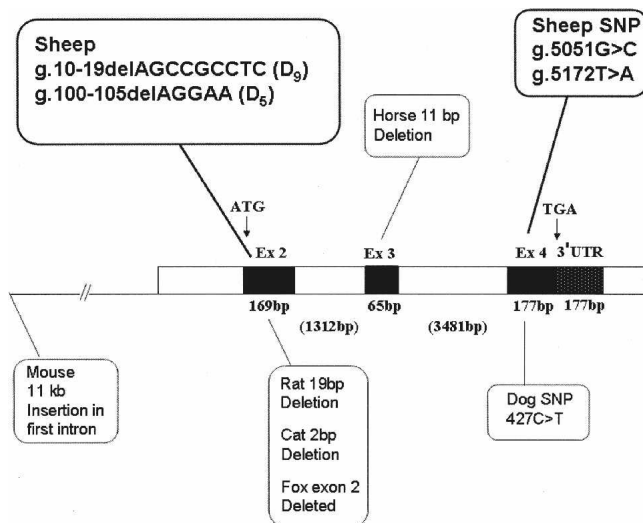


Figure 1. Schematic of the structure of the ovine *ASIP* gene showing the sizes of the three coding exons (black) and the intervening intron sequences (white). The exon–intron organization of the ovine gene is similar to that reported for the bovine, human, and mouse genes. Coding exons 2, 3, and 4 are separated by 1312- and 3481-bp intron sequences, respectively. The nucleotide positions of various described recessive black “non-agouti” mutations for the mouse, rat, horse, cat, fox, and dog are shown. The positions of four mutations identified from sheep in this study are shown.

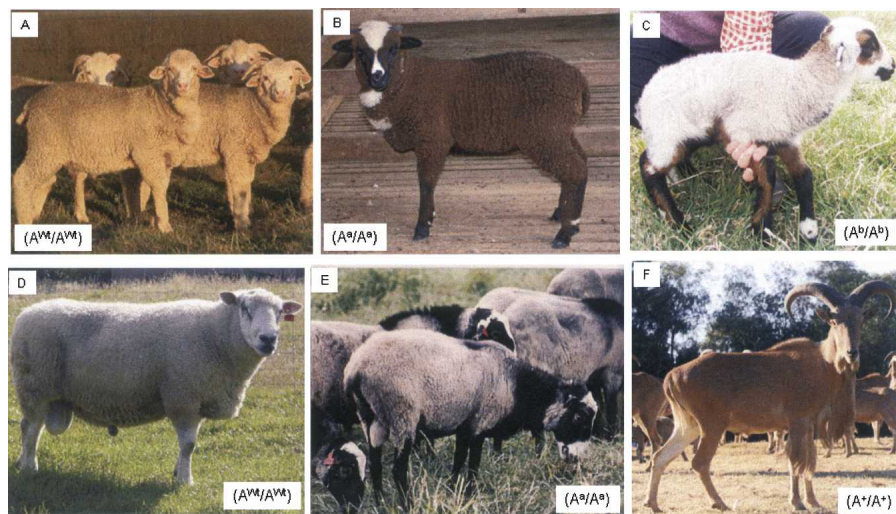


Figure 2. Illustration of sheep coat color patterns. Three coat colors in the Australian Merino; (A) the dominant white; (B) the non-agouti, also known as recessive self-color-black; and (C) the badgerface pattern. The dominant white coat color in Texel is displayed in D. Romanov sheep (E) appear to have a pattern similar to recessive self-color-black and are proposed as homozygous for the most recessive *ASIP* allele, A^o . (F) The Barbary sheep wild-type coat color. Hypothesized genotypes for *ASIP* are indicated.

ASIP expression in white sheep is controlled by an *ITCH* promoter

By use of gene-specific primers designed from *ASIP* ESTs (data not shown), RT-PCR analysis of skin and internal tissues from white and recessive black Merinos indicated that the dominant white phenotype of Merino sheep was caused by high levels of deregulated expression of the *agouti* gene from an itchy homolog E3 ubiquitin protein ligase (*ITCH*) gene promoter. The same alternatively spliced forms of ovine *ASIP* were expressed in liver, kidney, heart, spleen, and skin tissues tested from white Merinos but were not amplified from any tissues of a self-color black Merino (Fig. 4A–F). *ASIP* ubiquitous expression has also been reported in humans (Voisey et al. 2002) and cattle (Girardot et al. 2005) and represents a departure from what is seen in mice, where expression is generally confined to the skin (Bultman et al. 1992). RT-PCR (Fig. 4A,B) and 3' RACE (Fig. 4C) experiments also indicated all the *ASIP* transcripts started with two noncoding exons, designated It and It', and the same transcripts were present in all the white Merino sheep tissues examined. The expression of ovine *ITCH*, however, was not affected with expression of the same order of magnitude in both the black and white sheep tissues (Fig. 4D,F). The "normal" expression of *ITCH* explains why sheep do not experience the spectrum of immunological conditions evident in the "itchy" mouse, in which both *ITCH* and *ASIP* expression is disrupted due to a genomic inversion affecting both loci (Perry et al. 1998). *ASIP* and *ITCH* expression levels in the skin and other tissues was confirmed by real-time quantitative PCR analysis of three biological replicates each of Merino white and black sheep tissue (data not shown).

Alternatively spliced *ASIP* transcripts

RT-PCR identified several *ASIP* transcripts from the skin of white Merino sheep. Primer Agt9 (positioned in the first noncoding exon It) and primer Agt6 (in the 3' UTR of *ASIP*) (Supplemental Table S2) were used to amplify cDNA derived from the skin of white Merino sheep. These products were cloned and sequenced, and seven

alternative transcripts (GenBank accession nos. EU420024–EU420030) were obtained (Fig. 5A). Significant differences were evident for the sequences of the ovine skin noncoding exons compared with those reported for cow, mouse, and pig (Vrieling et al. 1994; Leeb et al. 2000; Girardot et al. 2005). Every ovine skin transcript had the two noncoding exons that we have labeled It and It' in addition to one or two other noncoding exons designated IA to IE (Fig. 5A). A terminal G nucleotide on the sequence of *ASIP* transcripts derived by 5' RACE determined exon It to be the most 5' noncoding exon. The noncoding exons It and It' were also part of the 5' region of the downstream *ITCH* gene (Fig. 5A) and were 67% identical to the noncoding exons in *ASIP* transcripts from the sienna yellow (*Asy*) mouse (Duhl et al. 1994).

The expression and RT-PCR results indicated that the dominant white phenotype of Merino sheep is caused by high levels of expression of functional

ASIP transcripts driven by an *ITCH* promoter region. Comparative analysis of the human, dog, and mouse genomes (<http://genome.ucsc.edu/>) places the *ITCH* gene downstream of *ASIP* in these species. The *ITCH-ASIP* hybrid transcripts therefore suggested the presence of a tandem duplication or rearrangement and deletion in this region of the ovine genome.

Identification of a duplication encompassing the *ASIP*, *AHCY*, and *ITCH* regions

As the ovine genome sequence was not yet determined, to characterize the ovine *ASIP-ITCH* genomic region, we obtained and

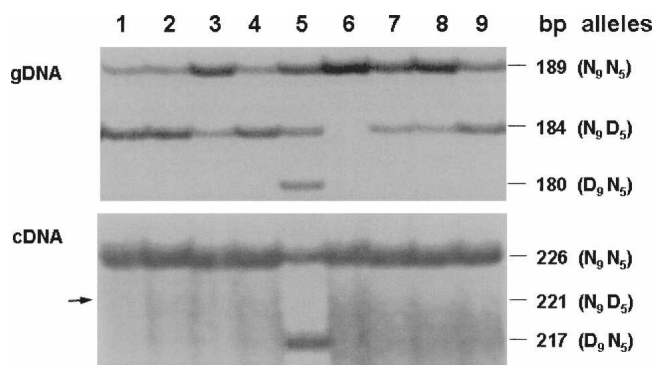


Figure 3. Allele-specific expression of *ASIP* transcripts in nine white Merino sheep. Autoradiographs of PCR products from gDNA and cDNA from the skin of nine white sheep are shown. The size difference between the 189 (226)- and 184 (221)-, and the 189 (226)- and 180 (217)-bp fragments is due to the corresponding D_5 and D_9 deletions in the latter fragments. Seven sheep had both the N_9N_5 and N_9D_5 alleles. In one sheep (lane 5) all three alleles N_9N_5 , N_9D_5 , and D_9N_5 were present. One sheep had only the nondeleted N_9N_5 alleles (lane 6). The nine white animals showed allele-specific expression of the functional N_9N_5 and D_9N_5 alleles. Expression of the nonfunctional N_9D_5 allele (arrow) was not detected. The different band intensities in the gDNA autoradiograph likely reflect the different allele copy numbers present in each animal.

ASIP copy number variants of white and black sheep

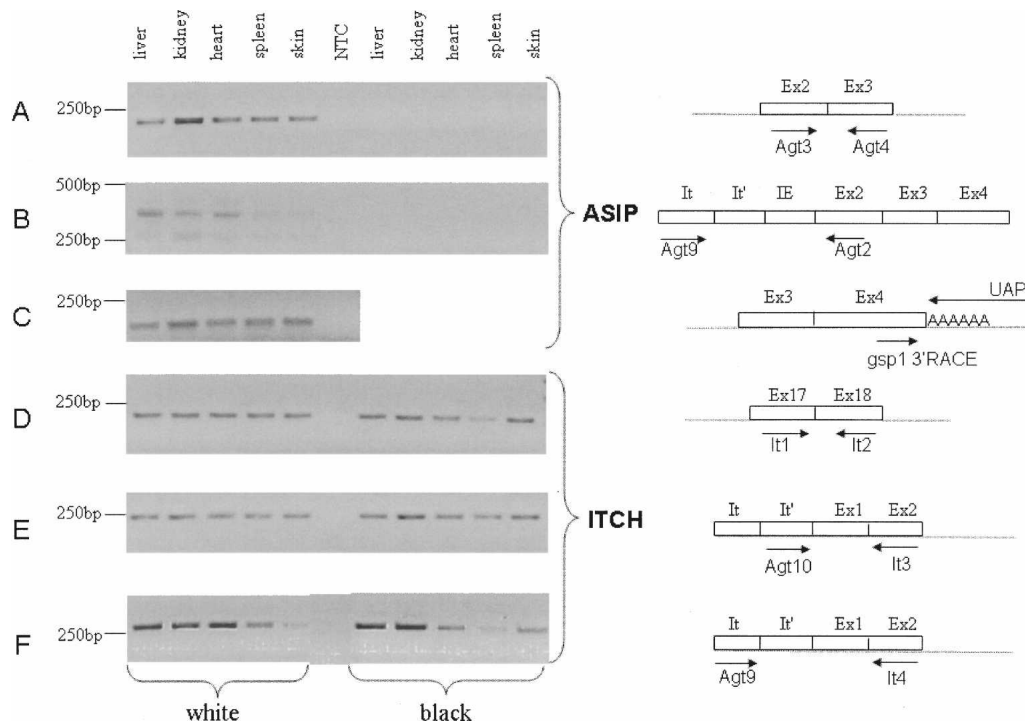


Figure 4. RT-PCR end-point gene expression analysis of ovine *ASIP* and *ITCH* in five tissues of a white and a self-color black Merino sheep. The position of primers used to amplify each product is shown in a schematic to the right of the corresponding 1.5% agarose gel image. It, It', and IE are noncoding exons. Open boxes Ex2, Ex3 and Ex4 (A, B, and C) and open boxes Ex17, Ex18, Ex1, and Ex2 (D, E, and F) represent coding exons of the *ASIP* or *ITCH* genes, respectively. Other alternatively spliced transcripts of the agouti gene have also amplified as evident from the bands in panel B. The most abundant *ASIP* transcripts are those with exon IE (~368 bp as shown in the panel B schematic). It and It' were both present in *ITCH* transcripts and generated bands of the expected size (based on similarity to the bovine sequence), but no alternatively spliced forms were evident. 3' RACE (panel C) showed *ASIP* transcripts in all tissues studied to have the same polyadenylation site.

sequenced three Romanov sheep and five Texel sheep BAC clones. Sequencing of these ovine BAC clones identified a large (~190 kb) tandem genomic duplication (Fig. 5A; Supplemental Fig. S1). Alignment of *ASIP* noncoding exons It (77 bp), It' (78 bp), and IA (54 bp) sequences identified from alternatively spliced transcripts from the skin of white Merino sheep to BAC clone INRA-164H8 and INRA-229C6 sequences from a Romanov sheep positioned them ~119, 123, and 131 kb, respectively, 3' of the *ASIP* start codon (Fig. 5A). The ID (56 bp) and IE (88 bp) noncoding exons were located ~6.6 kb and 298 bp, respectively, 5' of the *ASIP* start codon (Fig. 5A). The IB (178 bp) and IC (97 bp) noncoding exons were not present in INRA-164H8 sequence, which ended ~10 kb 5' of the *ASIP* start codon. Comparative alignment of the noncoding exon IB and IC sequences to the dog, human, and cow genome sequences placed these exons further 5' of the *ASIP* start codon in these species (data not shown).

The Merino *ASIP* intron and exon sequence data and the INRA BAC clone sequence data were used to search the CHORI Texel sheep library BAC-end sequences to identify BAC clones in the region of the ovine *ASIP* gene. In addition, the mapping of the full set of sheep BACs to the cow, dog, and human genomes (Dalrymple et al. 2007) was used to identify BACs potentially located across the putative duplication and flanking genomic regions. Twenty-nine clones that spanned ~2 Mb of sequence encompassing the *ASIP*, S-adenosylhomocysteine hydrolase (*AHCY*), *ITCH*, and flanking genomic regions of ovine chromosome 13 was obtained for analysis. The BAC clones were then ordered across the region and five BACs predicted to span the

complete region including the junction point, and 5' and 3' breakpoints were sequenced. We obtained ~500 kb of contiguous sequence that contained a 190-kb tandem duplication, including the complete *ASIP* and *AHCY* coding regions and the *ITCH* promoter and noncoding exon sequences It, It', and IA (Fig. 5A). Sequence data from clones CH243-455O4, CH243-234K21, and CH243-489F15 spanned the junction between the tandem duplicated copies. BAC clone CH243-160L8 spanned the 3' breakpoint, and clone CH243-373J16 spanned the 5' breakpoint.

Romanov, Texel, and Merino sheep haplotypes

Sequencing of the Romanov and Texel Sheep BAC clones and Merino sheep PCR products from genomic DNA identified seven ovine *ASIP* haplotypes (Supplemental Table S1). Haplotypes 1 and 2 were determined by cloning and sequencing the PCR products from the genomic DNA of a white (A^{W^c}) and a self-color black (A^c) Merino sheep. The white animal contained haplotypes 1 and 2, and the self-color black animal contained only haplotype 2. Ovine *ASIP* haplotypes 3 and 4 were determined by sequencing of INRA BAC clones INRA-218G7, INRA-229C6, and INRA-164H8 from Romanov sheep. Ovine *ASIP* haplotypes 5, 6, and 7 were identified from the Texel sheep BAC clone sequences. Haplotype 5 was designated *ASIP* copy 1 as it was positioned 5' to haplotypes 6 and 7, both designated *ASIP* copy 2 (Fig. 5A). Haplotypes 6 and 7 (*ASIP* copy 2) were 99.9% identical to the functional haplotypes 1 and 3 from Merino and Romanov sheep, while haplotype 5 (*ASIP* copy 1) showed greatest identity (99.7%) to the non-functional A allele containing haplotype 4 from Romanov sheep

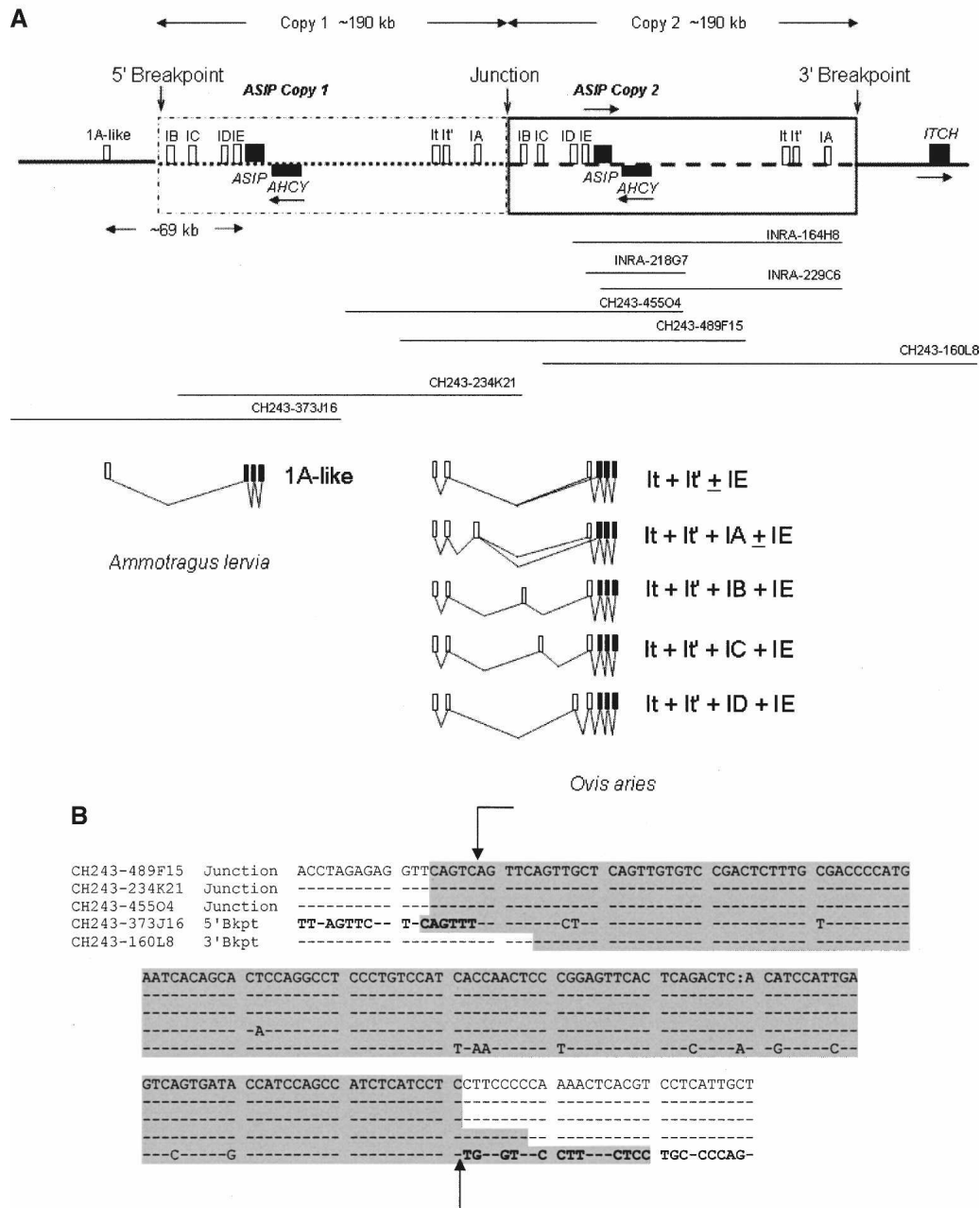


Figure 5. Schematic showing the sequenced ovine BAC clones below the structure of the ovine genomic tandem duplication and an alignment of the breakpoint and junction point sequences. (A) Noncoding exons are shown as open boxes. Protein coding exons are shown as solid black boxes. Arrows above or below the genes indicate the direction of transcription. Clones INRA-164H8, INRA-218G7, and INRA-229C6 were from the INRA Romanov Sheep BAC library (Vaiman et al. 1999), and clones CH243-160L8, CH243-234K21, CH243-455O4, CH243-373J16, and CH243-489F15 were from the CHORI-243 Texel sheep BAC library (Dalrymple et al. 2007). The three INRA BAC clones did not span the junction, 5' or 3' breakpoints. Clones CH243-455O4, CH243-489F15, and CH243-234K21 spanned the junction between copies; clone CH243-160L8 spanned the 3' breakpoint and clone CH243-373J16 the 5' breakpoint. Seven *ASIP* transcripts identified from the skin of a white Merino sheep (*Ovis aries*) and one transcript identified from the ventral skin of a Barbary sheep (*Ammotragus lervia*) are shown in the lower section. The coding exons 2, 3, and 4 and Barbary sheep noncoding exon 1A-like are numbered according to the nomenclature of Bultman et al. (1992) and Vrieling et al. (1994). All other noncoding exons are named alphabetically, IA to IE. Exons It and It' are noncoding exons of both *ITCH* and *ASIP* Merino transcripts (see also Fig. 4). The positions of the 5' and 3' breakpoints are located 5' of the *ASIP* and *ITCH* coding sequence regions, respectively. The *ITCH* promoter, including noncoding exons It, It', and IA, was duplicated and positioned upstream of the duplicated *ASIP* noncoding exons IB to IE, creating a new ovine hybrid *ITCH/ASIP* promoter. The complete *ASIP* and *AHCY* coding exons were also within the ~190-kb duplicated segment. Not drawn to scale. (B) DNA sequences from five Texel sheep BAC clones comprising the regions spanning the 5' and 3' breakpoints and the junction point between the gene duplication. Sequence identity to the master sequence is shown with a dash. The boundaries of the 143 bp of sequence similarity are marked with vertical arrows. Flanking sequences unique to the 5' and 3' breakpoint regions, respectively, are in bold. The highlighted (gray) regions of clone CH243-160L8 sequence of the 3' Breakpoint was identified by Repbase (Kohany et al. 2006) as having 82% identity to a region of Bovidae non-LTR/RTE BDDF2 repetitive SINE sequence. The highlighted (gray) regions of clones CH243-373J16 (of the 5' breakpoint) and clones CH243-489F15, CH243-234K21, and CH243-455O4 (all of the junction point) were identified as having 87%–88% identity to a region of Bovidae non-LTR BOV2 repetitive SINE sequence.

(see also Fig. 6). The position of the sheep functional haplotypes 6 and 7 at *ASIP* copy 2 places them 3' to, and therefore under, the regulation of the duplicated *ITCH* promoter (Fig. 5A).

Haplotypes 1, 6, and 7 all contained the proposed functional N_9 , N_5 , and T alleles. Haplotype 2 contained the predicted nonfunctional D_5 and functional T alleles and was otherwise very similar to Haplotype 1. Haplotype 3 contained the D_9 , N_5 , and T alleles and was also otherwise very similar to the functional haplotypes 1, 6, and 7. Haplotypes 4 and 5 both contained the predicted nonfunctional A allele and were quite different to all other haplotypes. Using haplotype 1 as the reference sequence, haplotypes 4 and 5 had 35 and 40 nucleotide substitutions, respectively. Transcripts from haplotypes 1, 3, 6, and 7 would be predicted to produce functional agouti protein, while transcripts from haplotypes 2, 4, and 5 would be predicted to produce nonfunctional agouti protein.

Nonallelic pairing, crossover, and gene conversion events arguably occur more frequently between almost identical tandem duplications (Lindsay et al. 2006; Myers and McCarroll 2006), and thus, the tandem duplications of the *ASIP* gene are likely candidates for these events. Indeed, the sequences of the *ASIP* copy 1 and 2 haplotypes suggest mutations, recombination, and gene conversion events possibly have occurred between them. It is conceivable that Haplotype 5 (*ASIP* copy 1) could be derived from recombination and/or gene conversion between Texel sheep equivalents of haplotypes 1 and 4. In addition, Texel sheep haplotype 6 (*ASIP* copy 2), which was otherwise identical to haplotype 7 (*ASIP* copy 2), shared two nucleotides (at positions 4124 bp and 5051 bp) with haplotype 5 (*ASIP* copy 1), which are likely a result of mutation or gene conversion events. Haplotype 5 also had an A nucleotide at position 3096 bp not present in any other haplotypes that is likely a result of a mutation.

Analysis of the duplication junction and breakpoint sequences

We analyzed the duplication junction and breakpoint sequences for regions that could facilitate duplication events. By comparing the DNA sequence of each of the BAC clones containing the junction, 5' and 3' breakpoints, a region of sequence similarity of ~143 bp was identified at all three sites (Fig. 5B). These sequences were identified by Repbase (Kohany et al. 2006) as having 82%–88% identity to Bovidae non-LTR BOV2 and BDDF2 repetitive SINE sequence regions. The duplication may have been facilitated by homologous recombination between these repetitive elements.

Analysis of *ASIP* copy number variation in domestic Merino sheep

Sequence flanking the shared 143-bp sequence at the junction and 5' breakpoint was used to develop an asymmetric competi-

tive PCR copy number assay (Supplemental Fig. S2). The junction point and 5' breakpoint PCR products (see Methods) were used initially to assess the presence and absence of duplicated copy alleles in the genomes of white and recessive black Merino sheep. All white Merinos successfully assayed ($n = 177$) amplified both a junction point and 5' breakpoint PCR product and therefore had duplicated *ASIP* alleles (Table 1). In contrast, all of the successfully assayed recessive black Merinos ($n = 180$) amplified a 5' breakpoint product only and thus had only single copy silent (refer expression data) *ASIP* alleles.

To calibrate the ABI3130xl copy number assay, PCR products of the junction point and 5' breakpoint were cloned into pCR 2.1-TOPO, and a standard curve was established using a dilution series of plasmid DNA containing the junction point mixed with plasmid DNA containing the 5' breakpoint from 0%–100%. The resulting standard curve was linear (Supplemental Fig. S3). The genomic DNA of 16 white Merino sheep (eight that were confirmed as carriers of a single copy recessive black-allele by pedigree analysis) was next analyzed (in quadruplicate) in an initial evaluation of the ABI3130xl copy number assay. Three of the confirmed carriers assayed contained a single junction point allele as would be predicted for a carrier animal (Supplemental Fig. S4, lanes 2,3,8). However, the five other carriers contained two junction points, which indicated triplicated alleles and suggested even greater diversity in copy number could occur at the ovine *ASIP* locus. The ABI3130xl copy number test was used to assess *ASIP* copy number variation in the white sheep of the Merino sheep coat color panel. The copy number genotypes of the assayed carrier and random white sheep indicated that white Merino sheep contain multiple copy *ASIP* alleles (three to six copies) per genome (Table 1).

Comparative analysis of *ASIP* copy number variation and expression in Barbary sheep (*Ammotragus lervia*), an ancient Caprinae species

Barbary sheep (Fig. 2) have a wild-type agouti coat color pattern—tan body and pale belly—and are assumed to be homozygous for the wild-type *ASIP* allele (A^+). PCR analysis ($n = 40$) amplified only the 5' breakpoint product, indicating single copy *ASIP* alleles (Table 1). The 5'RACE experiment determined that unlike the silent single copy *ASIP* of recessive black Merino sheep, the single copy Barbary sheep *ASIP* has a functional promoter and is responsible for the pale belly phenotype. The partial *ASIP* transcript (GenBank accession no. EU420031) from Barbary sheep ventral skin had a single noncoding 1A-like exon and coding exons 2, 3, and partial exon 4 sequence (Fig. 5A) with high similarity to the homologous ventral-specific mouse (Bultman et al. 1992; Vrieling et al. 1994) and pig 1A (Drogemuller et al. 2006) transcripts. Analysis of the Texel sheep BAC clone genomic se-

Table 1. Presence of junction points and estimated *ASIP* copy numbers associated with dominant white coat color in different sheep populations

Population	Proposed genotypes	Total number of junction points (<i>ASIP</i> copies)					Total
		0 (2)	1 ^a (3)	2 (4)	3 (5)	4 (6)	
Recessive black Merino	A^c/A^c or A^b/A^b	180	—	—	—	—	180
White carrier Merino	A^{wt}/A^c	0	47	29	9	1	86
Random white Merino	A^{wt}/A^{wt} or A^{wt}/A^c	0	8	67	16	0	91
Barbary sheep	A^+	40	—	—	—	—	40

^aThe single junction point indicates these white animals have a duplicated allele and a single copy black allele.

quence positioned the Barbary sheep noncoding 1A-like exon sequence ~69 kb 5' of *ASIP* copy 1 exon 2 in domestic sheep, 27 kb further upstream of the 5' breakpoint (Fig. 5A). Our data support a functional *ASIP* promoter driving expression of a single *ASIP* gene determining coat color patterns in this ancient species of the Caprinae (goat–antelope) subfamily.

Discussion

It has taken more than a century of mouse classical genetics and two decades of molecular genetic analysis to begin to understand the molecular mechanisms regulating yellow coat color and the associated pleiotropic effects from deregulated murine *ASIP* expression (Wolff 2003). The molecular regulation of coat color of other mammalian species at the *ASIP* locus, however, still remains comparatively unknown. The expression of the mouse *ASIP* gene ordinarily is tightly spatially and temporally regulated and restricted primarily to the skin (Bultman et al. 1992). In humans, (Voisey et al. 2002), cattle (Girardot et al. 2005, 2006), and, as we have now shown, in domestic sheep, *ASIP* is more widely expressed with various transcripts identified from several tissues. While there are no apparent negative effects of this ubiquitous *ASIP* expression in sheep, the possible consequences of the wider tissue expression is still to be investigated. Interestingly, in Icelandic sheep, pleiotropic effects of the dominant white or tan (A^{Wt}) allele result in reduced fecundity and greater seasonality in reproduction (Adalsteinsson 1975). However, in mice ubiquitous *ASIP* expression results in adult onset obesity, suggesting that the signaling pathways that normally regulate reproduction and body weight may be different in these mammalian species.

It was also recently shown that the *ASIP* coding sequence has been completely deleted from the gibbon genome (Nakayama and Ishida 2006) with no apparent detrimental effect, while it remains intact in all other investigated primate genomes (Mundy and Kelly 2006). Nakayama and Ishida (2006) speculate that the gibbon *ASIP* deletion may affect its energy homeostasis and so contribute to the smaller body mass of the gibbon relative to other primates. Studies in humans and animal models have elucidated a role for components of the melanocortin pathway in immunity, energy homeostasis, and reproduction (Henry 2003; Carroll et al. 2005). *ASIP* regulation of lipid metabolism in adipose tissue has been examined in mouse and humans (Voisey et al. 2002), but to date, no studies have investigated the potential impact of increased or deregulated *ASIP* expression on ovine energy balance or fertility. However, lean sheep fed ad libitum can become obese and have proven to be a useful animal model for human obesity (McCann et al. 1991; Henry 2003). Investigating the impact of *ASIP* copy number variation on key production traits of leanness, body fat, and fertility could provide new insights into the physiology of energy metabolism and reproduction of livestock and other mammalian species.

Mammalian species thus far investigated have had a single copy of the *ASIP* gene identified within their genomes. The genetic cause of coat color patterns in domestic sheep at the *ASIP* locus has been difficult to determine due in part to the prospect of a second *ASIP*-like locus, which was proposed by Smit et al. (2002). Further, with the exception of the mouse (Bultman et al. 1992, 1994) and the pig (Drogemuller et al. 2006), mutations in the coding sequence that disrupt functional *ASIP* protein were associated with the recessive black phenotypes of these species (Vage et al. 1997; Kuramoto et al. 2001; Rieder et al. 2001; Eizirik et al. 2003; Kerns et al. 2004). We have shown that the regulation

of the recessive black coat color phenotypes by the ovine *ASIP* gene is not attributable to simple coding mutations as described for other mammals and that the dominant white phenotype involves variation in gene copy number and deregulated expression.

Analysis of the ovine *ASIP* coding regions of recessive self-color black and badgerface Merino sheep identified four mutations, two of which (a 5-bp deletion and a g.5172T>A SNP) would be predicted to disrupt the functional protein. Both these coding sequence mutations, however, failed to completely associate with the recessive black Merino phenotypes. Of the recessive black Merinos, only ~60% and 11% were homozygous for either the D5 or A allele, respectively, and none were homozygous for both. All white Merinos investigated were either homozygous for the normal alleles or heterozygous for one or the other of these mutations. Additionally all white Merinos had at least one duplicated *ASIP* allele, while all of the recessive black Merinos contained only single copy alleles.

ASIP expression was detected in all tissues examined from white sheep, with multiple transcripts identified from both the skin and internal tissues. Expression was not detected from any tissues of recessive self-color black sheep. Further, all transcripts identified from white sheep skin began with the *ITCH* noncoding exons It and It', and none contained the 5-bp or g.5172T>A predicted nonfunctional mutations. These data indicated that expression in the white sheep was driven from the duplicated copy of an *ITCH* promoter positioned upstream of *ASIP* and that the identified functional mutations are not present in these expressed *ASIP* copies. The data also indicated that in domestic sheep the progenitor *ASIP* promoter, unlike the promoter of the single copy *ASIP* of ancient Barbary sheep, was not functional. Because *ASIP* expression was not detected in the single copy alleles of the recessive black sheep, the likely causative mutation of the recessive black phenotype is an as yet unidentified regulatory mutation of the progenitor gene promoter region.

In mice, the molecular events associated with an unusually high reversion rate of *agouti* recessive black coat color pattern mutations to more dominant alleles were identified to be as a result of insertions and deletions mediated by homologous recombination between repetitive elements (Bultman et al. 1994). In the gibbon, the removal of a 100-kb region including the *ASIP* coding sequence was mediated by Alu repeat elements (Nakayama and Ishida 2006). A large duplication involving the *KIT* locus, causing dominant white skin color in pigs, has originated by homologous recombination between LINE elements (Marklund et al. 1998; Giuffra et al. 2002). The sequences at the borders of the duplicated ovine *ASIP* gene segments contain repeat elements identified as BOV2 and BDDF2 non-LTR SINE sequences (Kohany et al. 2006). Thus, it seems likely that the ovine dominant white and recessive black phenotypes are mediated at least in part by homologous recombination involving these repeat elements. The sequencing and subsequent characterization of the human genome revealed that 5% of the human genome consists of long highly similar duplicated sequences known as low copy repeats (LCRs) or segmental duplications (Lindsay et al. 2006). It is now emerging that such segmental duplications are hotspots for allelic and nonallelic homologous recombination events that can result in genomic disorders causing clinical diseases (Bischof et al. 2006).

The exchange of sequence variations between duplicated loci known as gene conversion events mediated through allelic and nonallelic recombination (Myers and McCarroll 2006) can lead to a high degree of similarity between duplicated genes

ASIP copy number variants of white and black sheep

(Lindsay et al. 2006). A high degree of nucleotide sequence similarity (>99%) occurred between the 5379 bp of the multiple *ASIP* gene copies of each of the BAC clone sequences from the start of exon 2 to the end of the 3'-untranslated region (Supplemental Table S1). Gene conversion events together with allelic and non-

allelic homologous recombination between repetitive sequences are likely to be contributing to variation and evolution of ovine *ASIP* sequences.

Based on our results, we propose a model for the evolution of the ovine *ASIP* locus (Fig. 6). The model proposes that a duplication,

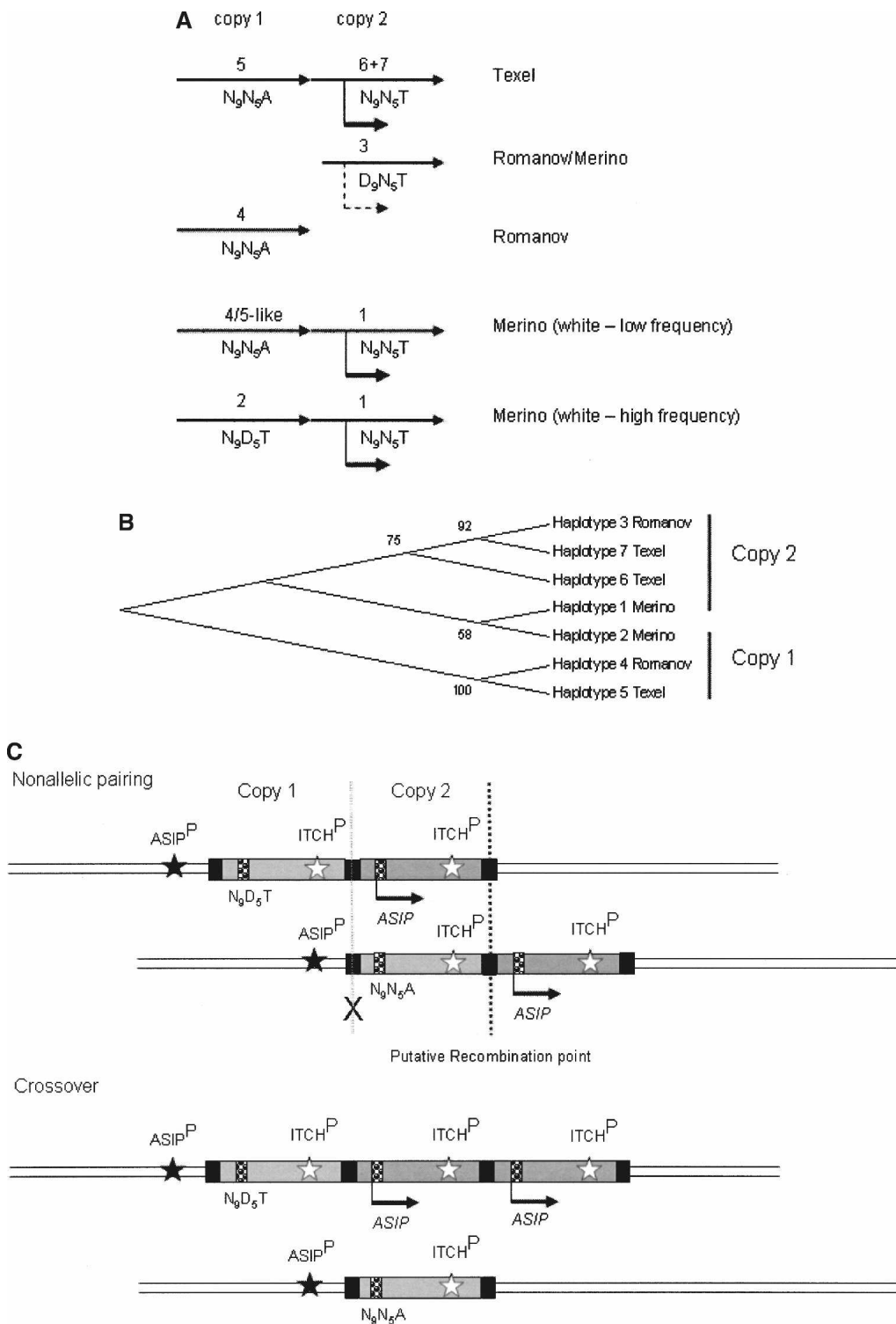


Figure 6. (Legend on next page)

mediated by homologous sequences in SINES flanking the *ASIP* coding and *ITCH* promoter regions, occurred in an ancestor of domestic sheep, positioning a second copy of the *ITCH* promoter upstream of a second *ASIP* coding sequence, deregulating its expression. Subsequently, possibly with the creation of the duplicated segment, or by other successive mutation events, the progenitor *ASIP* promoter was inactivated. Sheep with genotypes that have at least one duplicated *ASIP* allele are always white as strong (artificial) selection pressure for white coat color has maintained the functional (N₅, T) *ASIP* alleles at the expressed (copy 2) position (Fig. 6A). With the expressed (copy 2) *ASIP* functional coding sequence under strong selection pressure, the two *ASIP* regions subsequently diverged in sequence, generating functional and nonfunctional haplotype clusters (Supplemental Table S1; Fig. 6B). The most likely major route of generation of single copy loci was nonallelic pairing and recombination between duplicated alleles (Fig. 6C). The single copy recessive black causing alleles, with functional or nonfunctional coding sequence haplotypes, subsequently resulted from nonallelic recombination and/or gene conversion events (Fig. 6C). Our data from asymmetric competitive PCR copy number assays of recessive black and white Merinos show that while recessive black Merinos always have a single *ASIP* copy, white Merinos can have two to four and possibly even five *ASIP* copy alleles, further supporting this model.

Tandem gene duplication represents an underinvestigated source of molecular variation in livestock species. The duplication/deletion of the *agouti* gene is the first characterized example in sheep of the involvement of gene duplication in the creation of a genetic variation that contributes to a major breed and production trait—coat color phenotypes. This gene copy-number polymorphism introduces variation into the sheep genome and causes a phenotypic trait to which natural or artificial selection may then apply. The ovine chromosome 13 tandem duplication, encompassing the *ASIP* and *AHCY* genes and promoter region of the *ITCH* gene, is an ideal locus for the investigation of the consequences of duplication on sequence variation and evolution via natural and artificial selection in commercially important livestock species.

Classical genetic analysis first proposed alleles of the *agouti* gene as a major determinant of coat color phenotypes in sheep more than a half a century ago (Rendel 1957; Brooker and Dolling 1969). With a major goal of identifying the molecular cause of unwanted recessive black sheep phenotypes, the molecular genetic investigation of the ovine *agouti* gene in the Australian

Merino sheep was begun a decade ago (Parsons et al. 1997, 1999a,b). Until now, without the benefit of an ovine genome sequence, progress in understanding the molecular mechanisms regulating coat color in domestic and wild sheep populations at the *ASIP* locus has been elusive. The characterization of a large tandem duplication encompassing the *ASIP*, *AHCY*, and *ITCH* genes provides an explanation for this difficulty. In the future, further comparative investigation of the *agouti* architecture in domestic sheep breeds and extant wild sheep populations will, in evolutionary terms, determine when this dominant *agouti* mutation first occurred and its impact on and evolution since the domestication of sheep.

Methods

Panel of sheep coat color phenotypes

Samples of 6 mL blood (EDTA tubes) and 0.9-mm skin biopsies were collected from Australian Merino sheep. All experiments were approved by and performed according to the guidelines of the CSIRO Chiswick NSW Animal Ethics Committee (AEC approval nos. 01/19, 02/03, 03/62, 04/30). Color photographs displaying the dorsal, ventral, sides, and face were taken of each pigmented animal for determination of the self-color black or badgerface Merino phenotypes. Sheep were sourced from a wide geographic distribution throughout Australia, and animals from different producers were presumed to be unrelated. The Merino DNA panel consisted of samples from 94 baldy self-color black sheep (from 28 different producers), 48 spotted self-color black sheep (from 16 producers), 48 badgerface animals (from 12 producers), 87 white carrier animals (57 confirmed to be carriers by commercial pedigree testing and 30 suspected carriers) from a total of 20 different producers, and 96 random white animals (of unknown carrier status) from 22 producers. Barbary sheep (*A. lervia*) blood and ventral skin samples were sourced from the Western Plains Zoo (Dubbo, NSW Australia).

Preparation of gDNA and total RNA

QiAmp DNA Mini kit (Qiagen) was used to extract genomic DNA from whole blood. Skin biopsies were collected in RNA Later (Ambion) and stored at -80°C . Total RNA was prepared from the skin of white and recessive black Merino sheep and from the ventral skin of Barbary sheep using TRIzol (Invitrogen) in accordance with the manufacturer's recommendation. In each extraction, 5 mL of TRIzol was used to extract 200–500 mg of skin. To ensure removal of contaminating DNA, total RNA was treated

Figure 6. A model for the organization and evolution of the ovine *ASIP* locus in dominant white and recessive black domestic sheep. The proposed positions (A) and clustering (B) of copy 1 (inactive) and copy 2 (active) haplotypes identified from Texel, Romanov, and Merino sheep are shown. Solid arrows indicate high levels of expression of haplotypes at copy 2. A dashed arrow indicates expression of a haplotype 3-like D₉ allele detected in Merino skin. The two haplotypes identified in the Romanov BAC library may not be derived from a duplicated *ASIP-ITCH* region as pure Romanov sheep have a recessive black-like phenotype. However, haplotype 4 clusters with haplotype 5 in copy 1 of the Texel and haplotype 3 clusters with haplotype 6 and 7 from the Texel and haplotype 1 from the Merino (B). This clustering suggests that the Romanov haplotypes are derived from the two different copies of the putative original gene duplication. The low frequency presence of a haplotype 4/5-like copy 1 region in the Merino sheep population is deduced from the identification of N₉N₅A genotypes in the black Merinos and is consistent with the organization of the region in the Texel sheep CHORI-243 library BACs. (C) Schematic showing resolution of nonallelic pairing between duplicated *ASIP-ITCH* copies with crossover products showing reciprocal deletion and triplication. The 190-kb segments of copy 1 and copy 2 of a white Merino are shown as light and dark gray boxes, respectively. Black boxes indicate the similar SINE sequence regions at the junction, 5' and 3' breakpoints. In this example, the *ASIP* genotype shown represents a white animal heterozygous at copy 1 for the nonfunctional alleles (N₉D₅T/N₉N₅A) and homozygous at copy 2 for the functional alleles (N₉N₅T/N₉N₅T). Arrows below the *ASIP* genes indicate the direction of transcription driven by the *ITCH* promoter (*ITCH*^P). Nonallelic pairing and crossover between the junction and 3' breakpoints (dark dashed line) would result in creation of a single copy nonfunctional (N₉N₅A) allele, as shown. The resulting single *ASIP* copy is not expressed as the ancestral *ASIP* promoter (*ASIP*^P) is silent. A crossover point (gray cross) that would result in the positioning of a nonfunctional (N₉N₅A) allele under the regulation of the duplicated *ITCH* promoter is unlikely to occur as expression of "A" alleles was not detected in Merino sheep. Variable positioning of functional haplotypes at copy 1 and 2 combined with nonallelic pairing and crossover explains the different single copy alleles identified in black Merinos. Mutations and gene conversion events could also contribute to the diversity at the locus.

with DNase1 (Ambion DNA-free). RNA quality was visually assessed using agarose gel electrophoresis and a UV transilluminator and quantified by spectrophotometry.

ASIP PCR, cloning, and sequencing

PCR primers (Supplemental Table S2 online) for the amplification of intron sequences from genomic DNA templates were designed from sheep *ASIP* EST sequence. PCR primers were designed to amplify intron 1 regions between noncoding exon 1E and coding exon 2, intron 2 regions between exon 2 and exon 3, and intron 3 regions between exons 3 and 4. PCR reactions (20 μ L) contained 50 ng gDNA, 200 μ M dNTPs, 10 pmole of each primer, 1 \times Q solution, 1.5 mM MgCl₂, 0.5 units Taq polymerase (Qiagen), and 1 \times reaction buffer (Qiagen). PCR conditions were 3 min at 94°C; 30 sec at 94°C, 1 min at 57°C, and 2 min at 72°C for 35 cycles; and 5 min at 72°C. PCR products were amplified from a white Merino ram and a self-color black Merino lamb. PCR products were cloned using a TOPO TA Cloning Kit (Invitrogen) and sequenced using Big Dye Terminator 3.1 reaction mix. PCR products spanning exons 2, 3, and 4 were generated (see Supplemental Table S2 primer sequences) and sequenced directly to identify coding mutations in the DNA of selected animals from the panel of Merino coat color phenotypes. For direct sequencing, PCR products were processed with ExoSAP-IT (USB Corporation) according to the manufacturer's recommendations. All sequencing was done using ABI Prism Big Dye Terminator 3.1 chemistry and either an ABI 377 Prism DNA autosequencer or ABI 3130xl Genetic Analyser (Applied Biosystems). Sequence data were imported into Sequencher v4.2 (Gene Codes Corp.) for analysis. Sequences from the white and black Merino identified as Haplotypes 1 and 2 in Supplemental Table S1 have been deposited in GenBank (accession nos. EU420022 and EU420023).

Genotyping polymorphisms

The presence of D₉ and D₅ polymorphisms in exon 2 of the *ASIP* coding region was analyzed by PCR spanning both indels (see Supplemental Table S2 primer sequences) in a 20 μ L PCR reaction volume with Qiagen Taq performed according to the manufacturer's recommendations. Alleles were visualized by autoradiography and/or fluorescence on an ABI 377 Prism DNA autosequencer or ABI 3130xl Genetic Analyser (Applied Biosystems). A TaqMan MGB genotyping assay was developed for the g.5172T>A SNP in exon 4 of ovine *ASIP* (Assay by Design, Applied Biosystems). Genotypes of the panel of Merino coat color phenotypes were automatically assigned with a quality score of 90, using the Applied Biosystems 7900HT sequence detector

Characterization of *Ovis aries* BAC library clones

The three INRA Romanov Sheep BAC library clones (Vaiman et al. 1999) were obtained from the BAC-YAC Resource center of the Animal Genetics Department of INRA (http://dga.jouy.inra.fr/grafra/INRA_libraries_database_simplified.htm). Shotgun plasmid libraries were prepared by the Australian Genome Research Facility (AGRF; <http://www.agrf.org.au/>) from each of three INRA ovine BAC clones INRA-164H8, INRA-218G7, and INRA-229C6 DNA that had tested positive for *ASIP* coding sequence by PCR. Clones INRA-164H8 and INRA-218G7 were sequenced to 8 \times coverage and clone INRA-229C6 to 2 \times coverage by the AGRF (GenBank accession nos. EU185098–EU185100). Sequence data were imported into Sequencher v4.2 (Gene Codes) for analysis. Clones INRA-218G7 (~75 kb) and INRA-229C6 (~150 kb) partially shared the same sequence and were aligned to the longer clone

INRA-164H8 (~150 kb). Twenty-nine Texel Sheep BAC clones (Dalrymple et al. 2007) predicted to span the region within two Megabase pairs of *ASIP* were ordered from the Children's Hospital and Research Center at Oakland (CHRCO; <http://bacpac.chori.org/home.htm>). The 29 BAC clones were PCR analyzed for the presence or absence of both coding and noncoding regions of *ASIP*, *AHCY*, and *ITCH* genes. Primers used in this analysis are listed in Supplemental Table S2 online. BAC clone INRA-164H8 was used as the ovine reference sequence. BAC-end sequences from clones CH243-455O4 and CH243-489F15 that aligned to the reference sequence in a "head-to-head" orientation (with 5' ends closest) were predicted to span a junction between duplicated DNA copies. BAC clones with end sequences that aligned to the reference sequence in a "tail-to-tail" orientation (with 3' ends closest) were predicted to be fully contained within the reference sequence. Clone CH243-373J16 was predicted to provide sequence data further 5' and clone CH243-160L8 further 3' of the INRA-164H8 sequence. Clones CH243-234K21 and CH243-373J16 contained the putative nonfunctional "A" *ASIP* allele. These five clones of the 29 CHORI Texel sheep BAC clones—CH243-455O4, CH243-489F15, CH243-373J16, CH243-160L8, CH243-234K21—were selected for complete sequencing to 6 \times coverage (Macrogen; http://www.macrogen.com/eng/macrogen/macrogen_main2.jsp). Sequences were analyzed using Sequencher software v4.2 (GeneCodes). Sequences have been deposited in GenBank (GenBank accession nos. EU185093–EU185097).

Haplotypes phylogenetic analysis

The ovine *ASIP* haplotype sequences from this study were aligned and phylogenetic analyses performed using Molecular Evolutionary Genetics Analysis (MEGA) Software Version 4.0 (Tamura et al. 2007). The p-distance was used to estimate genetic distances and neighbor joining used to construct the phylogeny. Testing of inferred phylogeny was by bootstrap with 500 replications.

RT-PCR analysis

First-strand oligo(dT)₁₅ primed cDNA was generated using SuperScript III Reverse Transcriptase (Invitrogen) according to the manufacturer's instructions. PCR products were amplified, labeled with ³³P-dCTP, and separated by electrophoresis on a 5% polyacrylamide gel and analyzed by autoradiography or visualized with ethidium bromide staining following electrophoresis on a 1.0% agarose gel.

5' and 3' RACE

For 5' and 3' RACE experiments respectively, oligo(dT)₁₈ primed or UAP primed cDNA was prepared from 1–2 μ g of total RNA using SuperScript III Reverse Transcriptase (Invitrogen) according to the manufacturer's instructions. oligo(dT)₁₈ primed cDNA was poly A-tailed with dATP using Terminal Transferase (NEB) and excess primer and dNTPs removed using Qiaquick PCR Purification columns (Qiagen). 5' RACE products were amplified in touchdown PCR reactions (68°C –63°C) with primers GSP1 5' and UAP (Supplemental Table S2) and Qiagen Taq according to the manufacturer's instructions, followed by a nested PCR with primer GSP2 5' and UAP. Nested PCR was also used to amplify 3' RACE products. Primers GSP1 3' and UAP were used in the first PCR reaction followed by primers GSP2 3' and UAP in the second PCR reaction. PCR products were visualized by electrophoresis and ethidium bromide staining on 1.5% agarose gels. Products were gel purified and cloned for sequencing using TOPO TA Cloning (Invitrogen).

Junction point and 5' breakpoint assay

To estimate ovine *ASIP* copy numbers, an asymmetric competitive PCR protocol was developed following modification to the PCR method of Pielberg et al. (2003). Both the junction between duplicated DNA copies and the 5' breakpoint sequences were amplified in the one reaction tube. Primers Agt16 and Agt18 (Supplemental Table S2) spanning the junction between the duplicated copies produced a unique 242-bp product, while Agt16 and Agt17 spanning the 5' breakpoint sequence produced a 238-bp product (Supplemental Fig. S2). The products were amplified by asymmetric competitive PCR in a 5 μ L reaction volume in 384-well plates containing 5–50 ng gDNA, 500 μ M dNTP's, 3 mM MgCl₂, 1.25 \times reaction buffer (Qiagen), 5 pmole of specific forward primers (Agt17 and Agt18), 0.05 pmole of shared reverse primer (Agt16), and 0.15 units of HotStar Taq polymerase (Qiagen). PCR conditions were 15 min at 95°C; 30 sec at 95°C, 30 sec at 60°C, and 1 min at 72°C for 40 cycles; and 5 min at 72°C. A 1 μ L aliquot of a 1:10 dilution of PCR products was analyzed on an ABI 3130xl Genetic Analyser and the data analyzed using GeneMapper software (Liz 600 as size standard). The number of junction points (and *ASIP* copies) is estimated from the standard curve (Supplemental Fig. S3) using the ratio of the area under the 242-bp (junction point) peak compared with the total area represented by both peaks (junction point and breakpoint). To deduce discrete copy number categories for the data, peak areas were analyzed using the mean and standard deviation for each expected category (one, two, three, or four copies) and the normal statistical function.

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References

Adalsteinsson, S. 1970. Colour inheritance in Icelandic sheep and relation between colour, fertility and fertilisation. *J. Agric. Res. Iceland* **2**: 3–135.
 Adalsteinsson, S. 1975. Depressed fertility in Icelandic sheep caused by a single colour gene. *Ann. Genet. Sel. Anim.* **7**: 445–447.

Bennett, D.C. and Lamoreux, M.L. 2003. The colour loci of mice—A genetic century. *Pigment Cell Res.* **16**: 333–344.
 Bischof, J.M., Chiang, A.R., Scheetz, T.E., Stone, E.M., Casavant, T.L., Sheffield, V.C., and Braun, T.A. 2006. Genome-wide identification of pseudogenes capable of disease-causing gene conversion. *Hum. Mutat.* **27**: 545–552.
 Bonilla, C., Boxill, L.A., Donald, S.A., Williams, T., Sylvester, N., Parra, E.J., Dios, S., Norton, H.L., Shriver, M.D., and Kittles, R.A. 2005. The 8818G allele of the agouti signaling protein (*ASIP*) gene is ancestral and is associated with darker skin colour in African Americans. *Hum. Genet.* **116**: 402–406.
 Brooker, M.G. and Dolling, C.H.S. 1969. Pigmentation of sheep. II. The inheritance of coloured patterns in black Merino. *Aust. J. Agric. Res.* **20**: 387–394.
 Bultman, S.J., Michaud, E.J., and Woychik, R.P. 1992. Molecular characterization of the mouse agouti locus. *Cell* **71**: 1195–1204.
 Bultman, S.J., Klebig, M.L., Michaud, E.J., Sweet, H.O., Davison, M.T., and Woychik, R.P. 1994. Molecular analysis of reverse mutations from nonagouti (a) to black-and-tan (a') and white-bellied agouti (A^w) reveals alternative forms of agouti transcripts. *Genes & Dev.* **8**: 481–490.
 Carroll, L., Voisey, J., and van Daal, A. 2005. Gene polymorphisms and their effects in the melanocortin system. *Peptides* **26**: 1871–1885.
 Dalrymple, B.P., Kirkness, E.F., Nefedov, M., McWilliam, S., Ratnakumar, A., Barris, W., Zhao, S., Shetty, J., Maddox, J.F., O Grady, M., et al. 2007. Using comparative genomics to reorder the human genome sequence into a virtual sheep genome. *Genome Biol.* **8**: R152, doi: 10.1186/gb-2007-8-7-r152.
 Drogemuller, C., Giese, A., Martins-Wess, F., Wiedemann, S., Andersson, L., Brenig, B., Fries, R., and Leeb, T. 2006. The mutation causing the black-and-tan phenotype of Mangalitzta pigs maps to the porcine *ASIP* locus but does not affect its coding sequence. *Mamm. Genome* **17**: 58–66.
 Duhl, D.M., Vrieling, H., Miller, K.A., Wolff, G.L., and Barsh, G.S. 1994. Neomorphic agouti mutations in obese yellow mice. *Nat. Genet.* **8**: 59–65.
 Eizirik, E., Yuhki, N., Johnson, W.E., Menotti-raymond, M., Hannah, S.S., and O'Brien, S.J. 2003. Molecular genetics and evolution of melanism in the cat family. *Curr. Biol.* **13**: 448–453.
 Eppig, J.T., Bult, C.J., Kadin, J.A., Richardson, J.E., Blake, J.A., Anagnostopoulos, A., Baldarelli, R.M., Baya, M., Beal, J.S., Bello, S.M., et al. 2005. The Mouse Genome Database (MGD): From genes to mice—A community resource for mouse biology. *Nucleic Acids Res.* **33**: D471–D475.
 Girardot, M., Martin, J., Guibert, S., Leveziel, H., Julien, R., and Oulmouden, A. 2005. Widespread expression of the bovine Agouti gene results from at least three alternative promoters. *Pigment Cell Res.* **18**: 34–41.
 Girardot, M., Guibert, S., Laforet, M.P., Gallard, Y., Larroque, H., and Oulmouden, A. 2006. The insertion of a full-length *Bos taurus* line element is responsible for a transcriptional deregulation of the Normande Agouti gene. *Pigment Cell Res.* **19**: 346–355.
 Giuffra, E., Tornsten, A., Marklund, S., Bongcam-Rudloff, E., Chardon, P., Kijas, J.M., Anderson, S.I., Archibald, A.L., and Andersson, L. 2002. A large duplication associated with dominant white colour in pigs originated by homologous recombination between LINE elements flanking KIT. *Mamm. Genome* **13**: 569–577.
 Hayman, R.H. and Cooper, D.W. 1965. The frequency of pigmented sheep in the Australian Merino. *Wool Technol. Sheep Breeding* **12**: 81–85.
 Henry, B.A. 2003. Links between the appetite regulating systems and the neuroendocrine hypothalamus: Lessons from the sheep. *J. Neuroendocrinol.* **15**: 697–709.
 Kanetsky, P.A., Swoyer, J., Panossian, S., Holmes, R., Guerry, D., and Rebbeck, T.R. 2002. A polymorphism in the agouti signalling protein gene is associated with human pigmentation. *Am. J. Hum. Genet.* **70**: 770–775.
 Kerns, J.A., Newton, J., Berryere, T.G., Rubin, E.M., Cheng, J.F., Schmutz, S.M., and Barsh, G.S. 2004. Characterization of the dog Agouti gene and a nonagouti mutation in German Shepherd dogs. *Mamm. Genome* **15**: 798–808.
 Kohany, O., Gentles, A., Hankus, L., and Jurka, J. 2006. Annotation, submission and screening of repetitive elements in Repbase: RepbaseSubmitter and Censor. *BMC Bioinformatics* **7**: 474.
 Kuramoto, T., Nomoto, T., Sugimura, T., and Ushijima, T. 2001. Cloning of the rat Agouti gene and identification of the rat nonagouti mutation. *Mamm. Genome* **12**: 469–471.
 Kwon, H.Y., Bultman, S.J., Loffler, C., Chen, W.J., Furdon, P.J., Powell, J.G., Usala, A.L., Wilkison, W., Hansmann, I., and Woychik, R.P. 1994. Molecular structure and chromosomal mapping of the human homolog of the Agouti gene. *Proc. Natl. Acad. Sci.* **91**: 9760–9764.

- Leeb, T., Deppe, A., Kriegsmann, B., and Brenig, B. 2000. Genomic structure and nucleotide polymorphisms of the porcine Agouti signalling protein gene (ASIP). *Anim. Genet.* **31**: 335–336.
- Lindsay, S.J., Khajavi, M., Lupski, J.R., and Hurler, M.E. 2006. A chromosomal rearrangement hotspot can be identified from population genetic variation and is coincident with a hotspot for allelic recombination. *Am. J. Hum. Genet.* **79**: 890–902.
- Marklund, S., Kijas, J., Rodriguez-Martinez, H., Ronnstrand, L., Funari, K., Moller, M., Lange, D., Edfors-Lilja, I., and Andersson, L. 1998. Molecular basis for the dominant white phenotype in the domestic pig. *Genome Res.* **8**: 826–833.
- McCann, J.P., Bergman, E.N., and Beermann, D.H. 1991. Dynamic and static phases of severe dietary obesity in sheep: Food intakes, endocrinology and carcass and organ chemical composition. *J. Nutr.* **122**: 496–505.
- Mundy, N.I. and Kelly, J. 2006. Investigation of the role of the Agouti signalling protein gene (ASIP) in coat colour evolution in primates. *Mamm. Genome* **17**: 1205–1213.
- Myers, S.R. and McCarroll, S.A. 2006. New insights into the biological basis of genomic disorders. *Nat. Genet.* **38**: 1363–1364.
- Nakayama, K. and Ishida, T. 2006. Alu-mediated 100-kb deletion in the primate genome: the loss of the Agouti signalling protein gene in the lesser apes. *Genome Res.* **16**: 485–490.
- Parsons, Y.M., Fleet, M.R., and Cooper, D.W. 1997. Investigation of the gene responsible for recessive pigmentation in Australian Merino sheep. *Proc. Assoc. Advanc. Anim. Breed. Genet.* **12**: 447–451.
- Parsons, Y.M., Fleet, M.R., and Cooper, D.W. 1999a. The Agouti gene: A positional candidate for recessive self-colour black pigmentation in Australian Merino sheep. *Aust. J. Agric. Res.* **50**: 1099–1103.
- Parsons, Y.M., Fleet, M.R., and Cooper, D.W. 1999b. Isolation of the ovine Agouti coding sequence. *Pigment Cell Res.* **12**: 394–397.
- Perry, W.L., Hustad, C.M., Swing, D.A., O'Sullivan, T.N., Jenkins, N.A., and Copeland, N.G. 1998. The itchy locus encodes a novel ubiquitin protein ligase that is disrupted in a^{18H} mice. *Nat. Genet.* **18**: 143–146.
- Pielberg, G., Day, A.E., Plastow, G.S., and Andersson, L. 2003. A sensitive method for detecting variation in copy numbers of duplicated genes. *Genome Res.* **13**: 2171–2177.
- Rendel, J. 1957. Heredity of colour and white designs in farm animals. *Kungliga Hogskole Landbruksadami Tidsskrift* **96**: 207–263.
- Rieder, S., Taourit, S., Mariat, D., Langlois, B., and Guerin, G. 2001. Mutations in the agouti (ASIP), the extension (MC1R), and the brown (TYRP1) loci and their association to coat colour phenotypes in horses (*Equus caballus*). *Mamm. Genome* **12**: 450–455.
- Smit, M.A., Shay, T.L., Beever, J.E., Notter, D.R., and Cockett, N.E. 2002. Identification of an agouti-like locus in sheep. *Anim. Genet.* **33**: 383–385.
- Sponenberg, D.P. 1997. Genetics of colour and hair texture. In *The genetics of sheep* (eds. L.R. Piper and A. Ruvinsky), pp. 51–86. New York, NY.
- Tamura, K., Dudley, J., Nei, M., and Kumar, S. 2007. MEGA4: Molecular evolutionary genetics analysis (MEGA) software version 4.0. *Mol. Biol. Evol.* **24**: 1596–1599.
- Vage, D.I., Lu, D., Klungland, H., Lien, S., Adalsteinsson, S., and Cone, R.D. 1997. A non-epistatic interaction of agouti and extension in the fox, *Vulpes vulpes*. *Nat. Genet.* **15**: 311–315.
- Vaiman, D., Billault, A., Tabet-Aoul, K., Schibler, L., Vilette, D., Oustry-Vaiman, A., Soravito, C., and Cribiu, E.P. 1999. Construction and characterization of a sheep BAC library of three genome equivalents. *Mamm. Genome* **10**: 585–587.
- Voisey, J. and van Daal, A. 2002. Agouti: From mouse to man, from skin to fat. *Pigment Cell Res.* **15**: 10–18.
- Voisey, J., Imbeault, P., Hutley, L., Prins, J.B., and van Daal, A. 2002. Body mass index-related human adipocyte agouti expression is sex-specific but not depot-specific. *Obes. Res.* **10**: 447–452.
- Voisey, J., Gomez-Cabrera, M.D.C., Smit, D.J., Leonard, J.H., Sturm, R.A., and van Daal, A. 2006. A polymorphism in the Agouti signalling protein (ASIP) is associated with decreased levels of mRNA. *Pigment Cell Res.* **19**: 226–231.
- Vrieling, H., Duhl, D.M., Millar, S.E., Miller, K.A., and Barsh, G.S. 1994. Differences in dorsal and ventral pigmentation result from regional expression of the mouse Agouti gene. *Proc. Natl. Acad. Sci.* **91**: 5667–5671.
- Wolff, G.L. 2003. Regulation of yellow pigment formation in mice: A historical perspective. *Pigment Cell Res.* **16**: 2–15.
- Wolff, G.L., Roberts, D.W., and Mountjoy, K.G. 1999. Physiological consequences of ectopic agouti gene expression: The yellow obese mouse syndrome. *Physiol. Genomics* **1**: 151–163.

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